



Article scientifique

Article

2020

Accepted version

Open Access

This is an author manuscript post-peer-reviewing (accepted version) of the original publication. The layout of the published version may differ .

---

## Diagnosis and management of hypersensitivity reactions to vaccines

---

Sarti, Lucrezia; Lezmi, Guillaume; Mori, Francesca; Giovannini, Mattia; Caubet, Jean-Christoph Roger J-P

### How to cite

SARTI, Lucrezia et al. Diagnosis and management of hypersensitivity reactions to vaccines. In: Expert review of clinical immunology, 2020, vol. 16, n° 9, p. 883–896. doi: 10.1080/1744666X.2020.1814745

This publication URL: <https://archive-ouverte.unige.ch//unige:159480>

Publication DOI: [10.1080/1744666X.2020.1814745](https://doi.org/10.1080/1744666X.2020.1814745)



## Diagnosis and management of hypersensitivity reactions to vaccines

Lucrezia Sarti , Guillaume Lezmi , Francesca Mori , Mattia Giovannini & Jean-Christoph Caubet

To cite this article: Lucrezia Sarti , Guillaume Lezmi , Francesca Mori , Mattia Giovannini & Jean-Christoph Caubet (2020): Diagnosis and management of hypersensitivity reactions to vaccines, Expert Review of Clinical Immunology, DOI: [10.1080/1744666X.2020.1814745](https://doi.org/10.1080/1744666X.2020.1814745)

To link to this article: <https://doi.org/10.1080/1744666X.2020.1814745>



Accepted author version posted online: 24 Aug 2020.



Submit your article to this journal [↗](#)



View related articles [↗](#)



View Crossmark data [↗](#)

**Publisher:** Taylor & Francis & Informa UK Limited, trading as Taylor & Francis Group

**Journal:** *Expert Review of Clinical Immunology*

**DOI:** 10.1080/1744666X.2020.1814745

**Diagnosis and management of hypersensitivity reactions to vaccines**

**Lucrezia Sarti<sup>1</sup>, Guillaume Lezmi<sup>2,3</sup>, Francesca Mori<sup>1</sup>, Mattia Giovannini<sup>1</sup>, Jean-Christoph Caubet<sup>4</sup>**

1. Allergy Unit, Department of Pediatrics, Anna Meyer Children's University Hospital, Florence, Italy
2. Service de Pneumologie et Allergologie Pédiatriques, Hôpital Necker-Enfants Malades, AP-HP, Paris, France.
3. Université Paris Descartes, Paris, France
4. Division of Pediatric Allergy, Department of Pediatrics, University Hospitals of Geneva, Geneva, Switzerland

### **Corresponding Author**

Lucrezia Sarti,

Address: AOU Meyer Viale Pieraccini 24, 50139, Florence, Italy

E-mail: [lucrezia.sarti@gmail.com](mailto:lucrezia.sarti@gmail.com)

Telephone: +39-05556621

## **Diagnosis and management of hypersensitivity reactions to vaccines**

Lucrezia Sarti<sup>1\*</sup>, Guillaume Lezmi<sup>2,3</sup>, Francesca Mori<sup>1</sup>, Mattia Giovannini<sup>1</sup>, Jean-Christoph Caubet<sup>4</sup>

<sup>1</sup>Allergy Unit, Department of Pediatrics, Anna Meyer Children's University Hospital, Florence, Italy

<sup>2</sup>Service de Pneumologie et Allergologie Pédiatriques, Hôpital Necker-Enfants Malades, AP-HP, Paris, France.

<sup>3</sup>Université Paris Descartes, Paris, France

<sup>4</sup>Division of Pediatric Allergy, Department of Pediatrics, University Hospitals of Geneva, Geneva, Switzerland

### **\*Corresponding Author:**

Lucrezia Sarti, AOU Meyer Viale Pieraccini 24, 50139, Florence, Italy

Telephone: +39-05556621

E-mail: lucrezia.sarti@gmail.com

### **Abstract**

**Introduction:** Many countries in Europe now recommend and enforce mandatory vaccinations to improve vaccination coverage. Thus, the number of adverse events following immunization (AEFI) may show an increase. Among these events, severe hypersensitivity reactions to vaccines are rare. However, it is important that they be identified and recognized so that they may be adequately managed.

**Areas covered:** The literature search was undertaken through PubMed and Embase to identify English-language papers focusing on hypersensitivity to vaccines.

**Expert opinion:** Hypersensitivity reactions following vaccinations are rare and are classified according to their chronology and extension: immediate when they occur within the first 4 hours following administration and non-immediate when they occur later. Local reactions are the most common adverse event following injection of vaccines and generally do not require any allergy workup. Immediate reactions, however, are potentially IgE-mediated and require an allergy workup. In general, a previously known food allergy (i.e. egg or milk) is not a contraindication to immunizations. Patients with a known allergy to gelatin, yeast, latex, antibiotics or other specific components of vaccines require an allergy workup before administration of the vaccine.

**Key words:** vaccine, hypersensitivity reactions, vaccine allergy, systemic reactions, local reactions, hypersensitivity, egg allergy, gelatin.

## Article highlights

- Local reactions to the vaccine are the most frequent adverse events following vaccinations. They are benign, not at risk of anaphylaxis, and generally require no allergy workup. Further vaccines can be administered safely without precaution.
- All immediate reactions after vaccine administration should be assessed by an allergist.
- Immediate reactions (<4 hours) are potentially IgE-mediated and require an allergy workup to prevent the occurrence of anaphylaxis after further administration.
- Egg allergy is not a contraindication to influenza vaccine. In the case of previous anaphylaxis to egg, some guidelines recommend administering the vaccine without specific precaution, while others recommend that an experienced staff administers the vaccine. In the absence of a prior history of anaphylaxis after egg consumption, influenza vaccines can be administered without precaution in egg-allergic patients.
- Allergy to gelatin, yeast, latex and antibiotics or other specific components of vaccines require an allergy workup before administration of the vaccine.

ACCEPTED MANUSCRIPT

## 1. Introduction

Vaccines are a cornerstone of pediatric healthcare. The introduction of immunizations for the prevention of life-threatening infections was an important driver of improvements in infant and childhood morbidity and mortality in the 20th century. For this reason, nowadays in the vast majority of developed countries, vaccines on the National Immunization Program are free of charge for children, adults or both and are given in local council immunization sessions, primary healthcare provider clinics and some public hospitals. Enforcing mandatory vaccinations or strongly recommending vaccinations is one of the strategies that some countries in Europe have adopted to protect the community when vaccination coverage was not satisfactory. A recent study [1] showed that 35.4% of European countries had policies of mandatory vaccinations for at least one vaccine.

The success of immunization programs in eliminating vaccine-preventable diseases depends on the community knowledge and acceptance of the balance between the benefits of immunization and the potential vaccine risks. Parents are the ones who commonly perceive that their child has experienced an adverse event following immunization (AEFI), and within this group, the subsequent expectation of an AEFI and vaccine safety concerns may be heightened [2]. In this context, the allergist has a key role in identifying the potential reactions to investigate, in order to give patients a practical answer to their concerns.

The literature search was undertaken through PubMed and Embase. English-language papers focusing on hypersensitivity to vaccines were identified, using the keywords “vaccine hypersensitivity”.

## 2. Definition

An adverse drug reaction (ADR) is defined as a harmful and unintended effect occurring at doses normally used in humans for the prevention, diagnosis and treatment of diseases or, in general, for the modification of a physiological function [3]. An allergic reaction is defined as a harmful idiosyncratic response produced by a specific immune mechanism [4]. When drug reactions resembling allergy occur, they are called drug hypersensitivity reactions (DHRs) before showing the evidence of either drug-specific antibodies or T cells mediation. DHRs may be allergic or nonallergic in nature. These reactions are typically unpredictable [5]. For general communication, when an allergic drug reaction is suspected, DHR is the preferred term, because it may be difficult to differentiate between true drug allergy and nonallergic DHR based on the clinical presentation alone, especially in cases of acute severe DHR [5].

The definition of the type of reaction after immunizations is vitally important and often challenging. An AEFI includes any untoward medical occurrence following immunization, which does not necessarily have a causal relationship with the administration of the vaccine. Reported adverse events

can either be true adverse events, or coincidental events that are not due to the vaccine or immunization process but are temporally associated with immunization [6].

Any type of vaccine can cause an allergic reaction; however, in many cases, a suspected allergy to a vaccine is not conclusively confirmed [7-8]. Allergic reactions to vaccines have been reported with an incidence ranging from 1 case per 50.000 doses to 1 per 1.000.000 doses [9]. Hypersensitivity can occur as a result of the immunizing antigen or most often one of the other components of the vaccine (suspension fluid, preservatives, stabilizers, antibiotics and adjuvants).

### **3. Classification of the adverse reactions to vaccines**

Immune reactions to drugs and vaccines, can be grouped into four types according to the Gell and Coombs classification [10]: type I or immediate reactions; type II or cytotoxic reactions; type III or reactions mediated by immune complexes; and type IV or delayed hypersensitivity reactions. Hypersensitivity reactions to vaccines are commonly classified by their clinical extension (local or systemic reaction) or according to the timing of the symptoms (immediate and or delayed) [11].

Distinguishing the type of reaction to a vaccine based on time of onset of symptoms and with different organ involvement is essential to prevent a re-exposure to a vaccine that can precipitate systemic and immediate reactions, which could potentially be life-threatening.

Local reactions or injection site reactions are the most frequent adverse events following immunization and have an important impact on clinical practice. Indeed, patients that manifest these reactions are often falsely labeled as allergic [6, 11-14]. Systemic reactions range from fever, headache, myalgia, generalized urticaria to anaphylaxis, that can affect potentially two or more systems: skin (i.e. erythema, pruritus, urticaria, angioedema, maculopapular rash), respiratory tract (i.e. stridor, wheezing, dyspnea), gastrointestinal (i.e. vomiting, diarrhea, abdominal pain) and cardiovascular systems (i.e. weakness, syncope, palpitations, tachycardia and hypotension) [4]. These reactions are less common, but their adequate identification and management are crucial because they include anaphylaxis [11].

Immediate reactions to vaccine are defined as occurring within minutes of exposure to the allergen and generally within 4 hours [9]; however, it is rare for an anaphylaxis reaction to occur beyond the first hour. Immediate reactions include injection site reactions and, rarely, systemic reactions.

Delayed reactions to vaccines are defined as occurring within hours or days after exposure [4,6]. Most delayed reactions are limited and do not contraindicate the administration of future doses of the same vaccine [6]. Delayed reactions include [4]: cytotoxic reactions (type II), i.e. thrombocytopenia after administration of the measles-rubella vaccine [15-17]; reactions mediated by immune complexes (type III), i.e. serum sickness [18-20], Arthus reaction [21-22], erythema nodosum [23-24] or Henoch-



Schönlein purpura [25]; cellular reactions or delayed hypersensitivity reactions (type IV), i.e. contact dermatitis and subcutaneous nodules. Delayed urticaria and/or angioedema, or maculopapular rashes, are relatively common symptoms that can occur after vaccinations [11,28]. The pathogenesis of these reactions is not fully understood; however, the role of basophils' activation [26] and a reaction to circulating immune complexes [27] has been proposed in cases of reaction to Hepatitis B vaccine [11, 28].

### **3.1 Injection site reactions to vaccines**

Injection site reactions are the most frequent adverse reactions following immunization [11,13,14]. Injection site reactions include two major patterns: a) pain, redness, and/or swelling and b) persistent subcutaneous itchy nodules at the injection site [11-29]. Their frequency depends on the composition of the vaccine, the number of injections previously administered, and the immunological and inflammatory responses of the host [30,31]. Injection site reactions are particularly frequent with acellular Pertussis (aP)-containing vaccines [32, 33]. Other reactions, such as sterile abscesses, morphea and nevi, with or without hypertrichosis are anecdotal [11].

Pain, redness, and/or swelling at the injection site are the most common local reactions and are generally mild. They are observed in 23 to 81% and 44 to 84% of infants and toddlers following vaccinations with 7 or 13 valent-pneumococcal conjugate vaccines, respectively [30], and in more than 75% of children between four and six years of age following a booster vaccination for diphtheria-tetanus-pertussis-poliomyelitis [34]. These reactions could result from non-specific inflammation induced by microbial or other components used as adjuvants [14]. Large injection site reactions typically occur within 24 to 72 hours following immunization and disappear in two to three days [11,14]. Reactions extending beyond the nearest joint or lasting more than three days are sometimes defined as severe local reactions [13]. Large injection site reactions most frequently occur after injections of toxoid-containing vaccines but may be observed after the injection of any vaccine [11, 13]. They may result from toxoid or aluminum hydroxide-induced inflammation and may occur after any injections of a vaccine [11,35]. Large injection site reactions may also result from an Arthus reaction in previously immunized patients who have developed high titers of specific IgG against the microbial components of the vaccine [11]. In this case, IgG antibodies may bind to vaccine antigens at the injection site and form antigen/antibody complexes, which are thought to activate complement, leading to non-specific mast-cell degranulation and neutrophil recruitment. Arthus reactions develop only in previously immunized patients and typically occur after the fourth or fifth injection [11].

Extensive limb swelling (ELS) is generally characterized as extending beyond the elbow or knee [11]. ELS can occur at any age after administration of a wide variety of vaccines, especially after polyvalent pneumococcal vaccine, diphtheria, tetanus toxoids and aP-containing vaccines [36]. ELS is

defined as swelling that measures at least 10 cm and it was observed in 1.3% of children following the fourth dose of aP-containing vaccines [37]. ELS occurs commonly within the first 24 hours after vaccination [36], and is usually painless [11, 37, 38]. Ultrasound examination of 12 children with ELS suggested the potential implication of extravasation mechanisms [39].

Injection site reactions, both mild and large, are benign, resolve spontaneously, and most patients with previous large injection site reactions tolerate subsequent vaccine doses [40]. No allergy workup is generally required, and injection site reactions should not delay subsequent vaccination [9,14]. However, high titers of specific IgG to the vaccine in patients with large injection site reactions are strongly suggestive of an Arthus reaction. In this case, future administration may be delayed as long as IgG titers are protective [11]. In children up to six years of age, injection site reactions may be less frequent but more pronounced if the vaccine is injected in the thigh rather than the arm [41,42].

Persistent (> 6 months) subcutaneous itchy nodules are observed in approximately 1% of children following injection of vaccines containing aluminum, such as diphtheria-tetanus-pertussis-polio-hemophilus influenza type b, pneumococcal, or meningococcal conjugate vaccines [43, 44]. They typically develop weeks after injection [44]. They may increase and become itchier during infections and are often associated with local hypertrichosis and eczema [45]. Patch tests for aluminum salts are positive in 77 to 95% of patients, suggesting delayed type IV hypersensitivity to aluminum [43, 45-49]. However, 8% of control subjects without persistent nodules also have positive patch tests for aluminum salts [44]. The nodule may persist for several years before disappearing. Positivity for the patch tests often disappears over time, suggesting a loss of hypersensitivity [50]. Persistent subcutaneous itchy nodules are benign but may lead to unnecessary investigations and postponement of further vaccination [45, 47]. In clinical practice, persistent subcutaneous itchy nodules do not require any investigation and do not contraindicate vaccination.

No allergy workup is needed for most injection site reactions. These reactions have not been associated with subsequent anaphylactic reactions. Determination of specific IgG concentrations in large injection site reactions may be useful. Prevention of relapses is based on intramuscular injection of the vaccines [45, 51, 52].

### **3.2 Systemic reactions to vaccines**

Very rarely do vaccines cause immediate hypersensitivity reactions, and among these, severe systemic reactions are even less frequent.

Anaphylaxis, the most severe form of acute IgE-mediated reactions, can involve multiple organ systems and can present with variable severity. The rate of anaphylaxis to vaccines has been estimated to be approximately 1 per million vaccine doses [9,53]. Current data are limited to estimating the risk of anaphylaxis associated with vaccination. The majority of studies that estimate the rate of

anaphylaxis to vaccines used passive surveillance systems that lacked an unvaccinated comparison group [9,54]. Passive surveillance systems depend on voluntary reports and cases of anaphylaxis are generally identified based on the presence of suggestive symptoms (Brighton criteria)[57] without confirmation of causality. In addition, several vaccines are often administered together. For these reasons, the exact rate of anaphylaxis for each vaccine is difficult to estimate.

In a recent study, Su et al. [55] searched the VAERS database (Vaccine Adverse Event Reporting System) [56] for reports of anaphylaxis after vaccination in the US for a period of 27 years and focused their analysis on 828 reports that met the Brighton Collaboration case definition for anaphylaxis [57] or included a diagnosis of anaphylaxis by a physician and in addition described symptoms within one day of receiving the vaccine. The most vaccine-induced anaphylaxis in children and young adults (< 19 age) were found to be MMR, Varicella and DTaP/Tdap vaccines, while the influenza vaccine was most commonly reported for adults [55]. The authors estimated a rate of anaphylaxis during the 27 year period to be 0.6 per million doses distributed of MMR and 0.2 per million doses distributed of PCV23. In a shorter test period of 10 years, the estimated rate of anaphylaxis was 1.2 per million doses distributed of Varicella vaccines and in a further test period of 6 years the median estimated annual rate of anaphylaxis due to influenza vaccines was 0.2 per million doses distributed. The authors suggested that the low rate of anaphylaxis in respect to previous studies was due to the fact that VAERS does not collect data on doses administered but estimates rates based on doses distributed which consequently creates a large denominator [55].

McNeil et al. [58], using Vaccine Safety Datalink (VSD) to enroll patients in a three year period (2009-2011), estimated that the rate of anaphylaxis was 1.31 (95% CI, 0,90-1,84) per million vaccine doses administered [58,59]. The advantage of using VSD is that the sites maintain a linked database of health care encounters, including immunization registers with detailed information on vaccines administered.

In both studies [55,58] the most frequently implicated vaccine in anaphylactic reactions was influenza. However, this might reflect its greater frequency of administration. The two studies mentioned above [55,58] are in agreement about the demographic characteristic of patients with anaphylaxis. In particular they found that the anaphylaxis reaction to vaccines in the adult population is more frequent in females and that the median age of patients who experienced anaphylaxis was similar: 12 years in Su's study [55] and 17 years in McNeil's study [58]. Finally, in both studies, atopy was present in the clinical history of patients with anaphylaxis reaction to vaccines: 59% in Su et al. [55] and 85% in McNeil et al. [58].

In the management of patients with suspected anaphylaxis to vaccines, it is important to remember that there are many immediate and systemic adverse events that could be misdiagnosed as anaphylaxis and many of these occur more frequently than vaccine related anaphylaxis [9]. For example, vasovagal syncope and hypotonic hyporesponsive episodes following immunization may be confused with anaphylaxis [9].

There are other systemic reactions to vaccines that are more frequent than anaphylaxis, including fever, headache, myalgia delayed or immediate skin symptoms (i.e. urticaria and/or angioedema, or maculopapular or other nonspecific rashes) and respiratory symptoms (i.e. rhinitis, wheezing). Other systemic reactions are extremely rare, such as Guillain-Barré Syndrome, Immune Thrombocytopenic Purpura, vasculitis and Serum Sickness. These reactions are not discussed in this review. The “Institute for vaccine safety, Johns Hopkins Bloomberg School of public health” provides an updated discussion and revision of the literature about these rare systemic reactions to vaccine [60].

### **3.2.1 Diagnostic workup (Figure 1)**

In the case of suspected IgE-mediated reaction (i.e. urticarial rashes or anaphylaxis occurring within 4 hours from vaccine administration) where further doses of vaccine are required, a complete allergy workup is mandatory in order to avoid future reactions with the same vaccine or the possibility of cross-reactivity with components of other vaccines or foods. Skin tests (Skin Prick tests and Intradermal Tests) are recommended at least 3 weeks, but no more than one year, after the suspected IgE-mediated reaction [61], although ideally, they should be conducted within six months. In case of positive skin tests, the diagnosis of allergy is confirmed. However, positive and negative predictive values of skin tests to vaccines have not yet been established.

From a practical point of view, the occurrence of immunization could be assessed through the evaluation of disease-protecting antibody titers [12]. In case of a confirmed protective immunity induced by the first dose of vaccination, further vaccine doses could be delayed, always being aware that the duration of protection may be shorter than that of a standard administration schedule.

### **3.2.2 Scheme of vaccination of patients with immediate systemic reactions.**

In the case of non-anaphylactic immediate systemic reactions, if the allergy workup (skin tests and specific IgE) is negative, the vaccine can be administered as usual and the patients observed for at least 30 min [4].

In the presence of severe immediate and systemic reactions (suggestive of being IgE-mediated), if the allergy work-up is negative, the vaccine should be administered in two doses: 1/10 of the total amount followed 30-60 minutes later by the remaining dose with a subsequent observation period of at least 30 minutes and even better if observed for one hour [14, 63]. So far, there have been no reports of patients with negative ID testing with the vaccine followed by a serious anaphylactic reaction upon revaccination.

If the skin tests or serum specific IgE titers are found to be positive in a patient with a history consistent with IgE-mediated reactivity to one of the components of the vaccine, it is advisable to use a vaccine lacking that component.

In any cases of a positive allergy workup in patients with suggestive history of severe immediate and systemic reaction to a vaccine or its component and vaccination is considered essential, the suspect vaccine or another vaccine containing the suspect component should be administered with a graded desensitization protocol. The scheme most commonly used is the one proposed by the American Academy of Pediatrics [14, 64]:

1. 0.05 ml of the 1:10 dilution in physiological saline solution
2. 0.05 ml of full-strength vaccine
3. 0.10 ml of full-strength vaccine
4. 0.15 ml of full-strength vaccine
5. 0.20 ml of full-strength vaccine
6. For vaccines requiring a volume of 1 ml, we can add a last dose of 0.5 ml

Each dose is administered every 15 minutes and at the end of the procedure the patient remains under observation for at least 30-60 minutes. This procedure is performed in patients considered at risk of severe reactions because they have been diagnosed as “allergic” to a vaccine or its components. For this reason, the desensitization or fractionated doses administration needs to be performed by trained personnel in a hospital setting with lifesaving facilities available.

The approach to the investigation and subsequent revaccination of patients who reacted after administration of multiple or combined vaccines is more time-consuming. Indeed patients need to be skin tested for all the suspected vaccines in a single session and, if the clinical history of the reaction is suggestive of an IgE mediated reaction and the skin tests are inconclusive, all the individual vaccines should be administered separately in different sessions.

In regard to the management of patients with risk factors, it is important to emphasize that for patients with mastocytosis it is recommended that vaccinations are performed with single vaccines and that observation time is 30 minutes at least, but a controlled setting is not usually required [14]. The management of patients with possible allergies to any component of vaccines as a risk factor is discussed in the specific section below.

#### **4. Hypersensitivity reactions to vaccine components**

Vaccine antigens are rarely the cause of hypersensitivity reaction in the vaccinated individual. They have been reported, however, particularly with tetanus toxoids [65], pneumococcal antigens [66] and hepatitis B [26-27]. In a recent study CRM (197) the non-toxic mutant form of diphtheria toxin has been identified as an allergen that can elicit anaphylaxis reaction in patients immunized with PCV 13 [67].

Many other vaccine components have been hypothesized as being possibly responsible for hypersensitivity reactions to vaccines, but the reactions involving other vaccine components are more frequent than one involving the microbial component [6,12]. However, for some of them the direct causality between the component and the reaction has not been demonstrated, or it is not clear. In these cases, the management of the patient must be evaluated on a case-by-case basis, always remembering that most patients who develop a delayed reaction can receive the vaccine with a low risk of a mild reaction which is usually outweighed by the benefit of the vaccination [11].

Vaccine components that are known to cause hypersensitivity reactions are reported in Table 1, 2.

It is important to note that in the table some components of vaccines which have previously been described as responsible for delayed and injection site reactions (phenol, formaldehyde and 2-phenoxyethanol) are not listed, because they are mentioned only in outdated and single-case reports, without any recently confirmed data [120-122]. In only one recent study of Nagao et al. [123] the potential implication of 2-phenoxyethanol in anaphylaxis following influenza vaccine was suggested, although not confirmed.

#### **4.1 Potential allergens in vaccines**

##### **4.1.1 Egg proteins**

Literature underlines the rarity of severe reactions following vaccines potentially contaminated with egg proteins [79,82, 124, 125]. Recent data [4,6, 8,9,11-14, 61, 126, 127] confirm that no precaution is necessary for egg-allergic children who must undergo MMR or MMRV, even in those with a history of anaphylaxis to egg, because the safety of these vaccines depends on the minimum amounts of ovalbumin which is the agent that potentially contaminates the vaccines (0-1 ng / ml) [125].

However, an allergist should evaluate those children who have experienced a reaction with a previous MMR/MMRV vaccine, in order to exclude the possibility that the reaction occurred as a result of an hypersensitivity to some other components of the vaccine (especially gelatin). The same recommendation can be given for the tick born encephalitis vaccine. This vaccine is also grown on chicken embryo fibroblast and therefore contains low amounts of ovalbumin (< 1ng/ml) [14].

Some precautions are required in egg allergic patients who must be subjected to YF vaccination. Considering the number of children with egg allergy that undergo the administration of YF vaccine compared to the other vaccines (i.e. anti-influenza vaccination and anti MMR/MMRV), data on its safety in these patients are still lacking [9]. The concentrations of ovalbumin in YF vaccines are higher than in MMR/MMRV or influenza vaccines [14], ranging from 0,13 to 4,42 ug/ml, depending on the study and product batches [80,128]. Therefore, in egg allergic patients, skin testing including a skin prick test and, if negative, an intradermal test is recommended before the administration of YF

vaccine [9,11]. If skin testing is positive, the vaccine must be administered in graded doses under hospital observation. If the tests are negative, vaccination can be carried out as usual [9].

For the management of patients with allergies to egg and anti-influenza vaccinations, see specific sections below.

#### **4.1.2 Milk**

Hypersensitivity reactions possibly related to the presence of milk derivatives were described for MMR [117] and more recently for OPV [114] and DTaP or Tdap vaccines [113]; although these studies are debated and have not been confirmed.

There is a general consensus in literature to remark that no precautions are required when administering these vaccines to milk-allergic patients, even in those with history of anaphylaxis to milk. However, if a patient known to be allergic to milk suffers an allergic reaction to one of these vaccines, the possibility of milk protein contaminating should be considered [4,6, 9, 14]. It is noted that milk proteins are not included in the table of vaccine allergens from the “Institute for vaccine safety, Johns Hopkins Bloomberg School of public health” updated on December 2018 [68].

#### **4.1.3 Gelatin**

Gelatin, an animal protein used widely in foods and medication as a stabilizer in vaccines, was previously recognized as the principal cause of hypersensitivity reaction to MMR/MMRV vaccines and to tick-borne encephalitis vaccine [11,14].

In particular, a hypersensitivity reaction to vaccines is attributed to porcine or bovine gelatin, in that they show important cross-reactivity. The exact mechanism for patients to become sensitized to gelatin is unknown. However, recent studies have proposed galactose- $\alpha$ -1,3-galactose (alpha-gal), an allergen involved in hypersensitivity reactions to red meat and after exposure to tick bites, as a potential cross-reactive allergen responsible for hypersensitivity reactions to gelatin contained in vaccines [92,97,103,130]. Another possible cross-reactive allergen proposed in a recent study was bovine serum albumin, a major allergen (Bos d 6) in beef and a minor allergen in cows' milk [131]. Finally, Bogdanic J et al. showed that 16% and 38% respectively of beef and pork meat sensitized children, have IgE antibodies to gelatins that are cross-reactive [132].

It should be noted that in some countries, such as Japan and Germany, vaccine manufacturers have removed gelatin from vaccines or changed to a less allergenic gelatin (thoroughly hydrolyzed), with a resultant decrease in allergic reactions [133-137].

Thus, in patients allergic to gelatin, a gelatin-free vaccine should be preferred, because the content of gelatin in vaccine is not negligible (ranging from 500ug/0,5 ml to 12 mg/0,5 ml). If a gelatin-free vaccine is unavailable and the vaccination is required, a skin test with the vaccine itself should be performed before vaccine administration. Patients with negative skin tests can receive the full

vaccine dose, whereas patients with positive skin tests should receive the vaccine in fractionated doses [11, 12, 14].

#### **4.1.4 Yeast protein**

While yeast protein is present in HepB and HPV vaccines, only a few studies have demonstrated a possible relationship between the hypersensitivity reaction after immunization and the rare cases of yeast allergy, especially in HepB vaccines [110- 111]. Because the amount of yeast protein can reach 25 mg per dose (in HepB vaccine) [138] and because of the limited data present in literature, patients with suspected or confirmed yeast allergies should undergo a preliminary allergic evaluation with a skin prick test or serum specific IgE with *S. cerevisiae*. If the tests are negative, vaccination can be performed as usual, ~~instead~~ however, if they are positive a skin test with the vaccine needs to be performed. If positive, vaccine administration can proceed in fractionated doses [4,9]. It should be noted that the amount of yeast protein in the quadrivalent HPV vaccine is less than 7 ug/dose [138].

#### **4.1.5 Natural rubber latex**

Natural rubber latex (NRL) can be present in the rubber stopper of some vaccine vials and plungers in some prefilled syringes. Even if it has been rarely reported as responsible for hypersensitivity reaction [118,119], it is a potential cause of anaphylactic reaction in NRL allergic patients. For this reason, it should be suggested that patients with a confirmed allergy to NRL be vaccinated with caution with latex-free equipment, such as gloves [4]. In case of a hypersensitivity reaction occurring in a patient immunized with a vaccine that contains latex in its packaging, latex allergy should be excluded. It is worth noting that if clinical manifestations of the patients are indicative of contact allergy, immunization can be performed without precaution [4].

#### **4.1.6 Antibiotics**

Some antibiotics, such as neomycin, gentamycin, streptomycin and polymyxin B, used during the production process for vaccines in order to avoid bacterial contamination are considered potential allergens because these antimicrobial agents can cause contact or, rarely, systemic hypersensitivity reactions when used in clinical settings for disease therapy. However, hypersensitivity reactions associated with trace amounts of antibiotics present in vaccines have not been well documented [9]. There is only one ancient reported case of anaphylaxis associated with neomycin in an MMR vaccine [139]. Even though rare, if a patient provides a history of an immediate-type reaction to neomycin or other antibiotics, it is appropriate to investigate with skin testing before immunization with a vaccine containing these constituents. Most patients who develop a non-immediate reaction can receive the vaccine with a low risk of mild reaction outweighed by the benefit of the vaccination [11, 14].



## **5. Focus on influenza vaccine**

### **5.1 General considerations**

Influenza immunization is recommended annually for individuals at risk of severe influenza disease, including young children, pregnant women, people with chronic medical conditions, and the elderly [140-143]. The vaccine formulation changes yearly, based on the strains of influenza anticipated to circulate in the upcoming season [9]. The risk of adverse events following immunization with influenza vaccines (IVs) is therefore a common concern in clinical practice.

IVs are generally prepared by propagation of the virus in embryonated chicken's eggs and thus contain variable and very low amounts of the egg protein ovalbumin. Currently available IVs include the adjuvanted or non-adjuvanted trivalent and quadrivalent inactivated influenza vaccines (IIVs) and live attenuated intranasal trivalent and quadrivalent influenza vaccines (LAIVs). Cell culture-based IIVs (ccIIVs), in which the viruses are grown in animal cells and liquid culture rather than eggs, have been recently developed. However, ccIIVs may contain egg protein, because some of the viruses provided to the manufacturer at the beginning of the process are egg-derived [141]. The only IVs considered to be egg-free are the recombinant trivalent and quadrivalent hemagglutinin influenza vaccines (RIV3, RIV4) [141].

### **5.2 Epidemiology of anaphylaxis following IVs**

The risk of anaphylaxis following administration of trivalent inactivated influenza vaccines (IIV3s) was estimated to be 1.35/million doses between 2009 and 2011 in the United States (US) [58]. The incidence of anaphylaxis following vaccination with a high-dose IIV3, containing four times the standard concentration of hemagglutinin to improve the immune response in adults  $\geq 65$  years of age, was estimated to be one/million distributed doses [144]. However, all distributed doses are not necessarily administered to the patients and thus this figure may be underestimated. The incidence of anaphylaxis following immunization by quadrivalent IIVs (IIV4) for the 2013-2015 seasons in the US was estimated to be 0.17/million distributed doses [145]. In a post licensure analysis of the 521 adverse events reported following the new MF59-adjuvanted trivalent IIV (aIIV3), approved for adults  $\geq 65$  years of age in the US, there were no cases of anaphylaxis, whereas anaphylaxis accounted for 0.2 to 0.4% of adverse events reported for the non-adjuvanted IIVs during the same period [146]. Accordingly, in clinical trials, vaccination with the MF59-adjuvanted trivalent and quadrivalent IIVs was not associated with any particular risk of allergic reaction in the pediatric population relative to vaccination with the non-adjuvanted IIV3s and IIV4s [147, 148]. Seven cases of anaphylaxis were reported after the first two seasons of trivalent LAIV use in the US, during which approximately 2.5 million patients were immunized [149]. In a study assessing a total of 782,125 doses of intranasal LAIVs administered during the 2013-2014 season, no cases of anaphylaxis were reported, whereas more than 6.6 million doses of IIV3 were administered and 15 cases of anaphylaxis

recorded during the same period [150]. Anaphylaxis and hypersensitivity reactions after immunization with RIV3 were reported at a similar frequency as those reported after vaccination with IIV3s [151, 152]. Anaphylaxis following influenza vaccination is a rare event and may occur with all types of IVs.

### **5.3 IV and chicken's egg allergy**

There has been a longstanding concern about the risk of anaphylaxis following administration of IVs to patients with egg allergy, particularly those with previous anaphylactic reactions to egg. This led to changes in manufacturing processes, resulting in vaccines with only trace amounts of ovalbumin and the development of egg-free vaccines. In the US, the ovalbumin content of IVs from 2011 through 2015 was  $\leq 1 \mu\text{g}/0.5 \text{ mL}$  dose for injectable vaccines and  $0.24 \mu\text{g}/0.2 \text{ mL}$  dose for the nasal LAIV [153]. In 1998, James *et al.* demonstrated that children with egg allergy, including those reporting anaphylaxis to egg, could be safely immunized with IIVs containing 0.02-1.2  $\mu\text{g}/\text{mL}$  of egg protein, either in two graded doses or in one single dose [154]. Since then, the safety of IIV3s has been investigated in more than 4,000 children and adult patients with egg allergy, including patients with previous anaphylaxis, resulting in no reported cases of anaphylaxis following immunization with IIV3s [84-87, 155,156]. In a study evaluating the safety of IIV3s possibly containing higher concentration of ovalbumin than the 1.2  $\mu\text{g}/\text{mL}$  usually deemed to be safe, none of the 152 egg-allergic patients receiving 292 doses developed anaphylaxis or mild allergic reactions [88]. During the 2009 influenza pandemic, the risk of anaphylaxis following immunization with the AS03-adjuvanted H1N1 vaccine, containing less than 0.03  $\mu\text{g}/\text{mL}$  of ovalbumin, was compared between 830 children and adult patients with egg allergy and 393 control subjects [83]. None of the patients with egg allergy or the control subjects developed anaphylaxis and the proportion of patients with possible mild allergic reactions was similar in both groups. Overall, these studies showed that injectable IIVs are safe in egg-allergic recipients, even in those with previous severe reactions to egg, and that the risk of allergic reaction following immunization with IIVs appears to be similar between individuals with and without egg allergy. These studies also showed that pre-vaccine skin tests with the vaccines are unnecessary, since they do not predict the occurrence of an allergic reaction following influenza vaccination. However, in these studies, many patients with previous severe allergic reaction to egg and considered to be at high risk of anaphylaxis following IVs were vaccinated in two or more divided doses in a graded approach and not with a single dose. In addition, these studies included both patients naïve for previous IVs and others who received IVs in the past and were thus previously sensitized. The risk of a hypersensitivity reaction following influenza vaccination may differ between these two groups.

The safety of LAIVs has been assessed in 1,129 children with egg allergy, including 412 children with anaphylaxis to egg, receiving 1,330 doses [89,90,157]. Seventeen children experienced mild reactions post immunization, and no anaphylaxis was observed. In these studies, the concentration of

ovalbumin in the LAIVs was  $< 0.24 \mu\text{g}/\text{dose}$ . Interestingly, during intranasal challenges with egg protein performed in eight children, no symptoms were elicited at  $1 \mu\text{g}/\text{ml}$  and the concentration of egg protein found to trigger nasal symptoms was  $10 \mu\text{g}/\text{ml}$  or higher [158]. The risk of anaphylaxis following immunization with LAIVs in patients with egg allergy is therefore expected to be lower than with IIVs.

#### **5.4 Other IV components and anaphylaxis**

As for other vaccines, IVs contain various components that may cause allergic reactions. IgE antibodies against viral antigens, such as hemagglutinins were shown to be potential triggers of anaphylaxis after influenza vaccination of children in Japan during the 2011 season [123]. In addition, although very rare, latex present in the vial stopper or syringe plunger was associated with anaphylaxis following influenza vaccination of patients with a latex allergy [118]. Several cases of anaphylaxis have been reported in adults with and without egg allergy after vaccination with RIV3, which does not contain egg proteins, preservatives, or antibiotics, suggesting that other components may be involved [152, 159].

The causal relationship between vaccine components and allergic reactions is however difficult to confirm. For example, the generation of specific IgE antibodies against H1N1, H3N2, and B influenza vaccine components is part of the normal immune response to the vaccine, especially in young children [160]. Among patients from Canada who presented allergic symptoms within 24 hours following immunization with the AS03-adjuvanted monovalent pandemic H1N1 vaccine in 2009, an IgE-mediated mechanism was rarely demonstrated [161]. Skin-prick tests (SPTs) and intradermal tests (IDTs) with the vaccine and its components were positive in only 4% of cases, 3% of control subjects, and 9% of patients with anaphylaxis. Of note, the diagnostic value of skin testing is considered to be low for IVs. In healthy adult volunteers, IDTs to IIV3 were found to be falsely positive for 3 of 20 subjects at a 1:100 dilution, 11/20 subjects at a 1:10 dilution, and 13/20 at full strength [162]. Finally, data from a case-control study performed to determine risk factors for anaphylaxis and allergic-like events following immunization with the AS03-adjuvanted monovalent pandemic H1N1 vaccine in Canada identified food allergies and acute respiratory illness at the time of the vaccination as potential risk factors [163].

#### **5.5 Allergy workup**

##### **5.5.1 Egg allergy**

There is now strong evidence that individuals with egg allergy can receive any licensed age appropriate IV. Patients with non-severe egg allergy can be immunized under the same conditions as nonallergic patients, without specific precautions [14, 141, 142]. A single dose of IVs is recommended for patients who have experienced anaphylaxis after egg consumption [14, 141, 142].

In this case, some guidelines recommend administering IV without any additional precautions, given that standard vaccination practice includes the ability to recognize and manage severe hypersensitivity reactions [142]. Other guidelines [14, 141] state that IVs should be administered to patients with previous anaphylaxis to egg by an experienced staff in an inpatient or outpatient setting with 15 minutes [141], or a minimum of one hour post vaccination surveillance period [14].

### **5.5.2 Previous immediate reactions following influenza vaccination**

For systemic reactions occurring within the two to four hours following influenza vaccination, an allergic workup, including an SPT with the undiluted vaccine and, if negative, IDT with the vaccine (1:100 and, if negative, 1:10 dilutions) is indicated to show evidence of an IgE-mediated mechanism. If skin-tests are positive, the diagnosis of allergy is confirmed, and further administration should be performed in a clinical setting with graded doses in one day using an intravenous line and the patient should be observed for two hours post-immunization [14].

If skin-tests are negative, the diagnosis of allergy is not excluded, and management depends on the severity of the previous reaction to vaccine. In case of a previous anaphylaxis following influenza vaccination, the patient should be immunized in a clinical setting in two divided doses of 10% of the total vaccine dose and then the other 90% 30 minutes later. If there is a non-severe reaction (urticaria), the patient should be immunized with a single dose in a clinical setting. In both cases, a two-hour post-vaccination surveillance period is required [14].

## **6. Conclusion**

Severe hypersensitivity reactions to vaccines are a very rare eventuality and even rarer are the subsequent contraindications to the second dose of the same vaccine or especially to other vaccines. In patients without a history of allergy, with an allergic disease not related to a vaccine, or with a family history of allergy, no precaution prior to immunization with all types of vaccines is necessary. On the other hand, all patients with prior suspected hypersensitivity reaction to a vaccine have to be evaluated by an allergist to formulate the best approach for subsequent immunization and to avoid having children labeled as “allergic to vaccine” before a certain diagnosis. In all cases, routine vaccinations need to be administered in an adequate setting with trained personnel, medications and equipment needed to treat hypersensitivity reactions.

## **7. Expert opinion**

With policies strengthening the indications for vaccination in Europe, the problem of adverse events following immunization is becoming more stringently identified and regulated in clinical practice.

The main concern is the occurrence of anaphylaxis. Recent data showed that the incidence of anaphylaxis is approximately 1 case per million injected doses, and death is exceedingly rare. In

patients with a suspicion of IgE-mediated reaction, an allergy workup is required if further immunizations are needed, both to avoid further potentially life-threatening reactions and to identify the causal agent that might lead to a hypersensitivity reaction in other situations (i.e. latex, gelatin). It is important to note that even in the case of positive allergy workup, the vaccine is not contraindicated, and can be administered according to a desensitization protocol, under medical supervision. From another point of view, injection site reactions are the most common adverse event following injection of vaccines and constitute one of the main post-vaccination issues. Although most of these reactions are benign, there are clearly associated with decreases in the vaccination rate. This is mainly due to fear of anaphylactic reaction during recall injection.

Insufficient understanding and knowledge of the real risk of severe hypersensitivity reactions to vaccines is responsible for the fact that too many patients are still needlessly referred to hospital for vaccine injection because of the fear of potential severe reactions. For the same reason, vaccinations are too often delayed even in the case of non-immediate or local reactions. Improving knowledge of side effects and their management is therefore crucial in the promotion of vaccinations to protect both the individual and the community at a time when anti-vaccination movements are very active. Communication skills need to be upgraded, improved and targeted to patients and their families in order to fully explain the different scenarios associated with hypersensitivity reactions to vaccines. Moreover, the aim of successful management of suspected vaccine hypersensitivity reactions, at least in the case of the most frequently used vaccines in European countries (MMR/MMRV, influenza, DTP), is to reduce hospital admissions for the administration of vaccinations in a protected environment and to therefore stimulate the practice and belief that vaccines can be safely administered directly by local doctors.

In our opinion, future research should be aimed at identifying adverse reactions to lesser-known vaccines, such as yellow fever and Japanese encephalitis, increasingly needed due to the prevalence of travel in a globalized world. Studies in this area are still limited to date.

Furthermore, there has been a longstanding concern about the risk of anaphylaxis following administration of vaccines containing small amounts of egg proteins to patients with egg allergy, particularly those with previous anaphylactic reactions to egg. It is now clear that egg allergy is not a contraindication to influenza and MMR/MMRV vaccination. However, further large and multicentric studies are urgently needed to assess the real diagnostic value of skin test in egg allergic patients who need to be vaccinated with YF vaccine and the necessity of performing a graded dose administration versus full dose, as has been recently performed for influenza vaccine in egg allergic children.

In addition, more data are needed to provide correct definition of the incidence of anaphylaxis to vaccines with studies based on a confirmatory allergy work-up, especially in patients allergic to foods

such as milk and egg. At the same time, it is important to try limiting specific allergy tests to patients who have a clinical history of a reaction to a specific vaccine. Specifically, pediatric data are needed in order to focus on specific issues that could be associated with the hypersensitivity reactions management in this age group.

Changes in manufacturing processes of vaccines continue to increase their safety profiles.

In fact, to minimize the risk of hypersensitivity reactions the development of newer vaccines which use new manufacturing processes is crucial. For example, the use of new adjuvants in order to decrease the frequency of local reactions that have been responsible for a decreased vaccine coverage rate. An example of recent vaccines associated with low risk of hypersensitivity reaction are the recombinant trivalent and quadrivalent hemagglutinin influenza vaccines, which are considered to be egg-free, and devoid of preservatives and antibiotics.

In this regard, there is still a need for more research on newer adjuvants used in vaccines also taking into account that some adjuvanted vaccines are administered together with a potential additional risk which is not something evaluated in clinical trials.

Other routes of immunization such as the intranasal route have also demonstrated their safety for annual immunization against influenza. Recent development strategies have targeted specific populations which seems to be a productive path for more research. For example, the elderly population is a high-risk group for developing severe influenza disease. and aging is associated with a decreased immune response to vaccine. Influenza vaccines containing higher titers of hemagglutinin or adjuvant to increase the immune response in this age-group have now been developed and used in the US since the 2016-2017 season, with a good safety profile.

To summarize, the ultimate goal in this field is to establish unique guidelines to help identify potential patients who might require specific allergy workup and vaccination in a hospital setting while trying to guarantee a safe and desirable level of vaccination coverage for the entire population. In terms of clinical practice, this knowledge could lead to the definition and establishment of a network between first level vaccination centers and specialized referral centers, in order to formulate a shared management environment for selected cases of potential hypersensitivity reactions to vaccines. This would help immensely to maximize the efficiency and optimize the cost-benefit ratio of the vaccination process.

### **Funding**

This paper was not funded.

### **Declaration of interest**

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

### Reviewer disclosures

Peer reviewers on this manuscript have no relevant financial or other relationships to disclose.

### References

Papers of special note have been highlighted as:

\* of interest

\*\* of considerable interest

1. Bozzola E, Spina G, Russo R, et al. Mandatory vaccinations in European countries, undocumented information, false news and the impact on vaccination uptake: the position of the Italian pediatric society. *Ital J Pediatr.* 2018;44(1):67.
2. Parrella A, Gold M, Marshall H. et al. Parental perspectives of vaccine safety and experience of adverse events following immunisation. *Vaccine.* 2013;31(16):2067-2074.
3. Aronson JK, Ferner RE. Clarification of terminology in drug safety. *Drug Saf.* 2005;28(10):851-870.
4. Echeverría-Zudaire LA, Ortigosa-del Castillo L, Alonso-Lebrero E, et al. Consensus document on the approach to children with allergic reactions after vaccination or allergy to vaccine components. *Allergol Immunopathol (Madr).* 2015;43(3):304-325.
5. Demoly P, Adkinson NF, Brockow K, et al. International Consensus on drug allergy. *Allergy.* 2014;69(4):420-437.  
**\* International CONsensus (ICON) on drug allergy issued by the International Collaboration in Asthma, Allergy and Immunology (iCAALL), formed by the European Academy of Allergy and Clinical Immunology (EAACI), the American Academy of Allergy, Asthma and Immunology (AAAAI), the American College of Allergy, Asthma and Immunology (ACAAI), and the World Allergy Organization (WAO)**
6. McNeil MM, DeStefano F. Vaccine-associated hypersensitivity. *J Allergy Clin Immunol.* 2018;141(2):463-472.

7. Baxter CM, Clothier HJ, Perrett KP. Potential immediate hypersensitivity reactions following immunization in preschool aged children in Victoria, Australia. *Hum Vaccin Immunother.* 2018;14(8):2088-2092.
8. Micheletti F, Peroni D, Piacentini G, et al. Vaccine allergy evaluation and management at the specialized Green Channel Consultation Clinic. *Clin Exp Allergy.* 2012;42:1088-1096.
9. Dreskin SC, Halsey NA, Kelso JM, et al. International Consensus (ICON): allergic reactions to vaccines. *World Allergy Organ J.* 2016;9(1):32.  
**\*\* International Consensus (ICON) on allergic reaction to vaccines with the active participation of representatives from the World Allergy Organization (WAO), the European Academy of Allergy and Clinical Immunology (EAACI), the American Academy of Allergy, Asthma, and Immunology (AAAAI), and the American College of Allergy, Asthma, and Immunology (ACAAI)**
10. Coombs RRA, Gell PGH. Classification of allergic reactions responsible for drug hypersensitivity reactions. In: Coombs RRA, Gell PGH editors, *Clinical Aspects of Immunology*, Philadelphia (PA): Davies; 1968. p. 575-596.
11. Caubet JC, Ponvert C. Vaccine allergy. *Immunol Allergy Clin North Am.* 2014;34:597-613.
12. Kelso JM, Greenhawt MJ, Li JT, et al. Adverse reactions to vaccines practice parameter 2012 update. *J Allergy Clin Immunol* 2012;130:25-43.
13. WHO. Immunization safety surveillance: guidelines for immunization programme managers on surveillance of adverse events following immunization. [Internet] 2015 [accessed 2020 Mar 10]; Available from:  
[https://iris.wpro.who.int/bitstream/handle/10665.1/12620/9789290617457\\_eng.pdf](https://iris.wpro.who.int/bitstream/handle/10665.1/12620/9789290617457_eng.pdf).
14. Nilsson L, Brockow K, Alm J, et al. Vaccination and allergy: EAACI position paper, practical aspects. *Pediatr Allergy Immunol* 2017;28:628-640.  
**\*\* Position paper on vaccination and allergy of the European Academy of Allergy and Clinical Immunology (EAACI)**
15. Morin E, Sadarangani M. Recurrent immune thrombocytopenia following different vaccines. *BMJ Case Rep.* 2019;12(9).
16. Khetsuriani N, Imnadze P, Baidoshvili L, et al. Impact of unfounded vaccine safety concerns on the nationwide measles-rubella immunization campaign, Georgia, 2008. *Vaccine.* 2010;28(39):6455-6462.
17. Nicolosi L, Vittucci A, Mancini R, et al. Vaccine risk assessment in children with a referred reaction to a previous vaccine dose: 2009-2011 retrospective report at the Bambino Gesù' children hospital, Rome, Italy. *Ital J Pediatr.* 2014;40:31.
18. Bonds RS, Kelly BC. Severe serum sickness after H1N1 influenza vaccination. *Am J Med Sci.* 2013;345(5):412-413.



19. Apisarnthanarak A, Uyeki TM, Miller ER, et al. Serum sickness-like reaction associated with inactivated influenza vaccination among Thai health care personnel: risk factors and outcomes. *Clin Infect Dis*. 2009 1;49(1):e18-22.
20. Laribièrè A, Miremont-Salamé G, Reyre H, et al. Surveillance of adverse effects during a vaccination campaign against meningitis C. *Eur J Clin Pharmacol*. 2005;61(12):907-911.
21. Peng B, Wei M, Zhu FC, et al. The vaccines-associated Arthus reaction. *Hum Vaccin Immunother*. 2019 4:1-9.
22. Wang MH, Hu A, Lee YS, et al. Arthus Reaction. *J Emerg Med*. 2019 Apr;56(4):450-451.
23. Wu YL, Tsai MH, Liu LL. Erythema nodosum and hepatitis B: a case report and literature review. *J Microbiol Immunol Infect*. 2008;41(5):437-439.
24. Cohen PR. Combined reduced-antigen content tetanus, diphtheria, and acellular pertussis (tdap) vaccine-related erythema nodosum: case report and review of vaccine-associated erythema nodosum. *Dermatol Ther (Heidelb)*. 2013;3(2):191-197.
25. Da Dalt L, Zerbinati C, Strafella MS, et al. Italian Multicenter Study Group for Drug and Vaccine Safety in Children. Henoch-Schönlein purpura and drug and vaccine use in childhood: a case-control study. *Ital J Pediatr*. 2016 18;42(1):60.
26. Ebo DG, Bridts CH, Stevens WJ. IgE-mediated large local reaction from recombinant hepatitis B vaccine. *Allergy*. 2008;63:483-484.
27. Barbaud A, Tréchet P, Reichert-Pénétrat S, et al. Allergic mechanisms and urticaria/angioedema after hepatitis B immunization. *Br J Dermatol* 1998; 139: 925-926.
28. Barbaud A, Deschildre A, Waton J, et al. Hypersensitivity and vaccines: an update. *Eur J Dermatol*. 2013 1;23(2):135-141.
29. Caubet JC, Rudzeviciene O, Gomes E, et al. Managing a child with possible allergy to vaccine. *Pediatr Allergy Immunol*. 2014;25(4):394-403.
30. Yeh SH, Gurtman A, Hurley DC, et al. Immunogenicity and safety of 13-valent pneumococcal conjugate vaccine in infants and toddlers. *Pediatrics*. 2010;126(3):e493-505.
31. Guerra FA, Blatter MM, Greenberg DP, et al. Safety and immunogenicity of a pentavalent vaccine compared with separate administration of licensed equivalent vaccines in US infants and toddlers and persistence of antibodies before a preschool booster dose: a randomized, clinical trial. *Pediatrics*. 2009;123(1):301-312.
32. Gold MS, Noonan S, Osbourn M, et al. Local reactions after the fourth dose of acellular pertussis vaccine in South Australia. *Med J Aust*. 2003;179(4):191-194.
33. Skowronski DM, Remple VP, Macnabb J, et al. Injection-site reactions to booster doses of acellular pertussis vaccine: rate, severity, and anticipated impact. *Pediatrics*. 2003;112(6 Pt 1):e453.

34. Gajdos V, Soubeyrand B, Vidor E, et al. Immunogenicity and safety of combined adsorbed low-dose diphtheria, tetanus and inactivated poliovirus vaccine (REVAXIS ®) versus combined diphtheria, tetanus and inactivated poliovirus vaccine (DT Polio ®) given as a booster dose at 6 years of age. *Hum Vaccin*. 2011;7(5):549-556.
35. Rennels MB, Deloria MA, Pichichero ME, et al. Extensive swelling after booster doses of acellular pertussis-tetanus-diphtheria vaccines. *Pediatrics*. 2000;105(1):e12.
36. Woo EJ, Burwen DR, Gatumu SN, et al. Extensive limb swelling after immunization: reports to the Vaccine Adverse Event Reporting System. *Clin Infect Dis* 2003;37(3):351-358.
37. Southern J, Waight PA, Andrews N, et al. Extensive swelling of the limb and systemic symptoms after a fourth dose of acellular pertussis containing vaccines in England in children aged 3-6years. *Vaccine*. 2017;35(4):619-625.
38. Schmitt HJ, Beutel K, Schuind A, et al. Reactogenicity and immunogenicity of a booster dose of a combined diphtheria, tetanus, and tricomponent acellular pertussis vaccine at fourteen to twenty-eight months of age. *J Pediatr*. 1997;130(4):616-623.
39. Marshall HS, Gold MS, Gent R, et al. Ultrasound examination of extensive limb swelling reactions after diphtheria-tetanus-acellular pertussis or reduced-antigen content diphtheria-tetanus-acellular pertussis immunization in preschool-aged children. *Pediatrics*. 2006;118(4):1501-1509.
40. Rennels MB, Black S, Woo EJ, et al. Safety of a fifth dose of diphtheria and tetanus toxoid and acellular pertussis vaccine in children experiencing extensive, local reactions to the fourth dose. *Pediatr Infect Dis J* 2008;27(5):464-465.
41. Jackson LA, Yu O, Nelson JC, et al. Injection site and risk of medically attended local reactions to acellular pertussis vaccine. *Pediatrics*. 2011;127(3):e581-587.
42. Jackson LA, Peterson D, Nelson JC, et al. Vaccination site and risk of local reactions in children 1 through 6 years of age. *Pediatrics*. 2013;131(2):283-289.
43. Bergfors E, Hermansson G, Nyström Kronander U, et al. How common are long-lasting, intensely itching vaccination granulomas and contact allergy to aluminium induced by currently used pediatric vaccines? A prospective cohort study. *Eur J Pediatr*. 2014;173(10):1297-1307.
44. Bergfors E, Trollfors B, Inerot A. Unexpectedly high incidence of persistent itching nodules and delayed hypersensitivity to aluminium in children after the use of adsorbed vaccines from a single manufacturer. *Vaccine*. 2003;22(1):64-69.

45. Bergfors E, Trollfors B. Sixty-four children with persistent itching nodules and contact allergy to aluminium after vaccination with aluminium-adsorbed vaccines-prognosis and outcome after booster vaccination. *Eur J Pediatr.* 2013;172(2):171-177.
46. Bergfors E, Björkelund C, Trollfors B. Nineteen cases of persistent pruritic nodules and contact allergy to aluminium after injection of commonly used aluminium-adsorbed vaccines. *Eur J Pediatr.* 2005;164(11):691-697.
47. Salik E, Løvik I, Andersen KE, et al. Persistent Skin Reactions and Aluminium Hypersensitivity Induced by Childhood Vaccines. *Acta Derm Venereol.* 2016;96(7):967-971.
48. Bergfors E, Inerot A, Falk L, et al. Patch testing children with aluminium chloride hexahydrate in petrolatum: A review and a recommendation. *Contact Dermatitis.* 2019;81(2):81-88.
49. Goiset A, Darrigade AS, Labrèze C, et al. Aluminium sensitization in a French paediatric patch test population. *Contact Dermatitis.* 2018;79(6):382-383.
50. Gente Lidholm A, Bergfors E, Inerot A et al. Unexpected loss of contact allergy to aluminium induced by vaccine. *Contact Dermatitis.* 2013;68(5):286-292.
51. Pittman PR. Aluminium-containing vaccine associated adverse events: role of route of administration and gender. *Vaccine.* 2002;20:S48-50.
52. Diggle L, Deeks J, Pollard A. Effect of needle size on immunogenicity and reactogenicity of vaccines in infants: a randomized, controlled trial. *Brit Med J.* 2006;333:571-578.
53. Bohlke K, Davis RL, Marcy SM, et al. Risk of anaphylaxis after vaccination of children and adolescents. *Pediatrics.* 2003;112:815-820.
54. Stratton K, Ford A, Rusch E, et al. editors, *Adverse Effects of Vaccines: Evidence and Causality*, Washington (DC): National Academies Press; 2012.
55. Su JR, Moro PL, Ng CS, et al. Anaphylaxis after vaccination reported to the Vaccine Adverse Event Reporting System, 1990-2016. *J Allergy Clin Immunol* 2019;143(4):1465-1473.
56. Shimabukuro TT, Nguyen M, Martin D, et al. Safety monitoring in the Vaccine Adverse Event Reporting System (VAERS). *Vaccine.* 2015;33:4398-4405.
57. Ruggeberg JU, Gold MS, Bayas JM, et al. Anaphylaxis: case definition and guidelines for data collection, analysis, and presentation of immunization safety data. *Vaccine.* 2007;25:5675-5684.
58. McNeil MM, Weintraub ES, Duffy J, et al. Risk of anaphylaxis after vaccination in children and adults. *J Allergy Clin Immunol* 2016;137:868-878.

59. McNeil MM, Gee J, Weintraub ES, et al. The Vaccine Safety Datalink: successes and challenges monitoring vaccine safety. *Vaccine*. 2014;32:5390-5398.
60. Institute For Vaccine Safety Johns Hopkins Bloomberg School Of Public Health. Potential adverse events following immunization. [Internet] 2019 [accessed 2020 Mar 10]; Available from: <http://www.vaccinesafety.edu/vs-list-of-AEs.htm>
61. Radice A, Carli G, Macchia D, et al. Allergic reactions after vaccination: translating guidelines into clinical practice. *Eur Ann Allergy Clin Immunol*. 2019;51(2):51-61.
62. Wood RA, Berger M, Dreskin SC, et al. An algorithm for treatment of patients with hypersensitivity reactions after vaccines. *Pediatrics*. 2008;122:e771-e777.
63. Fritsche PJ, Helbling A, Ballmer-Weber BK. Vaccine hypersensitivity-update and overview. *Swiss Med Wkly*. 2010;140:238-246.
64. Active and passive immunization. In: Pickering LK, Baker CJ, Kimberlin DW, et al. editors, *Red Book: 2009 Report of the Committee on Infectious Diseases*, Elk Grove Village (IL): American Academy of Pediatrics; 2009. p. 48.
65. Das S, Mondal S. Tetanus Toxoid Induced Anaphylaxis. *J Vaccines Vaccin*. 2012;3:126.
66. Ponvert C, Scheinmann P, de Blic J. Anaphylaxis to the 23-valent pneumococcal vaccine: a second explored case by means of immediate-reading skin tests with pneumococcal vaccines. *Vaccine*. 2010;28(52):8256-8257.
67. Arroabarren E, Anda M, Sanz ML. Anaphylaxis to pneumococcal vaccine; CRM (197): novel cause of vaccine allergy. *Pediatr Allergy Immunol*. 2016;27:433-437.
68. Institute For Vaccine Safety Johns Hopkins Bloomberg School Of Public Health. Potential allergen in vaccines per 0.5 ml dose. [Internet] 2018 [accessed 2020 Mar 10]; Available from: <http://vaccinesafety.edu/components-Allergens.htm>.
69. Netterlid E, Hindsén M, Björk J, et al. There is an association between contact allergy to aluminium and persistent subcutaneous nodules in children undergoing hyposensitization therapy. *Contact Dermatitis*. 2009;60:41-49.
70. Haag CK, Dacey E, Hamilton N, et al. Aluminum granuloma in a child secondary to DTaP-IPV vaccination: A case report. *Pediatr Dermatol*. 2019;36(1):e17-e19.
71. Yu AM, Ito S, Leibson T, et al. Pediatric Wells syndrome (eosinophilic cellulitis) after vaccination: A case report and review of the literature. *Pediatr Dermatol*. 2018; 35(5): e262-e264.
72. Gordon SC, Bartenstein DW, Tajmir SH, et al. Delayed-type hypersensitivity to vaccine aluminum adjuvant causing subcutaneous leg mass and urticaria in a child. *Pediatr Dermatol*. 2018;35(2):234-236.

73. Lauren CT, Belsito DV, Morel KD, et al. Case Report of Subcutaneous Nodules and Sterile Abscesses Due to Delayed Type Hypersensitivity to Aluminum-Containing Vaccines. *Pediatrics*. 2016;138(4).
74. Leventhal JS, Berger EM, Brauer JA, et al. Cohen DE. Hypersensitivity reactions to vaccine constituents: a case series and review of the literature. *Dermatitis*. 2012; 23(3):102-109.
75. Zheng W, Dreskin SC. Thimerosal in influenza vaccine: an immediate hypersensitivity reaction. *Ann Allergy Asthma Immunol*. 2007; 99(6):574-575.
76. Rouleau I, De Serres G, Drolet JP, et al. Increased risk of anaphylaxis following administration of 2009 AS03-adjuvanted monovalent pandemic A/ H1N1 (H1N1pdm09) vaccine. *Vaccine*. 2013;31:5989–5996.
77. Oberle D, Pavel J, Rieck T, et al. Tenenbaum T. Anaphylaxis After Immunization of Children and Adolescents in Germany. *Pediatr Infect Dis J*. 2016;35(5):535-541.
78. Badiu I, Geuna M, Heffler E, et al. Hypersensitivity reaction to human papillomavirus vaccine due to polysorbate 80. *BMJ Case Rep*. 2012;2012:ber0220125797.
79. Kara Elitok G, Çelikboya E, Bulbul L, et al. Does Food Allergy Require Any Change in Measles-Mumps-Rubella Vaccination? *Indian J Pediatr*. 2019;86(10):915-920.
80. Rutowski K, Ewan PW, Nasser SM. Administration of yellow fever vaccine in patients with egg allergy. *Int Arch Allergy Immunol*. 2013;161:274-278.
81. Greenhawt M, Turner PJ, Kelso JM. Administration of influenza vaccines to egg allergic recipients: A practice parameter update 2017. *Ann Allergy Asthma Immunol*. 2018;120(1):49-52.
82. Cronin J, Scorr A, Russell S, et al. A review of a paediatric emergency department vaccination programme for patients at risk of allergy/anaphylaxis. *Acta Paediatr* 2012;101:941-5.
83. Gagnon R, Primeau MN, Des Roches A, et al. Safe vaccination of patients with egg allergy with an adjuvanted pandemic H1N1 vaccine. *J Allergy Clin Immunol*. 2010;126(2):317–323.
84. Owens G, MacGinnitie A. Higher-ovalbumin-content influenza vaccines are well tolerated in children with egg allergy. *J Allergy Clin Immunol* 2011;127(1):264-265.
85. Fung I, Spergel JM. Administration of influenza vaccine to pediatric patients with egg-induced anaphylaxis. *J Allergy Clin Immunol* 2012 ;129(4):1157-1159.
86. Chung EY, Huang L, Schneider L. Safety of influenza vaccine administration in egg-allergic patients. *Pediatrics* 2010;125(5):e1024-e1030.
87. Des Roches A, Paradis L, Gagnon R, et al; PCIRN (Public Health Agency of Canada/Canadian Institutes of Health Research Influenza Research Network). Egg-allergic patients can be safely vaccinated against influenza. *J Allergy Clin Immunol* 2012;130(5):1213-1216.

88. Webb L, Petersen M, Boden S, et al. Single-dose influenza vaccination of patients with egg allergy in a multicenter study. *J Allergy Clin Immunol* 2011;128(1):218-219.
89. Turner PJ, Southern J, Andrews NJ, et al. Safety of live attenuated influenza vaccine in atopic children with egg allergy. *J Allergy Clin Immunol* 2015;136(2):376-381.
- \* Study assessing the safety of live attenuated influenza vaccine in young people with egg allergy.**
90. Turner PJ, Southern J, Andrews NJ, et al. Safety of live attenuated influenza vaccine in young people with egg allergy: multicentre prospective cohort study. *BMJ* 2015;351:h6291.
91. Fina Aviles F, Campins Marti M, et al. MMR vaccine and egg allergy. Experience in a hospital immunization unit. *An Pediatr (Barc)*. 2007;67(4):362-367
92. Retterer MKC, Workman LJ, Bacon JR, et al. Specific IgE to gelatin as a cause of anaphylaxis to zoster vaccine. *J Allergy Clin Immunol*. 2018;141(5):1956-1957.
93. Sakaguchi M, Nakayama T, Inouye S. Food allergy to gelatin in children with systemic immediate-type reactions, including anaphylaxis, to vaccines. *J Allergy Clin Immunol*. 1996;98:1058-1061.
94. Sakaguchi M, Miyazawa H, Inouye S. Sensitization to gelatin in children with systemic non-immediate-type reactions to varicella vaccines. *Ann Allergy Asthma Immunol*. 2000;84: 341-344.
95. Kelso JM, Jones RT, Yunginger JW. Anaphylaxis to measles, mumps, and rubella vaccine mediated by IgE to gelatin. *J Allergy Clin Immunol*. 1993;91:867-872.
96. Pool V, Braun MM, Kelso JM, et al. Prevalence of anti-gelatin IgE antibodies in people with anaphylaxis after measles-mumps rubella vaccine in the United States. *Pediatrics* 2002;110:e71.
97. Stone CA, Hemler JA, Commins SP, et al. Anaphylaxis after zoster vaccine: Implicating alpha-gal allergy as a possible mechanism. *J Allergy Clin Immunol* 2017;139:1710-3.e2.
98. Sakaguchi M, Nakayama T, Fujita H, et al. Minimum estimated incidence in Japan of anaphylaxis to live virus vaccines including gelatin. *Vaccine*. 2000;19(4-5):431-436.
99. Sakaguchi M, Miyazawa H, Inouye S. Specific IgE and IgG to gelatin in children with systemic cutaneous reactions to Japanese encephalitis vaccines. *Allergy*. 2001;56:536-539.
100. Worm M, Sterry W, Zuberbier T. Gelatin-induced urticaria and anaphylaxis after tick-borne encephalitis vaccine. *Acta Derm Venereol*. 2000;80(3):232.
101. Singer S, Johnson CE, Mohr R, et al. Urticaria following varicella vaccine associated with gelatin allergy. *Vaccine*. 1999;17(4):327-329.
102. Ohsaki M, Tsutsumi H, Kumagai T, et al. The relevance of TH1 and TH2 cells in immediate and nonimmediate reactions to gelatin-containing vaccine. *J Allergy Clin Immunol*. 1999;103(2 Pt 1):276-281.

103. Stone CA, Commins SP, Choudhary S, et al. Anaphylaxis after vaccination in a pediatric patient: further implicating alpha-gal allergy. *J Allergy Clin Immunol Pract* 2019;7(1):322-324.
104. Patja A, Makinen-Kiljunen S, Davidkin I, et al. Allergic reactions to measles-mumps-rubella vaccination. *Pediatrics*. 2001;107:e27.
105. Taniguchi K, Fujisawa T, Ihara T, et al. Gelatin-induced T-cell activation in children with nonanaphylactic-type reactions to vaccines containing gelatin. *J Allergy Clin Immunol*. 1998;102:1028Y1032.
106. Sakaguchi M, Inouye S. IgE sensitization to gelatin: the probable role of gelatin-containing diphtheria-tetanus-acellular pertussis (DTaP) vaccines. *Vaccine*. 2000;18:2055-2058.
107. Kumagai T, Nakayama T, Kamada M, et al. The lymphoproliferative response to enzymatically digested gelatin in subjects with gelatin hypersensitivity. *Clin Exp Allergy* 2000;30:1430-1435.
108. Nakayama T, Aizawa C. Change in gelatin content of vaccines associated with reduction in reports of allergic reactions. *J Allergy Clin Immunol* 2000;106:591-592.
109. Nakayama T, Aizawa C, Kuno-Sakai H. A clinical analysis of gelatin allergy and determination of its causal relationship to the previous administration of gelatin-containing acellular pertussis vaccine combined with diphtheria and tetanus toxoids. *J Allergy Clin Immunol*. 1999;103:321-325.
110. DiMiceli L, Pool V, Kelso JM, et al. Vaccination of yeast sensitive individuals: review of safety data in the US vaccine adverse event reporting system (VAERS). *Vaccine*. 2006;24:703-707.
111. Brightman CA, Scadding GK, Dumbreck LA, et al. Yeast-derived hepatitis B vaccine and yeast sensitivity. *Lancet*. 1989;1:903.
112. Hammond GW, Parker J, Mimms L, et al. Comparison of immunogenicity of two yeast-derived recombinant hepatitis B vaccines. *Vaccine*. 1991;9(2):97-100.
113. Kattan JD, Konstantinou GN, Cox AL, et al. Anaphylaxis to diphtheria, tetanus, and pertussis vaccines among children with cow's milk allergy. *J Allergy Clin Immunol* 2011;128:215-218.
114. Parisi CA, Smaldini PL, Gervasoni ME, et al. Hypersensitivity reactions to the Sabin vaccine in children with cow's milk allergy. *Clin Exp Allergy*. 2013;43:249-254.
115. Slater JE, Rabin RL, Martin D. Comments on cow's milk allergy and diphtheria, tetanus, and pertussis vaccines. *J Allergy Clin Immunol*. 2011;128(2):434.

116. Yavuz ST, Sahiner UM, Sekerel BE, et al. Anaphylactic reactions to measles-mumps-rubella vaccine in three children with allergies to hen's egg and cow's milk. *Acta Paediatr.* 2011;100:e94–e96.
117. Rosales MJ, Alonso E, Ibañez MD, et al. Hypersensitivity reactions to measles, mumps and rubella vaccine. *Allergy.* 1997;52 supplement s37:216-217.
118. Russell M, Pool V, Kelso JM, et al. Vaccination of persons allergic to latex: a review of safety data in the Vaccine Adverse Event Reporting System (VAERS). *Vaccine.* 2004;23:664–666.
119. Lear JT, English JS. Anaphylaxis after hepatitis B vaccination. *Lancet.* 1995;345:1249.
120. Fabry H. Formaldehyde sensitivity. Two interesting cases. *Contact Derm Newsletter* 1968;3:51.
121. Ring J. Exacerbation of eczema by formalin-containing hepatitis B vaccine in formaldehyde-allergic patient. *Lancet.* 1986;2:522–523.
122. Vogt T, Landthaler M, Stolz W. Generalized eczema in an 18-month-old boy due to phenoxyethanol in DPT vaccine. *Contact Dermatitis.* 1998;38:50-51.
123. Nagao M, Fujisawa T, Ihara T, et al. Highly increased levels of IgE antibodies to vaccine components in children with influenza vaccine-associated anaphylaxis. *J Allergy Clin Immunol.* 2016;137:861-867.
124. Ponvert C, Bloch-Morot E. Les réactions d'hypersensibilité allergiques et non allergiques aux vaccins. *Rev Fr Allergol Immunol Clin.* 2013;53:9-17.
125. Anderson DV, Jorgensen IM. MMR vaccination of children with egg allergy is safe. *Dan Med J.* 2013;60:A4573.
126. Franceschini F, Bottau P, Caimmi S, et al. Evaluating children with suspected allergic reactions to vaccines for infectious diseases. *Allergy Asthma Proc.* 2018 1;39(3):177-183.
127. Chung EH. Vaccine allergies. *Clin Exp Vaccine Res.* 2014 Jan;3(1):50-57.
128. Smith D, Wong P, Gomez R, et al. Ovalbumin content in the yellow fever vaccine. *J Allergy Clin Immunol Pract.* 2015;3:794-795.
129. Sakai Y, Yamato R, Onuma M, et al. Non-antigenic and low allergic gelatin produced by specific digestion with an enzyme-coupled matrix. *Biol Pharm Bull.* 1998;21(4):330-334
130. Mullins R, James H, Platts-Mills T, et al. Relationship between red meat allergy and sensitization to gelatin and galactose-a-1,3-galactose. *J Allergy Clin Immunol.* 2012;129:1334-1342.



131. de Silva R, Dasanayake WMDK, Wickramasinhe GD, et al. Sensitization to bovine serum albumin as a possible cause of allergic reactions to vaccines. *Vaccine*. 2017;35(11):1494-1500.
132. Bogdanovic J, Halsey NA, Wood RA, et al. Bovine and porcine gelatin sensitivity in children sensitized to milk and meat. *J Allergy Clin Immunol*. 2009;124:1108-1110.
133. Fischer M, Lindsey N, Staples JE, et al. Japanese encephalitis vaccines: recommendation of the Advisory Committee on Immunization Practices (ACIP) MMWR Morb Mortal Wkly Rep. 2010;59:1-27.
134. Zent O, Hennig R. Post-marketing surveillance of immediate allergic reactions: polygeline-based versus polygeline-free pediatric TBE vaccine. *Vaccine*. 2004 16;23(5):579-584.
135. Nakayama T, Aizawa C. Change in gelatin content of vaccines associated with reduction in reports of allergic reactions. *J Allergy Clin Immunol*. 2000;106(3):591-592.
136. Ozaki T, Nishimura N, Muto T, et al. Safety and immunogenicity of gelatin-free varicella vaccine in epidemiological and serological studies in Japan. *Vaccine*. 2005 26;23(10):1205-1208.
137. Kuno-Sakai H, Kimura M. Removal of gelatin from live vaccines and DTaP-an ultimate solution for vaccine-related gelatin allergy. *Biologicals*. 2003;31(4):245-249.
138. Kelso JM. Allergic reactions after immunization. *Ann Allergy Asthma Immunol*. 2013;110(6):397-401.
139. Kwittken PL, Rosen S, Sweinberg SK. MMR vaccine and neomycin allergy. *Am J Dis Child*. 1993;147:128Y129.
140. WHO. Background Paper on Influenza Vaccines and Immunization, SAGE Working Group. [Internet] 2012 [accessed 2020 Mar 10]; Available from: [https://www.who.int/immunization/sage/meetings/2012/april/1\\_Background\\_Paper\\_Mar26\\_v13\\_cleaned.pdf](https://www.who.int/immunization/sage/meetings/2012/april/1_Background_Paper_Mar26_v13_cleaned.pdf).
141. Grohskopf LA, Alyanak E, Broder KR, et al. Prevention and Control of Seasonal Influenza with Vaccines: Recommendations of the Advisory Committee on Immunization Practices - United States, 2019-20 Influenza Season. *MMWR Recomm Rep*. 2019;68(3):1-21.
142. Committee on Infectious diseases. Recommendations for Prevention and Control of Influenza in Children, 2019-2020. *Pediatrics* 2019. 144(4):e20192478.
143. ECDC. Prevention and Control Influenza vaccination. [Internet] 2013 [accessed 2020 Mar 10]; Available from: [http://ecdc.europa.eu/en/healthtopics/seasonal\\_influenza/vaccines/Pages/influenza\\_vaccination.aspx#vaccinationstrategies](http://ecdc.europa.eu/en/healthtopics/seasonal_influenza/vaccines/Pages/influenza_vaccination.aspx#vaccinationstrategies).

144. Moro PL, Arana J, Cano M, et al. Postlicensure safety surveillance for high-dose trivalent inactivated influenza vaccine in the Vaccine Adverse Event Reporting System, 1 July 2010-31 December 2010. *Clin Infect Dis*. 2012;54(11):1608-1614.
145. Haber P, Moro PL, Lewis P, et al. Post-licensure surveillance of quadrivalent inactivated influenza (IIV4) vaccine in the United States, Vaccine Adverse Event Reporting System (VAERS), 2013-May 31, 2015. *Vaccine* 2016;34(22):2507-2512.
146. Haber P, Moro PL, Ng C, et al. Post-licensure surveillance of trivalent adjuvanted influenza vaccine (aIIV3; Fluad), Vaccine Adverse Event Reporting System (VAERS), United States. *Vaccine*. 2019;37(11):1516-1520.
147. Vesikari T, Kirstein J, Devota Go G, et al. Efficacy, immunogenicity, and safety evaluation of an MF59-adjuvanted quadrivalent influenza virus vaccine compared with non-adjuvanted influenza vaccine in children: a multicentre, randomised controlled, observer-blinded, phase 3 trial. *Lancet Respir Med*. 2018;6(5):345-356.
148. Nolan T, Bravo L, Ceballos A, et al. Enhanced and persistent antibody response against homologous and heterologous strains elicited by a MF59-adjuvanted influenza vaccine in infants and young children. *Vaccine*. 2014;32(46):6146-6156.
149. Izurieta HS, Haber P, Wise RP, et al. Adverse events reported following live, cold-adapted, intranasal influenza vaccine. *JAMA* 2005;294(21):2720-2725.
150. Yih WK, Kulldorff M, Sandhu SK, et al. Prospective influenza vaccine safety surveillance using fresh data in the Sentinel System. *Pharmacoepidemiol Drug Saf* 2016;25(5):481-492.
151. Woo EJ, Moro PL, Cano M, et al. safety surveillance of trivalent recombinant influenza vaccine: Reports to the Vaccine Adverse Event Reporting System. *Vaccine*. 2017;35(42):5618-5621.
152. Izikson R, Leffell DJ, Bock SA, et al. Randomized comparison of the safety of Flublok® versus licensed inactivated influenza vaccine in healthy, medically stable adults  $\geq$  50 years of age. *Vaccine*. 2015;33(48):6622-6628.
153. CDC. influenza Vaccine and People with Egg Allergies. [Internet] 2019 [accessed 2020 Mar 10]; Available from: <https://www.cdc.gov/flu/prevent/egg-allergies.htm>.
154. James JM, Zeiger RS, Lester MR, et al. Safe administration of influenza vaccine to patients with egg allergy. *J Pediatr*, 1998;133(5):624-628.
155. Greenhawt MJ, Spergel JM, Rank MA, et al. Safe administration of the seasonal trivalent influenza vaccine to children with severe egg allergy. *Ann Allergy Asthma Immunol*. 2012;109(6):426-430.
156. Esposito S, Gasparini C, Martelli A, et al. Safe administration of an inactivated virosomal adjuvanted influenza vaccine in asthmatic children with egg allergy. *Vaccine*. 2008;26(36):4664-8.

157. Des Roches A, Samaan K, Graham F, et al. Safe vaccination of patients with egg allergy by using live attenuated influenza vaccine. *J Allergy Clin Immunol Pract*. 2015;3(1):138-139.
158. Turner PJ, Erlewyn-Lajeunesse M. Intranasal live-attenuated influenza vaccine (LAIIV) is unlikely to cause egg-mediated allergic reactions in egg-allergic children. *J Allergy Clin Immunol Pract* 2015;3(2):312-313.
159. Woo EJ, Moro PL, Cano M, Jankosky C. Postmarketing safety surveillance of trivalent recombinant influenza vaccine: Reports to the Vaccine Adverse Event Reporting System. *Vaccine*. 2017;35(42):5618-5621.
160. Nakayama T. Causal relationship between immunological responses and adverse reactions following vaccination. *Vaccine*. 2019;37(2):366-371.
161. Rouleau I, De Serres G, Drolet JP, et al. Public Health Agency of Canada–Canadian Institutes for Health Research Influenza Research Network. Allergic symptoms after pandemic influenza vaccination rarely mediated by vaccine-specific IgE. *J Allergy Clin Immunol*. 2012;130(6):1423-1426.
162. Wood RA, Setse R, Halsey N; Clinical Immunization Safety Assessment (CISA) Network Hypersensitivity Working Group. Irritant skin test reactions to common vaccines. *J Allergy Clin Immunol*. 2007;120(2):478-481.
163. Rouleau I, De Serres G, Skowronski DM, et al. Risk factors associated with anaphylaxis and other allergic-like events following receipt of 2009 monovalent AS03-  
adjuvanted pandemic influenza vaccine in Quebec, Canada. *Vaccine*. 2014;32(28):3480-3487.

**Figure 1: Diagnostic workup for Immediate systemic reactions to vaccines.**

§ 1/10 concentration for SPT with vaccine is recommended in cases of severe anaphylaxis (63)

Useful information for the management of vaccine allergy can be obtained by checking the following links (62):

1. <http://www.vaccinesafety.edu/components-Allergens.htm>: list of allergens and where they are contained;
2. <http://www.cdc.gov/vaccines/pubs/pinkbook/downloads/appendices/B/latex-table.pdf> : list of vaccines at risk for latex allergic patients
3. <https://www.cdc.gov/vaccines/pubs/pinkbook/downloads/appendices/B/excipient-table-2.pdf>: media used for vaccines and excipients.

ACCEPTED MANUSCRIPT

**Table 1: Potential vaccine allergens: adjuvant, preservatives, antibiotics, carrier proteins (modified 68)**

Excipient	Type	Vaccine type	Type of hypersensitivity reaction	References
Aluminium	Adjuvant	DTaP/Tdap/DT/MEN B/HepB/HepA/HPV/DTaP+IPV/DTaP+IPV+HepB/Hib+HepB/ DTaP+IPV+Hib/PCV 13.	Delayed type hypersensitivity reactions (contact allergy, small granulomas, nodules)	43, 45, 47, 48, 69-73
Thimerosal	Preservative	DT/Td/influenza/Japanese encephalitis/Meningococcal	Contact allergy, systemic allergic reaction (rare)	6, 74, 75
ASO-3	Adjuvant	ASO-3 adjuvanted A/H1N1 pandemic influenza vaccine	Anaphylaxis and other immediate hypersensitivity reaction	74, 76, 77
Neomycin	Antimicrobial	Influenza/HepA/IPV/DTaP+IPV/MMR/DTaP+HepB+IPV/DTaP+IPV+Hib/MMRV/DTaP+IPV/Rabies/HepA/Varicella/Influenza/HepA+HepB	comment in the main text	
Polymixin B	Antimicrobial	Influenza/Polio/DTaP+IPV/DTaP+HepB+IPV	comment in the main text	
<b>Polysorbate 80</b>	Surfactant	HPV/influenza/HepB/DTaP/Japanese Encephalitis/ DTaP+IPV/ DTaP+HepB+IPV/DTaP+IPV+Hib/PCV13/Rotavirus/MEN B	Anaphylaxis and other immediate hypersensitivity reaction	78

**Abbreviation:** DTaP- diphtheria, tetanus and acellular pertussis; Tdap- tetanus, reduced diphtheria and acellular pertussis; DT- diphtheria, tetanus; Td- Tetanus and Diphtheria Toxoids adsorbed; MEN B- Meningococcal group B; HepB- Hepatitis B; HepA- Hepatitis A; HPV- Human papillomavirus; IPV- inactivated polio vaccine; Hib- Haemophilus influenzae type b; PCV13- Pneumococcal 13-valent; ASO-3- trade name for a squalen-based adjuvant; MMR- Measles, Mumps, Rubella; MMRV- Measles, Mumps, Rubella, Varicella;

**Table 2: Other potential vaccine allergens (modified 68)**

Excipient	Type	Vaccine type	Type of hypersensitivity reaction	References
<b>Potential allergens</b>				
Egg (ovoalbumin, egg protein)*	Residual medium	Influenza, MMR, YF, TBE.	Minor/local hypersensitivity reaction (macular rash, urticarial rash), anaphylaxis (rare)	79-91
Gelatin	Manufacturing residue/stabilizer	YF/MMR/MMRV/Varicella/influenza/Varicella Zoster/Japanese encephalitis/TBE	Immediate-type (anaphylaxis) and delayed-type (localized erythema, induration at the injection site) hypersensitivity reactions.	92-109
Yeast (Saccharomyces cerevisiae)	Medium nutrient	Hib+HepB/HepB/HPV/Meningococcal/DTaP+HepB+IPV/PCV13/HepB/HepA+HepB/Typhoid	Anaphylaxis and other immediate hypersensitivity reaction	110-112
Milk	Medium nutrient	DTaP/Td/Tdap/OPV/Typhoid fever (oral)/MMR	Anaphylaxis and other immediate hypersensitivity reaction	113-117
Latex**	Pharmaceutical closure	Tdap/Meningococcal/Hip+HepB/HepB/Influenza/HepA/HepB+HepA/DTaP/DTaP+IPV/DTaP+HepB+IPV/Rotavirus/Td	Anaphylaxis and other immediate hypersensitivity reaction	118-119

**Abbreviation:** DTaP- diphtheria, tetanus and acellular pertussis; Tdap- tetanus, reduced diphtheria and acellular pertussis; DT- diphtheria, tetanus; Td- Tetanus and Diphtheria Toxoids adsorbed; MEN B- Meningococcal group B; HepB- Hepatitis B; HepA- Hepatitis A; HPV- Human papillomavirus; IPV- inactivated polio vaccine; Hib- Haemophilus influenzae type b; PCV13- Pneumococcal 13-valent; AS0-3- trade name for a squalen-based adjuvant; MMR- Measles, Mumps, Rubella; MMRV- Measles, Mumps, Rubella, Varicella; YF- yellow fever; OPV- oral polio vaccine; TBE- tick born encephalitis.

\* Different vaccines are at risk of contain small amounts of residual egg proteins from the vaccine manufacturing process, concentrations are usually higher in vaccines cultured on embryonated chicken eggs (influenza, yellow fever, and rabies) and lower for vaccines cultured on fibroblasts of chicken embryos (MMR/MMRV, TBE).

\*\* Latex may be present in the rubber stopper of some vaccine vials and plungers in some prefilled syringes

ACCEPTED MANUSCRIPT

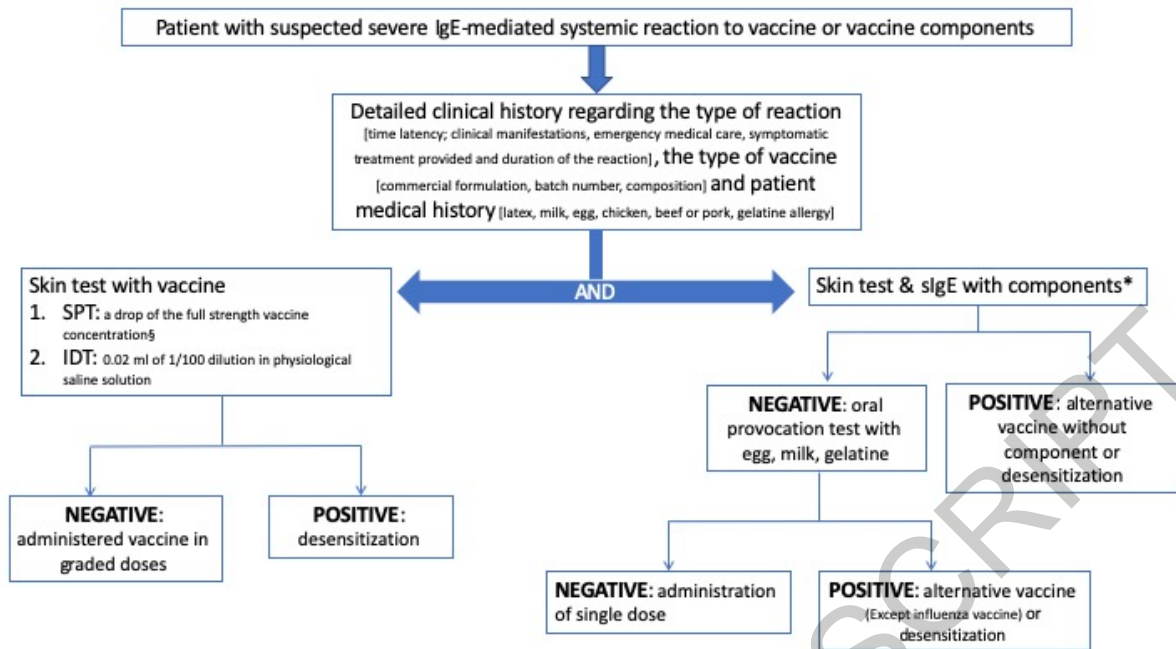


Fig 1

ACCEPTED MANUSCRIPT