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# How to cite

CRETTON, Sylvian et al. Pyridine-4(1*H*)-one Alkaloids from *Waltheria indica* as Antitrypanosomatid Agents. In: Journal of Natural Products, 2020, vol. 83, n° 11, p. 3363–3371. doi: 10.1021/acs.jnatprod.0c00671

This publication URL:https://archive-ouverte.unige.ch/unige:150312Publication DOI:10.1021/acs.jnatprod.0c00671

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# Pyridine-4(1H)-one Alkaloids from Waltheria indica as Antitrypanosomatid Agents

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**ABSTRACT:** Twelve new pyridine-4(1*H*)-one derivatives, namely 8-

demethoxywaltherione F (1), waltheriones R-V (2, 6, 7, 10, and 11), 1-

methoxywaltherione O (**3**), (*S*)-15-hydroxywaltherione G (**4**), (8*R*)-8-hydroxywaltherione M (**5**), (9*S*,13*S*)-2-hydroxymethylwaltherione C and (9*S*,10*S*,13*S*)-10-hydroxywaltherione C (**8** and **9**), (*S*)-13-methoxywaltherione V (**12**), as well as melovinone (**13**) and 5'methoxywaltherione A (**14**) were isolated from the CH<sub>2</sub>Cl<sub>2</sub> extract of the aerial parts of *Waltheria indica*. Their chemical structures were determined by means of a comprehensive analysis including <sup>1</sup>H NMR, DEPTQ, HSQC, HMBC, <sup>1</sup>H-<sup>1</sup>H COSY, ROESY and HRESIMS data. The absolute configurations were assigned via comparison of the experimental and calculated ECD data. In addition, the isolated constituents as well as the known waltheriones M–Q were evaluated for their in vitro antitrypanosomal activity. Compounds **2**, **5**, **7** as well as waltheriones M, P, and Q showed potent growth inhibition towards *Trypanosoma cruzi* with IC<sub>50</sub> values of 2.1, 0.8, 2.1, 1.3, 0.5 and 0.1  $\mu$ M, respectively, and selectivity indices of >12, >33, >13, 5, 25, and 14. These findings further demonstrate that the waltheriones are a promising class of anti-chagasic compounds worthy of further investigations. Trypanosomiases are an important group of human and animal diseases caused by parasitic protozoa of the genus *Trypanosoma*. *Trypanosoma brucei rhodesiense* (*T. b. rhodesiense*) and *T. b. gambiense* cause human African trypanosomiasis, also known as sleeping sickness, in sub-Saharan Africa, whereas in Central and Latin America *T. cruzi* is responsible for American trypanosomiasis or Chagas disease. For both diseases, drugs exist but are not always effective and cause severe side effects. It is therefore urgent to find new and more potent molecules to cure these diseases, which affect mostly poor populations.

As part of our search for compounds with antitrypanosomal activity from *Waltheria indica*,<sup>1, 2</sup> alkaloids present in small amounts in the plant were targeted. Twelve new compounds, namely 7 quinoline-4(1*H*)-ones **1**, **2**, **8**–**12**, three 5,6,7,8-tetrahydroquinoline-4(1*H*)-ones **3**–**5**, and two pyridine-4(1*H*)-ones **6** and **7**, as well as melovinone (**13**) and 5′-methoxywaltherione A (**14**) were isolated from the CH<sub>2</sub>Cl<sub>2</sub> extract of the aerial parts of *W. indica*. Compounds **1**–**14** and five known waltheriones  $(M-Q)^3$  were screened for their in vitro growth inhibition of *T. cruzi*.













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# **RESULTS AND DISCUSSION**

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The aerial parts of W. indica were extracted using CH<sub>2</sub>Cl<sub>2</sub> at room temperature, and the resulting extract was fractionated by flash chromatography. Further purifications of the fractions were carried out by semi-preparative HPLC and yielded 12 new compounds

1–12, as well as melovinone (13) and 5'-methoxywaltherione A (14).

NMR data (Tables 1 and 2), 2D NMR experiments and comparison with the literature<sup>1,</sup>

<sup>3</sup> suggested pyridine-4(1*H*)-one-type structures for compounds 1-14.

Table 1. <sup>1</sup>H NMR Data of Compounds 1–12 (600 MHz, in Methanol-d<sub>4</sub>)

	1	2	3	4	5	6	7	8	9	10	11	12
position						$\delta_{ m H}$ (mult.	, J in Hz)					
5			2.97, m	2.95, m	2.85, m	6.27, s	6.27, s					
6	7.01, d (7.2)	6.92, d (8.1)	1.54, m	1.60, m 1.34, m	1.92, d (13.9) 1.67, m							
7	7.43, dd (8.4, 7.2)	7.02, d (8.1)	а	2.25, m	2.00, m 1.82, m	2.55, t (7.7)	2.54, t (7.7)	7.11, d (8.4)	7.42, m	7.15, d (8.7)	7.54, d (8.9)	6.88, d (8.9)
8	7.31, d (8.4)		2.76, m	а	4.54, dd (10.1, 6.2)	1.64, m	1.64, m	7.62, d (8.4)	7.46, d (8.4)	7.30, d (8.7)	7.39, d (8.9)	7.27, m
10								2.13, m	4.28, dd (10.0, 5.8)	6.61, t (7.3)	2.35, ddd (13.5, 7.4, 3.0) 2.21, ddd (13.5, 11.2, 3.0)	2.77, t (10.5) 1.28, m
11	3.36, m	3.27, m	1.33, m	1.29, m	1.33, m	1.30, m	1.33, m	1.66, m 1.00, m	1.88, m 0.74, m	1.89, m	1.97, m 1.84, m	2.47, m 1.65, m
12	1.62, m	1.63, m	1.33, m	1.29, m	1.33, m	1.30, m	1.33, m	2.05, m 1.86, m	2.11, m 1.93, m	2.41, m	5.43, t (4.9)	7.27, m
13	1.42, m	1.43, m	1.33, m	1.29, m	1.33, m	1.30, m	1.33, m	6.31, d (2.6)	6.20, d (3.9)	а	4.31, s	5.33, s
14	1.30, m	1.65, m	1.64, m	1.78, m	1.65, m	1.30, m	1.60, m					
15	1.30, m	2.58, t (7.3)	2.61, t (7.6)	4.60, t (6.6)	2.61, t (7.7)	1.30, m	2.59, t (8.1)					
16	1.28, m					1.31, m						
17	1.30, m	7.14, d (7.6)	7.16, d (7.5)	7.32, m	7.16, d (7.2)	1.29, m	7.14, m					
18	0.90, m	7.21, t (7.6)	7.23, t (7.5)	7.30, m	7.23, t (7.2)	0.90, t (7.0)	7.23, t (7.6)					
19		7.11, m	7.12, t (7.5)	7.23, t (7.0)	7.12, t (7.3)		7.13, m					
20		7.21, t (7.6)	7.23, t (7.5)	7.30, m	7.23, t (7.2)		7.23, t (7.6)					
21		7.14, d (7.6)	7.16, d (7.5)	7.32, m	7.16, d (7.2)		7.14, m					
2								7.56, d (8.4)	7.73, m	6.39, d (2.3)		
3´								7.41, t (8.4)	7.40, m		6.94, d (8.4)	7.12, d (8.3)
4′								7.33, t (7.3)	7.34, t (7.3)	6.42, t (2.3)	7.19, td (7.6, 2.3)	7.42, td (7.8, 3.9)

<i></i>								7.41, t	7.40		676	7.08, t
5								(8.4)	7.40, m		6./6, m	(7.8)
6´								7.56, dd (8.3, 1.3)	7.73, m	6.39, d (2.3)	6.74, m	7.42, td (7.8, 3.9)
MeO-1			4.12, s	3.99, s								
Me-2	2.44, s	2.48, s		2.46, s	2.34, s	2.33, s	2.32, s		2.50, s	2.47, s	2.51, s	2.47, s
CH <sub>2</sub> OH-2			4.76, s					4.86, s				
MeO-3	3.78, s	3.78, s	3.85, s	3.77, s	3.75, s	3.76, s	3.75, s	3.86, s	3.84, s	3.83, s	3.84, s	3.82, s
MeO-8		4.00, s										
MeO-13												3.23, s
MeO-2´											3.66, s	3.70, s
MeO-3'										3.73, s		
MeO-5′										3.73, s		
<i>a a</i> : 1.		,										

<sup>*a*</sup> Signal too weak to be observed.

	1	2	3	4	5	6	7	8	9	10	11	12
position						$\delta_{\mathrm{C}},$	type					
2	142.7, C	142.6, C	141.9, C	140.7, C	140.8, C	142.0, C	142.3, C	147.0, C	145.3, C	142.7, C	143.8, C	143.9, C
3	142.4, C	143.1, C	146.1, C	145.2, C	146.2, C	146.3, C	146.5, C	140.3, C	142.5, C	142.7, C	143.4, C	142.4, C
4	а	а	а	а	а	а	а	а	а	а	175.8, C	а
4a								121.3, C	121.5, C	124.3, C	124.7, C	120.5, C
5	145.9, C	136.5, C	32.6, CH	32.2, CH	32.5, CH	115.5, CH	115.7, CH	142.3, C	140.2, C	144.5, C	140.2, C	138.9, C
6	125.9, CH	125.2, CH	а	26.0, CH <sub>2</sub>	24.1, CH <sub>2</sub>	151.4, C	151.7, C	141.6, C	138.8, C	137.4, C	139.1, C	138.8, C
7	131.4, CH	110.4, CH	а	35.2, CH <sub>2</sub>	28.4, CH <sub>2</sub>	33.6, CH <sub>2</sub>	33.8, CH <sub>2</sub>	124.5, CH	127.5, CH	132.8, CH	134.4, CH	129.0, CH
8	116.7, CH	147.7, C	а	а	67.6, CH	30.0, CH <sub>2</sub>	30.3, CH <sub>2</sub>	118.8, CH	118.1, CH	116.0, CH	118.6, CH	116.9, CH
8a								139.6, C	143.9, CH	140.3, C	140.3, C	141.5, C
9	141.5, C	132.3, C	а	а	145.8, C	30.4, CH <sub>2</sub>	30.5, CH <sub>2</sub>	86.5, C	89.0, C	143.6, C	51.1, C	48.1, C
10	124.2, C	124.9, C	а	а	а	30.4, CH <sub>2</sub>	30.5, CH <sub>2</sub>	32.9, CH <sub>2</sub>	69.6, CH	130.7, CH	26.1, CH <sub>2</sub>	22.6, CH <sub>2</sub>
11	36.6, CH <sub>2</sub>	36.0, CH <sub>2</sub>	30.4, CH <sub>2</sub>	29.8, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.5, CH <sub>2</sub>	18.4, CH <sub>2</sub>	28.8, CH <sub>2</sub>	26.4, CH <sub>2</sub>	27.9, CH <sub>2</sub>	26.1, CH <sub>2</sub>
12	33.4, CH <sub>2</sub>	33.1, CH <sub>2</sub>	30.4, CH <sub>2</sub>	29.8, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.5, CH <sub>2</sub>	26.6, CH <sub>2</sub>	27.7, CH <sub>2</sub>	38.2, CH <sub>2</sub>	66.1, CH	67.6, CH
13	30.8, CH <sub>2</sub>	30.1, CH <sub>2</sub>	30.4, CH <sub>2</sub>	29.8, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.4, CH <sub>2</sub>	30.5, CH <sub>2</sub>	81.2, CH	80.7, CH	29.0, CH <sub>2</sub>	68.8, CH <sub>2</sub>	101.8, CH
14	30.3, CH <sub>2</sub>	32.7, CH <sub>2</sub>	32.6, CH <sub>2</sub>	39.9, CH <sub>2</sub>	32.8, CH <sub>2</sub>	30.4, CH <sub>2</sub>	32.8, CH <sub>2</sub>					
15	30.3, CH <sub>2</sub>	36.8, CH <sub>2</sub>	36.7, CH <sub>2</sub>	74.8, CH	36.9, CH <sub>2</sub>	30.4, CH <sub>2</sub>	36.9, CH <sub>2</sub>					
16	32.9, CH <sub>2</sub>	144.0, C	143.9, C	146.2, C	144.0, C	32.9, CH <sub>2</sub>	144.1, C					
17	23.5, CH <sub>2</sub>	129.1, CH	129.2, CH	126.8, CH	129.4, CH	23.5, CH <sub>2</sub>	129.4, CH					
18	14.2, CH <sub>3</sub>	129.0, CH	129.0, CH	128.9, CH	129.2, CH	14.2, CH <sub>3</sub>	129.2, CH					
19		126.3, CH	126.4, CH	127.8, CH	126.6, CH		126.6, CH					
20		129.0, CH	129.0, CH	128.9, CH	129.2, CH		129.2, CH					
21		129.1, CH	129.2, CH	126.8, CH	129.4, CH		129.4, CH					
1′								143.5, C	141.5, C	145.5, C	135.0, C	128.5, C
2								126.8, CH	129.2, CH	107.0, CH	159.2, C	159.2, C
3′								129.1, CH	129.0, CH	162.1, C	113.2, CH	112.5, CH
4´								128.6, CH	129.0, CH	99.9, CH	129.1, CH	130.1, CH
5´								129.1, CH	129.0, CH	162.1, C	121.1, CH	122.3, CH
6′								126.8, CH	129.2, CH	107.0, CH	132.3, CH	130.4, CH

Table 2. <sup>13</sup>C NMR Data of Compounds 1–12 (150 MHz, in Methanol-d<sub>4</sub>)

	1	2	3	4	5	6	7	8	9	10	11	12
position -						$\delta_{\mathrm{C}},$	type					
MeO-1			67.5, CH <sub>3</sub>	65.8, CH <sub>3</sub>								
Me-2	14.0, CH <sub>3</sub>	13.8, CH <sub>3</sub>		9.9, CH <sub>3</sub>	13.5, CH <sub>3</sub>	13.3, CH <sub>3</sub>	13.6, CH <sub>3</sub>		14.6, CH <sub>3</sub>	14.0, CH <sub>3</sub>	14.3, CH <sub>3</sub>	14.4, CH <sub>3</sub>
CH <sub>2</sub> OH-2			54.2, CH <sub>2</sub>					57.1, CH <sub>2</sub>				
MeO-3	60.0, CH <sub>3</sub>	60.0, CH <sub>3</sub>	61.0, CH <sub>3</sub>	59.8, CH <sub>3</sub>	60.1, CH <sub>3</sub>	59.9, CH <sub>3</sub>	60.1, CH <sub>3</sub>	60.2, CH <sub>3</sub>	60.4, CH <sub>3</sub>	60.1, CH <sub>3</sub>	60.4, CH <sub>3</sub>	60.4, CH <sub>3</sub>
MeO-8		56.3, CH <sub>3</sub>										
MeO-13												56.3, CH <sub>3</sub>
MeO-2											55.5, CH <sub>3</sub>	55.3, CH <sub>3</sub>
MeO-3										55.5, CH₃		
MeO-5′										55.5, CH₃		
<sup>a</sup> Signal too y	veak to be o	bserved.										

Compound **1** was obtained as a pale yellow oil with the molecular formula C<sub>19</sub>H<sub>27</sub>NO<sub>2</sub> established by HRESIMS data (*m/z* 302.2113 [M+H]<sup>+</sup>, calcd for C<sub>19</sub>H<sub>28</sub>NO<sub>2</sub>, 302.2115). The UV spectrum showed absorptions at 240 and 328 nm, suggesting a quinolinone skeleton similar to waltherione F.<sup>1</sup> The HSQC and HMBC spectra showed the presence of three sp<sup>2</sup> carbons linked to a heteroatom at & 142.7, 142.4, and 141.5 (C-2, C-3, and C-9, respectively), two quaternary aromatic carbons at & 145.9 and 124.2 (C-5 and C-10, respectively), three aromatic methines at & 125.9, 131.4 and 116.7 (C-6, C-7, and C-8, respectively), a methoxy group at & 60.0 (MeO-3), seven methylenes at & 36.6, 33.4, 30.8, 30.3, 30.3, 32.9 and 23.5 (C-11 – C-17, respectively), and two methyl groups at & 14.0 and 14.2 (Me-2 and C-18, respectively). <sup>1</sup>H-<sup>1</sup>H COSY correlations between H-6 (&<sub>H</sub> 7.01), H-7 (&<sub>H</sub> 7.43) and H-8 (&<sub>H</sub> 7.31), and the HMBC correlations from MeO-3 (&<sub>H</sub> 3.78) to C-3, Me-2 (&<sub>H</sub> 2.44) to C-2, H<sub>2</sub>-11 (&<sub>H</sub> 3.36) to C-5, C-6, and C-13, the *n*-

terminal methyl group H<sub>3</sub>-18 ( $\delta_{\rm H}$  0.90) to C-17 and C-16 led to the conclusion that **1** is 3methoxy-2-methyl-5-octylquinolin-4(1*H*)-one (Figure 1).



**Figure 1**. Key <sup>1</sup>H-<sup>1</sup>H COSY (bold red lines) and HMBC (blue arrows) correlations of **1** and **2**.

Compound **1** shows close similarities with waltherione F, previously isolated from the same plant, because both compounds have a benzene ring fused to the 4-pyridinone moiety. Only the presence of a methoxy group at C-8 distinguishes waltherione F from **1**. Therefore, the alkaloid (**1**) was trivially named 8-demethoxywaltherione F.

The HRESIMS data of compound **2** exhibited a protonated molecule  $[M+H]^+$  at m/z366.2064 (calcd for C<sub>23</sub>H<sub>28</sub>NO<sub>3</sub>, 366.2064), corresponding to the molecular formula C<sub>23</sub>H<sub>27</sub>NO<sub>3</sub>. When compared to **1**, the NMR data of **2** indicated five aromatic protons at  $\delta_H$  7.21 (2H, t, J = 7.6 Hz, H-18, 20), 7.14 (2H, d, J = 7.6 Hz, H-17, 21) and 7.11 (1H, m, H-19) in the terminal position of the *n*-pentyl chain, instead of the *n*-terminal methyl group H-18 signal in **1**. A second difference was the presence of an additional methoxy group at  $\delta_H$  4.00, which correlated with C-8 in the HMBC spectrum (Figure 1). The structure of waltherione R (**2**) was thus identified as 3,8-dimethoxy-2-methyl-5-(5phenylpentyl)quinolin-4(1*H*)-one. Compound **3** showed a protonated molecule  $[M+H]^+$  at m/z 386.2327 (calcd for C<sub>23</sub>H<sub>32</sub>NO<sub>4</sub>, 386.2326), indicating a molecular formula C<sub>23</sub>H<sub>31</sub>NO<sub>4</sub>. The structure of **3** is close to waltherione O<sup>3</sup> (Figure S1, Supporting Information). Indeed, both compounds shared a C-2 hydroxymethyl group at  $\delta_{\rm H}$  4.76, a cyclohexene ring fused to a 4-pyridinone moiety, a methine at  $\delta_{\rm H}$  2.97, and an *n*-pentyl side chain with a phenyl group in the terminal position. However, an additional methoxy group was detected at  $\delta_{\rm H}$  4.12 for **3**. The ROE association observed between CH<sub>2</sub>OH and both MeO-1 and MeO-3 were used to unambiguously position the methoxy group at the nitrogen atom. The absolute configuration at C-5 was established by comparison of the experimental and reported<sup>3</sup> ECD spectra. The ECD spectrum of **3** showed sequential negative and positive Cotton effects (CE) at 260 and 290 nm (Figure 2A), which was similar to the ECD spectrum of waltherione O, and the absolute configuration at C-5 was therefore defined as (*R*). Thus, the structure of 1-methoxywaltherione O (**3**) was defined as (*R*)-2-hydroxymethyl-1,3-dimethoxy-(5-phenylpentyl)-5,6,7,8-tetrahydroquinolin-4(1*H*)-one.



Figure 2. Experimental and TDDFT simulated ECD spectra of compounds 3–5, 8, 9, 11, and 12.

Compound **4** shares the same HRESIMS protonated molecule  $[M+H]^+$  at m/z 386.2324 than **3**. Instead of a C-2 hydroxymethyl group in **3**, the NMR data of **4** indicated a hydroxy group at C-15 at  $\delta_{\rm C}$  74.8. This site of hydroxylation was unambiguously located

by the HMBC correlation between the deshielded H-15 at  $\delta_{\rm H}$  4.60 and the aromatic carbons C-17/C-21 at  $\delta_{\rm C}$  126.8. The position of the hydroxy group was further confirmed by the ROE association observed between H-15 and H-17/H-21 at  $\delta_{\rm H}$  7.32. The absolute configuration was assigned as (5*S*,15*S*) based on the comparison of calculated and experimental ECD spectra (Figure 2B). In the latter, compound **4** showed two negative CEs around 290 and 200 nm along with three sequential positive CEs around 260, 240, and 210 nm. The positive CE observed at 259 nm implies a (5*S*) configuration.<sup>1</sup> This compound was characterized as (5*S*,15*S*)-(5-hydroxy-5-phenylpentyl)-1,3-dimethoxy-2methyl-5,6,7,8-tetrahydroquinolin-4(1*H*)-one and named (*S*)-15-hydroxywaltherione G.

The NMR data of compound **5** were similar to those of waltherione M<sup>3</sup> (Figure S1, Supporting Information). The only difference was the chemical shift of the methine C-8 at  $\delta_{\rm C}$  67.6, indicating the presence of an oxygen atom instead of a methylene at  $\delta_{\rm C}$  27.6 for waltherione M. This observation was in agreement with the molecular formula C<sub>22</sub>H<sub>29</sub>NO<sub>3</sub> deduced from the HRESIMS protonated molecule [M+H]<sup>+</sup> at *m*/*z* 356.2217 (calcd for C<sub>22</sub>H<sub>30</sub>NO<sub>3</sub>, 356.2225). The comparison of the literature data of waltheriones E, H, K, N, and Q<sup>1,3</sup> and the ECD spectrum of **5** (Figure 2A) exhibiting positive CEs at 230 and 262 nm and a negative CE at 285 nm revealed the (5*S*,8*R*) absolute configuration. This compound was identified as 8-hydroxy-3-methoxy-2-methyl-5-(5phenylpentyl)-5,6,7,8-tetrahydroquinolin-4(1*H*)-one and named (*R*)-8hydroxywaltherione M.

Compound **6** exhibited an HRESIMS protonated molecule  $[M+H]^+$  at m/z 308.2584 (calcd for C<sub>19</sub>H<sub>34</sub>NO<sub>2</sub>, 308.2584), indicating a molecular formula C<sub>19</sub>H<sub>33</sub>NO<sub>2</sub>. In comparison to 8-deoxoantidesmone (C<sub>19</sub>H<sub>31</sub>NO<sub>2</sub>) previously isolated from the same

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plant,<sup>1</sup> two additional indices of hydrogen deficiency were observed. The NMR data of **6** revealed a linear dodecyl aliphatic side chain at C-6 instead of a cyclohexene ring fused to a 4-pyridinone moiety for 8-deoxoantidesmone. Indeed, the <sup>1</sup>H and HSQC spectra exhibited an olefinic proton at  $\delta_{\rm H}$  6.27 (1H, s, H-5) correlating in HMBC with C-3, C-6, and C-7 at  $\delta_{\rm C}$  146.3, 151.4 and 33.6, respectively (Figure 3). Likewise, the methylene protons H<sub>2</sub>-7 at  $\delta_{\rm H}$  2.55 correlated with C-5 at  $\delta_{\rm C}$  115.5, C-6, and C-9 at  $\delta_{\rm C}$  30.4. A signal at  $\delta_{\rm H}$  0.90 (3H, t, *J* = 7.0 Hz, H-18) corresponding to the *n*-terminal methyl showed long-range correlation with C-16 and C-17 at  $\delta_{\rm C}$  32.9 and 23.5, respectively. Waltherione S (**6**) was thus identified as 6-dodecyl-3-methoxy-2-methylpyridin-4(1*H*)-one.



Figure 3. Key HMBC (blue arrows) correlations of 6 and 7.

Compound **7** showed NMR data close to those of waltherione S (**6**), except that in the terminal position of the *n*-alkyl side chain a phenyl group was present. HMBC correlations from the methylene protons H<sub>2</sub>-15 at  $\delta_{\rm H}$  2.59 (2H, t, J = 8.1 Hz) to C-16 and C-17/C-21 ( $\delta_{\rm C}$  144.1 and 129.4, respectively), and between H-17/H-21 at  $\delta_{\rm H}$  7.14 (2H, m) to C-15 ( $\delta_{\rm C}$  36.9) allowed unambiguous assignment of the  $\omega$ -position of the phenyl moiety (Figure 3). The integrations of the methylene groups in the <sup>1</sup>H NMR spectrum indicated a nonyl side chain, which was in agreement with the molecular formula

 $(C_{22}H_{31}NO_2)$  deduced from the HRESIMS protonated molecule  $[M+H]^+$  at *m/z* 342.2430 (calcd for  $C_{22}H_{32}NO_2$ , 342.2428). Therefore, Waltherione T (**7**) was identified as 3-methoxy-2-methyl-6-(9-phenylnonyl)pyridin-4(1*H*)-one.

Comparison of the observed and reported<sup>4</sup> NMR data of compound **8** indicated the presence of a 4-quinolinone moiety fused to a bicyclic ether with an attached phenyl ring as described for waltherione C. The chemical shifts at  $\delta_C$  86.5 and 81.2 of C-9 and C-13, respectively, suggested the presence of an ether bridge between these two carbons. However, no C-2 methyl group was observed in the HMBC experiment but instead hydroxymethyl protons at  $\delta_H$  4.86 correlated to C-2 and C-3 at  $\delta_C$  147.0 and 140.3, respectively. The molecular formula of C<sub>22</sub>H<sub>21</sub>NO<sub>4</sub> deduced from the HRESIMS ion at *m*/*z* 364.1543 [M+H]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>22</sub>NO<sub>4</sub>, 364.1544) confirmed the presence of an additional oxygen atom in comparison to waltherione C. Comparison of the TDDFT simulated spectra of the (9*S*,13*S*) and (9*R*,13*R*) enantiomers with the experimental ECD spectrum defined the absolute configuration as (9*S*,13*S*) (Figure 2C). These configurations are opposite to those described for waltherione C.<sup>4</sup> Consequently, compound **8** was identified as (9*S*,13*S*)-2-hydroxymethylwaltherione C.

Compound **9** is a regioisomer of **8** and shares the same HRESIMS protonated molecule  $[M+H]^+$  at m/z 364.1543 (calcd for C<sub>22</sub>H<sub>22</sub>NO<sub>4</sub>, 364.1544). The difference between these two compounds lies in the position of the hydroxy group. ROE correlations between the deshielded H-10 at  $\delta_{\rm H}$  4.28 (1H, dd, J = 10.0, 5.8 Hz), H<sub>eq</sub>-11 at  $\delta_{\rm H}$  1.88 (1H, m), H<sub>ax</sub>-11 at  $\delta_{\rm H}$  0.74 (1H, m) and H-2′/H-6′ at  $\delta_{\rm H}$  7.73 (2H, m) located the hydroxy group at C-10 ( $\delta_{\rm C}$  69.6) in compound **9**. Four stereoisomers are probable for **9**, their simulated ECD spectra are shown in Figure 2D. The two stereoisomers (9*S*,10*S*,13*S* 

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and 9*S*,10*R*,13*S*) revealed a better fit with experimental data, with some differences around 210 nm. Considering the ROE correlation between H-10 and H<sub>eq</sub>-12 at  $\delta_{\rm H}$  2.11 (1H, m) and the simulated ECD spectra, the absolute configurations of C-10 bearing the hydroxy group, as well as C-9 and C-13 were defined as (9*S*,10*S*,13*S*) (Figure 2D). Compound **9** was identified as (9*S*,10*S*,13*S*)-10-hydroxywaltherione C.

NMR data indicated that compound **10** was characterized by a 3-methoxy-2-methylquinoline-4(1*H*)-one skeleton with a cycloheptyl moiety attached to C-5/C-6 and a phenyl ring at C-9, as for compounds **8** and **9**. However, no oxygen atom was present on the cycloheptyl moiety but instead a double bond in C-9/C-10 as suggested by the chemical shifts at  $\delta_{\rm C}$  143.6 and 130.7, respectively. Three aromatic protons at  $\delta_{\rm H}$  6.39 (2H, d, J =2.3 Hz, H-2' and H-6'), and at  $\delta_{\rm H}$  6.42 (1H, t, J = 2.3 Hz, H-4') indicated that the phenyl ring is *m*-disubstituted. A signal at  $\delta_{\rm H}$  3.73 integrating for six protons (two methoxy groups) and HMBC correlations completed the elucidation of the structure. It was named waltherione U and is the 3',5'-dimethoxy derivative of melochinone, an alkaloid previously isolated from *Melochia tomentosa*.<sup>5</sup> Both *M. tomentosa* and *W. indica* are Malvaceae of the New World.

Compound **11** showed close structural similarities with compounds **8**–**10**, sharing a 3methoxy-2-methyl-quinolin-4(1*H*)-one nucleus and a phenyl ring at C-9. However, instead of a cycloheptyl moiety fused at C-5/C-6, a cyclohexyl carrying an oxymethano bridge was identified. Indeed, in the DEPTQ and HSQC spectra, two methylene groups CH<sub>2</sub>-10 and CH<sub>2</sub>-11 at  $\delta_{c}$  26.1 and 27.9, respectively, an oxygenated CH-12 at  $\delta_{c}$  66.1, and a quaternary carbon C-9 at  $\delta_{c}$  51.1 were detected. HMBC and <sup>1</sup>H-<sup>1</sup>H COSY correlations (Figure 4) showed that the cyclohexyl moiety was fused at C-5/C-6 to the

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quinoline-4(1H)-one nucleus and the phenyl ring linked at C-9. In addition, the HMBC correlations confirmed the location of the oxymethano group between C-9 and C-12. In particular, the signal of the methylene H<sub>2</sub>-13 at  $\delta_{\rm H}$  4.31 (2H, s) correlated with C-6, C-9, C-10, and C-1' at  $\delta_{\rm C}$  139.1, 51.1, 26.1, and 135.0, respectively. The relative configuration of 11 was inferred from ROESY data. A ROE correlation between H<sub>2</sub>-10 and H<sub>2</sub>-13 suggested that the tetrahydropyran ring from C-9 to C-13 adopted a boat conformation to which the aromatic ring of the quinolinone skeleton was fused diaxially (Figure 5). Additional ROE correlations between H-7, H<sub>2</sub>-13 and MeO-2<sup>'</sup> of the equatorial phenyl ring indicated a gauche conformation to one another, and placed these three elements in the same axial plane of the molecule. Moreover, MeO-2' at  $\delta_{\rm H}$  3.66 (3H, s) correlating with H-3' at  $\delta_{\rm H}$  6.94 (1H, d, J = 8.4 Hz) indicated that the phenyl ring was o-substituted. The absolute configurations of C-9 and C-12 were defined by ECD data as (9S) and (12R)(Figure 2E). This compound was defined as (9S,12R)-3-methoxy-9-(2'-methoxyphenyl)-2-methyl-9,10,11,12-tetrahydro-12,9-(oxymethano)benzo[f] quinolin-4(1H)-one and named waltherione V.



**Figure 4**. Key <sup>1</sup>H-<sup>1</sup>H COSY (bold red lines) and HMBC (blue arrows) correlations of **11** and **12**.

The NMR data of compound **12** were compared with those of **11** and a difference at C-13 was obvious. Instead of a methylene group as for **11**, a methine bearing two oxygen atoms was detected at  $\delta_{\rm C}$  101.8 in **12**. In the HMBC spectrum, a methoxy group at  $\delta_{\rm H}$  3.23 (3H, s) correlated with C-13 implying that this group is attached to C-13 (Figure 4). The HRESIMS protonated molecule [M+H]<sup>+</sup> at *m/z* 408.1807 (calcd for C<sub>24</sub>H<sub>26</sub>NO<sub>5</sub>, 408.1811) confirmed the presence of an additional methoxy group for **12** compared to **11**. ROE correlations between H-13, MeO-13 and MeO-2′, and the absence of correlation between H-10 and H-13 indicated that H-13 is equatorially oriented (Figure 5). This observation was confirmed by the (9*R*,12*S*,13*S*) absolute configurations determined by comparison of experimental and calculated ECD spectra (Figure 2F). Thus, (*S*)-13-methoxywaltherione V (**12**) was identified as (9*R*,12*S*,13*S*)-3,13-dimethoxy-9-(2′-methoxyphenyl)-2-methyl-9,10,11,12-tetrahydro-12,9-(oxymethano)benzo[*f*]quinolin-4(1*H*)-one and named.



Figure 5. Key ROE correlations (blue dashed arrows) of 11 and 12.

The molecular formula C<sub>19</sub>H<sub>29</sub>NO<sub>3</sub> suggested by HRESIMS and NMR data of compound **13** were in accordance with those of melovinone, an alkaloid previously isolated from *Melochia tomentosa*.<sup>6</sup> Melovinone (**13**) is closely related to waltherione R (**2**). The only difference is an additional methoxy group at C-7 in melovinone. The NMR data of compound **14** showed close similarities to those of waltherione A.<sup>7</sup> The only difference is a second methoxy group on the phenyl ring. Comparison of the observed and reported physical data<sup>8,9</sup> confirmed that alkaloid **14** is 5′methoxywaltherione A.

Compounds 1-14 as well as waltheriones M–Q (Figure S1, Supporting Information), previously isolated and characterized from the same plant<sup>3</sup> but not tested against the etiological agent of Chagas disease, were evaluated for their in vitro growth inhibition of the amastigote form of *T. cruzi* (Tulahuen C4 strain). The cytotoxicity towards uninfected host cells, i.e. rat skeletal L6 cells, was also assessed and the data are shown in Table 5.

Table 3. Antitrypanosomal and Cytotoxic Activities (IC<sub>50</sub> in  $\mu$ M) of Alkaloids 1–14 and Waltheriones M–Q

compound	T. cruzi	Cytotoxicity <sup>a</sup>	$\mathrm{SI}^b$
1	10.0 <sup>c</sup>	15.0	1.5
2	$2.1^{c}$	>26	>12.5
3	$3.9^{c}$	>26	>6
4	$7.5\pm0.5^d$	>26	>3.5
5	$0.8 \pm 0.1^d$	>28	>33
6	5.0 <sup>c</sup>	>32	>6
7	$2.1^{c}$	>29	>13
8	$14.7 \pm 0.6^d$	>22	>1.5
9	>20°	>22	>1
10	13.1 <sup>c</sup>	>25	>1.5
11	$19.0 \pm 2.1^d$	>26	>1
12	$18.7 \pm 1.4^d$	>24	>1
13	12.9 <sup>c</sup>	>23	>1.5

14	14.7 <sup>c</sup>	>25	>1
Waltherione M	$1.3 \pm 0.2^d$	6.5	5.0
Waltherione N	$6.5 \pm 0.8^d$	26.2	4.0
Waltherione O	$7.0 \pm 0.3^d$	>28	>4
Waltherione P	$0.5 \pm 0.1^d$	12.0	24.5
Waltherione Q	$0.1 \pm 0.0^d$	1.8	13.8
Benznidazole <sup>e</sup>	$2.9^{c}$	>345	>119
Podophyllotoxin <sup>e</sup>		0.02	

<sup>*a*</sup> Tested against rat skeletal L6 cells

<sup>*b*</sup> SI (selectivity index) =  $IC_{50}$  cytotoxicity/ $IC_{50}$  antitrypanosomal activity

<sup>c</sup> Results are the mean of two independent assays

<sup>*d*</sup> Results are the mean of three independent assays  $\pm$  SD

<sup>e</sup> Positive controls

Compounds 2, 5, 7, and waltheriones M, P, and Q showed IC<sub>50</sub> values lower than the reference drug benznidazole (IC<sub>50</sub> =  $2.9 \mu$ M). They carry an *n*-alkyl side chain (pentyl or nonyl for compound 7) with a terminal phenyl ring and a 3-methoxy-2-methylpyridin-4(1*H*)-one moiety. The (5*S*) absolute configuration of compounds 5, waltheriones M, P, and Q increased the activity against *T. cruzi*. This observation is in accordance with previous results.<sup>1</sup> Compounds 8–12 and 14 are waltheriones with cyclization of the aliphatic side chain between C-6 and C-9, which was detrimental for the antitrypanosomal activity. Thus, the best compounds had submicromolar IC<sub>50</sub> values against intracellular *T. cruzi* and selectivity indices above 10, underlining the potential of the waltheriones as anti-chagasic agents.

#### **EXPERIMENTAL SECTION**

**General Experimental Procedures.** Optical rotation was measured on a Jasco (Easton, MD, USA) P-1030 polarimeter (MeOH, *c* in g/100 mL). UV spectra were

recorded on a Perkin-Elmer Lambda-25 UV-vis spectrophotometer (Wellesley, MA, USA) in MeOH. ECD spectra were recorded on a JASCO J-815 CD spectrometer in MeOH. NMR spectra were recorded on a Bruker Avance III HD 600 MHz NMR spectrometer equipped with a QCI 5 mm Cryoprobe and a SampleJet automated sample changer (Bruker BioSpin, Rheinstetten, Germany). Chemical shifts are reported in parts per million ( $\delta$ ) using the residual methanol- $d_4$  signals ( $\delta_{\rm H}$  3.31;  $\delta_{\rm C}$  49.0) as internal standards for <sup>1</sup>H and <sup>13</sup>C NMR experiments, respectively. HRMS data were obtained on a Q Exactive Focus Hybrid quadrupole-orbitrap mass spectrometer (Thermo Scientific, Waltham, MA, USA) using electrospray ionization in the positive-ion mode. The spray voltage was set at 3.5 kV; the sheath gas flow rate  $(N_2)$  at 50 units; the capillary temperature was 320 °C; the S lens RF level at 50 and the probe heater temperature at 425 °C. UHPLC was performed on an Acquity UPLC I-class System (Waters, Milford, MA, USA). The separation was performed on an Acquity BEH C<sub>18</sub> UPLC column (150  $\times$ 2.1 mm i.d.; 1.7  $\mu$ m, Waters), using a gradient (MeCN and H<sub>2</sub>O both containing 0.1%) formic acid) from 5 to 98% MeCN in 30 min, followed by an isocratic washing step at 98% MeCN for 3 min. After the washing step, the column was re-equilibrated with 5% MeCN for 5 min prior to the next injection. The flow rate was set to 0.4 mL/min, the column temperature to 40 °C, and the injection volume was 1 µL. Fractionation was performed on an Armen Spot preparative chromatographic system (Interchim, Montluçon, France) equipped with a quaternary pump, a UV detector, and a fraction collector. Semi-preparative chromatography was performed on an Armen Spot System with a Kinetex Axia Core-Shell C18 column ( $250 \times 21.2$  mm, 5  $\mu$ m; Phenomenex, Torrance, CA, USA).

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ECD Computational Details. Conformational analysis of compounds 3–5, 8, 9, 11 and 12 was carried out using MacroModel 9.1 software (Schrödinger, LLC, New York, NY, USA) by applying the OPLS-2005 force field in H<sub>2</sub>O. The selected conformers were subjected to geometrical optimization using the density function theory (DFT) with the CAM-B3LYP functional and the 6-31G\*\* basis-set as implemented with the Gaussian 09 program package.<sup>10</sup> Vibrational analysis was performed at the same level to confirm stability of the minima. Time-dependent density function theory calculations at the TD-DFT/B3LYP/6-31G\*\* level in MeOH using the "self-consistent reaction field" method with the conductor-like polarizable calculation model were employed to calculate excitation energy (denoted by wavelength in nm) and rotatory strength in dipole velocity (*R<sub>vel</sub>*) and dipole length (*R<sub>len</sub>*) forms. ECD curves were calculated based on rotatory strengths using a half bandwidth of 0.3 eV using SpecDis version 1.61.<sup>11</sup>

**Plant Material.** The aerial parts of *W. indica* were collected between June 2012 and February 2013 in Zinder (Niger). The identification was confirmed by Didier Roguet (Geneva Botanical Gardens). A voucher specimen is deposited at the Geneva Botanical Gardens (no. 19003).

**Extraction and Isolation.** The CH<sub>2</sub>Cl<sub>2</sub> extract (1.2 g) of the aerial parts was fractionated by flash chromatography as previously described.<sup>3</sup> The flow rate was set to 25 mL/min and UV absorbance was at 220 nm. The separation yielded 200 fractions, which were individually analyzed by UHPLC-MS. Fractions 26-28, 31-32, 40-41, 52-54, 85-88, 112-114, 121-122, 135-138, 154-156, 159-160, and 176-178 were selected for further purification. The final fractionation step was performed by semi-preparative HPLC with a Kinetex Axia Core-Shell C<sub>18</sub> column (250 × 21.2 mm, 5  $\mu$ m; Phenomenex)

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using MeOH/H<sub>2</sub>O/0.1% formic acid as solvents for a gradient elution of 40 to 100% MeOH in 80 min. The flow rate was set to 18 mL/min and UV absorbance was at 220 nm. Fractions 26-28 (4.3 mg) gave **8** (0.5 mg), fractions 31-32 (20.1 mg) gave **14** (3.7 mg), fractions 40-41 (2.2 mg) gave **12** (0.4 mg), fractions 52-53 (3.0 mg) gave **11** (1.0 mg), fraction 54 (2.0 mg) gave **9** (0.6 mg), fractions 85-86 (3.5 mg) gave **5** (0.8 mg), fractions 87-88 (2.0 mg) gave **3** (0.3 mg), fractions 112-113 (2.9 mg) gave **7** (0.8 mg), fractions 113-114 (2.2 mg) gave **10** (0.5 mg), fractions 121-122 (2.0 mg) gave **4** (0.3 mg), fractions 135-138 (2.5 mg) gave **6** (0.4 mg), fractions 154-156 (1.9 mg) gave **2** (0.3 mg), fractions 159-160 (2.1 mg) gave **1** (0.5 mg), and fractions 176-178 (3.0 mg) gave **13** (0.4 mg).

8-Demethoxywaltherione F(I): pale yellow oil; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 220 (3.0), 240 (3.5), 328 (1.6) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 302.2113 [M+H]<sup>+</sup> (calcd for C<sub>19</sub>H<sub>28</sub>NO<sub>2</sub>, 302.2115).

*Waltherione R* (2): pale yellow oil; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 243 (3.4), 332 (2.6) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 366.2064 [M+H]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>28</sub>NO<sub>3</sub>, 366.2064).

*1-Methoxywaltherione O* (**3**): pale yellow oil;  $[a]_D^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 211 (3.4), 268 (2.0) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$  ( $\Delta \varepsilon$ ) 285 (+8.02) and 262 (-18.14) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m/z* 386.2327 [M+H]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>32</sub>NO<sub>4</sub>, 386.2326).

*15-Hydroxywaltherione G* (4): pale yellow oil;  $[a]_D^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 235 (3.8), 263 (3.5) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$ 

 $(\Delta \varepsilon)$  290 (-3.89), 259 (+14.73), 229 (+7.48), and 262 (-18.02) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m/z* 386.2324 [M+H]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>32</sub>NO<sub>4</sub>, 386.2326).

8-Hydroxywaltherione M (5): pale yellow oil;  $[a]_D^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 210 (3.6), 264 (1.6) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$ ( $\Delta \varepsilon$ ) 285 (-5.24),262 (+15.88) and 230 (+7.85) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 356.2223 [M+H]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>30</sub>NO<sub>3</sub>, 356.2225).

*Waltherione S* (6): pale yellow oil; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 212 (3.4), 224 (3.5), 264 (1.6) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m*/*z* 308.2584 [M+H]<sup>+</sup> (calcd for C<sub>19</sub>H<sub>34</sub>NO<sub>2</sub>, 308.2584).

*Waltherione T* (7): pale yellow oil; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 194 (4.0), 206 (3.7), 263 (2.0) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m*/*z* 342.2430 [M+H]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>32</sub>NO<sub>2</sub>, 342.2428).

2-Hydroxymethylwaltherione C (8): pale yellow oil;  $[a]_D^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 223 (3.8), 245 (3.8), 340 (1.6) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$  ( $\Delta \varepsilon$ ) 338 (-5.32), 295 (+4.77), 250 (+4.81), 232 (+12.29), and 213 (-18.04) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 364.1543 [M+H]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>22</sub>NO<sub>4</sub>, 364.1544).

*10-Hydroxywaltherione C* (**9**): pale yellow oil;  $[a]_D^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 220 (4.6), 262 (4.6) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$  ( $\Delta \varepsilon$ ) 334 (-9.07), 299 (+5.83), 256 (+4.04), 230 (+7.92), and 213 (-39.01) nm; IR not

measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 364.1543 [M+H]<sup>+</sup> (calcd for C<sub>22</sub>H<sub>22</sub>NO<sub>4</sub>, 364.1544).

*Waltherione U* (10): pale yellow oil; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 226 (3.8), 336 (1.8) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS m/z 392.1856 [M + H]<sup>+</sup> (calcd for C<sub>24</sub>H<sub>26</sub>NO<sub>4</sub>, 392.1857).

*Waltherione V* (11): pale yellow oil;  $[a]_D^{22}$  +9 (*c* 0.05, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 223 (3.0), 251 (3.9), 327 (2.4) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$  ( $\Delta \varepsilon$ ) 280 (-29.50), 261 (-17.81), 243 (+4.03), 236 (+58.01), and 223 (+24.80) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m/z* 378.1701 [M+H]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>24</sub>NO<sub>4</sub>, 378.1700).

*13-Methoxywaltherione V* (*12*): pale yellow oil;  $[a]_{D}^{22}$  not measured due to lack of material; UV (MeOH)  $\lambda_{max}$  (log  $\varepsilon$ ) 246 (3.6), 330 (1.9) nm; ECD (0.3 mM, MeOH)  $\lambda_{max}$  ( $\Delta\varepsilon$ ) 333 (-10.48), 295 (+22.31), 265 (-18.84), 233 (-43.03), 223 (-35.21), and 215 (+18.74) nm; IR not measured due to lack of material; <sup>1</sup>H and <sup>13</sup>C NMR, see Tables 1 and 2; HRESIMS *m/z* 408.1807 [M+H]<sup>+</sup> (calcd for C<sub>24</sub>H<sub>26</sub>NO<sub>5</sub>, 408.1806).

Antitrypanosomal and Cytotoxicity Assays. The in vitro activity against *T. cruzi* as well as cytotoxicity assessment in L6 cells were determined as reported elsewhere.<sup>12</sup> The strain was *T. cruzi*, Tulahuen C2C4 (LacZ), amastigote forms grown in rat L6 skeletal myoblasts. Results are expressed in  $\mu$ M for pure compounds.

## ASSOCIATED CONTENT

## **Supporting Information**

The Supporting Information is available free of charge at https://www.pubs.acs.org/doi/

Structures of the waltheriones M-Q

HRMS, MS/MS, UV, 1D and 2D NMR spectra for compounds 1–12 Calculation data for ECD curves for compounds 4, 8, 9, 11 and 12.

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## Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

We thank Romina Rocchetti, Sonja Märki and Monical Cal for expert technical assistance in drug testing, and François Brand for the picture used in the abstract graphic. This work was supported by the Swiss National Science Foundation (SNF grant CRSII5\_183536). The Institute of Pharmaceutical Sciences of Western Switzerland is thankful to the Swiss National Science Foundation for the support in the acquisition of the NMR 600 MHz (SNF R'Equip grant 316030\_164095).

#### REFERENCES

- Cretton, S.; Breant, L.; Pourrez, L.; Ambuehl, C.; Marcourt, L.; Ebrahimi, S. N.; Hamburger, M.; Perozzo, R.; Karimou, S.; Kaiser, M.; Cuendet, M.; Christen, P., J. *Nat. Prod.* 2014, 77, 2304-2311.
- Cretton, S.; Breant, L.; Pourrez, L.; Ambuehl, C.; Perozzo, R.; Marcourt, L.; Kaiser, M.; Cuendet, M.; Christen, P., *Fitoterapia* 2015, 105, 55-60.
- Cretton, S.; Dorsaz, S.; Azzollini, A.; Favre-Godal, Q.; Marcourt, L.; Ebrahimi, S. N.; Voinesco, F.; Michellod, E.; Sanglard, D.; Gindro, K.; Wolfender, J.-L.; Cuendet, M.; Christen, P., *J. Nat. Prod.* 2016, 79, 300-307.
- Jadulco, R. C.; Pond, C. D.; Van Wagoner, R. M.; Koch, M.; Gideon, O. G.; Matainaho, T. K.; Piskaut, P.; Barrows, L. R., *J. Nat. Prod.* 2014, 77, 183-187.
- Kapadia, G. J.; Paul, B. D.; Silverton, J. V.; Fales, H. M.; Sokoloski, E. A., J. Am. Chem. Soc. 1975, 97, 6814-9.
- Kapadia, G. J.; Shukla, Y. N.; Basak, S. P.; Fales, H. M.; Sokoloski, E. A., *Phytochemistry* 1978, 17, 1444-5.
- Hoelzel, S. C. S. M.; Vieira, E. R.; Giacomelli, S. R.; Dalcol, I. I.; Zanatta, N.; Morel,
   A. F., *Phytochemistry* 2005, 66, 1163-1167.
- Jang, J. Y.; Le Dang, Q.; Choi, Y. H.; Choi, G. J.; Jang, K. S.; Cha, B.; Luu, N. H.; Kim, J.-C., *J. Agric. Food Chem.* 2015, 63, 3803.
- Jang, J. Y.; Le Dang, Q.; Choi, Y. H.; Choi, G. J.; Jang, K. S.; Cha, B.; Luu, N. H.; Kim, J.-C., J. Agric. Food Chem. 2015, 63, 3803-3803.
- Frisch, M. J. T., G.W.; Schlegel, H.B.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.;
   Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G.A.; Nakatsuji, H.; Caricato, M.;

- Li, X.; Hratchian, H.P.; Izmaylov, A.F.; Bloino, J.; Zheng, G.; Sonnenberg, J.L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J.A.; Peralta, J.E.; Ogliaro, F.; Bearpark, M.; Heyd, J.J.; Brothers, E.; Kudin, K.N.; Staroverov, V.N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J.C.; Iyengar, S.S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J.M.; Klene, M.; Knox, J.E.; Cross, J.B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R.E.; Yazyev, O.; Austin, A.J.; Cammi, R.; Pomelli, C.; Ochterski, J.W.; Martin, R.L.; Morokuma, K.; Zakrzewski, V.G.; Voth, G.A.; Salvador, P.; Dannenberg, J.J.; Dapprich, S.; Daniels, A.D.; Farkas, O.; Foresman, J.B.; Ortiz, J.V.; Cioslowski, J.; Fox, D.J.; , *Gaussian 09, Revision A.02, Gaussian, Inc., Wallingford CT* **2009**.
- Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G., *Chirality* 2013, 25, 243-249.
- Bernal, F. A.; Kaiser, M.; Wünsch, B.; Schmidt, T. J., *ChemMedChem* 2020, 15, 68-78.

# Abstract graphic

