



Article
scientifique

Revue de la
littérature

2024

Published
version

Open
Access

This is the published version of the publication, made available in accordance with the publisher's policy.

Neuroprotective effects of lactate and ketone bodies in acute brain injury

Plourde, Guillaume; Roumes, Hélène; Suissa, Laurent; Hirt, Lorenz; Doche, Émilie; Pellerin, Luc; Bouzier-Sore, Anne-Karine; Quintard, Hervé

How to cite

PLOURDE, Guillaume et al. Neuroprotective effects of lactate and ketone bodies in acute brain injury. In: Journal of cerebral blood flow and metabolism, 2024, vol. 44, n° 7, p. 1078–1088. doi: 10.1177/0271678X241245486


This publication URL: <https://archive-ouverte.unige.ch/unige:190211>

Publication DOI: [10.1177/0271678X241245486](https://doi.org/10.1177/0271678X241245486)

© The author(s). This work is licensed under a Other Open Access license

<https://www.unige.ch/biblio/aou/fr/guide/info/references/licences/>

Neuroprotective effects of lactate and ketone bodies in acute brain injury

Guillaume Plourde¹ , H el ene Roumes², Laurent Suissa³, Lorenz Hirt⁴,  Emilie Doche³, Luc Pellerin^{5,*}, Anne-Karine Bouzier-Sore^{2,*} and Herv e Quintard⁶

Abstract

The goal of neurocritical care is to prevent and reverse the pathologic cascades of secondary brain injury by optimizing cerebral blood flow, oxygen supply and substrate delivery. While glucose is an essential energetic substrate for the brain, we frequently observe a strong decrease in glucose delivery and/or a glucose metabolic dysregulation following acute brain injury. In parallel, during the last decades, lactate and ketone bodies have been identified as potential alternative fuels to provide energy to the brain, both under physiological conditions and in case of glucose shortage. They are now viewed as integral parts of brain metabolism. In addition to their energetic role, experimental evidence also supports their neuroprotective properties after acute brain injury, regulating in particular intracranial pressure control, decreasing ischemic volume, and leading to an improvement in cognitive functions as well as survival. In this review, we present preclinical and clinical evidence exploring the mechanisms underlying their neuroprotective effects and identify research priorities for promoting lactate and ketone bodies use in brain injury.

Keywords

Alternative brain fuels, brain metabolism, ketone bodies, lactate, neuroprotection, stroke, TBI, hypoxia

Received 4 November 2023; Revised 4 March 2024; Accepted 14 March 2024

Introduction

Acute brain injury refers to brain damage that occurs suddenly, such as traumatic brain injury (TBI), stroke, subarachnoid hemorrhage (SAH) or global ischemia, for example. The insult itself leads to a primary injury, which is followed by secondary brain damage, a pathologic cascade that arises hours to days after the initial injury. The main mechanisms leading to development of secondary brain injury are cerebral blood flow disturbances, hypoxia and a strong decrease of glucose delivery to the brain.¹ To improve patient outcome, the goal of neurocritical care, after detection of the acute brain injury, is to avoid or reverse the underlying cascades that occur during this secondary brain injury period by optimizing cerebral blood flow, oxygen supply and substrate delivery to the brain.¹

Glucose is an essential substrate to satisfy brain energy requirements. However, its availability is frequently reduced following acute brain injury.^{2,3} In the last decades, lactate and ketone bodies have been identified as alternative fuels to provide energy to the brain and preserve glucose for other critical metabolic needs.

Results are also accumulating concerning their potential neuroprotective effects and clinical applications.

In this review, we present available data supporting a therapeutic benefit of lactate and ketone bodies in

¹Division of Intensive Care Medicine, Department of Medicine, Centre hospitalier de l'Universit e de Montr al, Montr al, Canada

²Centre de R esonance Magn etique des Syst emes Biologiques (CRMSB), Univ. Bordeaux, CNRS, CRMSB/UMR 5536, Bordeaux, France

³Neurovascular Unit, CHU de Marseille, Marseille, France

⁴Division of Neurology, Department of Clinical Neuroscience, Centre hospitalier universitaire vaudois, Lausanne, Suisse

⁵IRMETIST Inserm U1313, Universit e et CHU de Poitiers, Poitiers, France

⁶Division of Intensive Care Medicine, Department of Anesthesiology, Clinical Pharmacology, Intensive Care and Emergency Medicine, H opitaux universitaires de Gen ve, Gen ve, Suisse

*Equal contribution.

Corresponding author:

Guillaume Plourde, Division of Intensive Care Medicine, Department of Medicine, Centre hospitalier de l'Universit e de Montr al, 1051 rue Sanguinet, Montr al, Canada.

Email: guillaume.plourde.1@umontreal.ca

acute brain injury. We also review preclinical and clinical evidence exploring the mechanisms underlying their neuroprotective effects, and identify research priorities in the field of neuroenergetics.

Glucose for brain energy supply

While the brain represents only 2% of the total body mass, its glucose consumption is around 20% of the systemic consumption, which represents more than 10 times the predicted value relative to its mass. Glucose is an essential substrate to maintain normal brain metabolism and function. It is required for glycolysis, which in turn provides 2 ATP, and 2 pyruvate molecules that can enter the tricarboxylic acid (TCA) cycle to ensure energy production, in majority through the generation of reducing equivalents (NADH and $FADH_2$), which are then used in the electron transport chain to generate around 30 additional ATP molecules (the exact yield of ATP from one round of the TCA cycle can vary but it has been experimentally estimated that NADH can contribute about 2.5 to 3 ATP, and each $FADH_2$ can contribute about 1.5 to 2 ATP) (Figure 1). In addition to providing energy to the brain, glucose is a precursor in the synthesis of

neurotransmitters and participates in the pentose phosphate pathway (PPP), also known as the hexose monophosphate shunt. The PPP is a metabolic pathway that runs parallel to glycolysis and is essential for NADPH and pentose sugar production, as well as for redox homeostasis. Indeed, the PPP plays a crucial role in maintaining reduced glutathione (GSH) levels in cells by supplying the necessary reducing equivalents in the form of NADPH. Reduced glutathione acts as a critical antioxidant and contributes to the cellular defense mechanisms against oxidative stress due to reactive oxygen species (ROS) production. The PPP is therefore a major pathway for neuronal protection after an acute brain insult during which high amount of ROS are produced.

During the acute phase following brain injury, cerebral glucose availability may become insufficient to meet the metabolic demands due to cerebral blood flow failure, blood-brain barrier dysfunction or diffusion impairment.² Indeed, glucose storage is very low in the brain: if glycogen can be found in astrocytes, its consumption during ischemia can sustain brain functions for only 2 minutes (measurements made in rats⁴) In human, a decrease in glycemia linked with a decrease in cerebral glucose concentration was demonstrated to

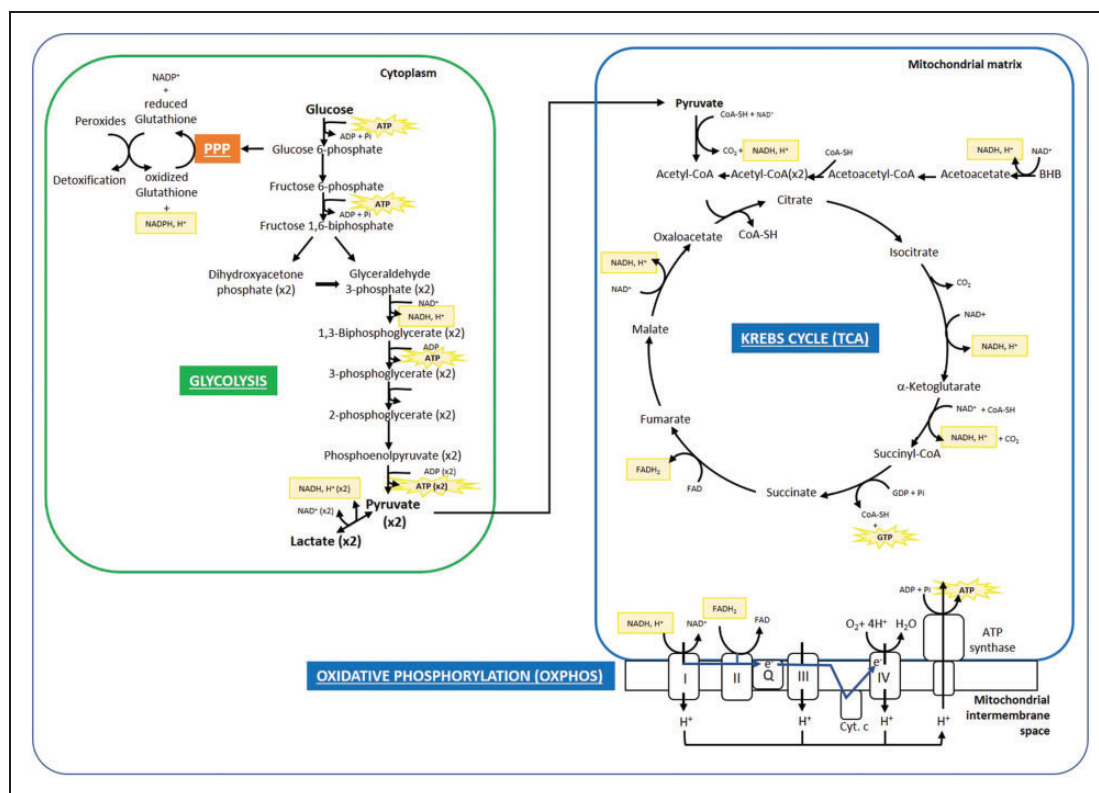


Figure 1. Glucose metabolism pathway. In the cytoplasm, glucose is metabolized to pyruvate through glycolysis. This pyruvate can either provide lactate or be redirected to the mitochondria to provide ATP through (i) the Krebs cycle (TCA cycle) coupled to and (ii) oxidative phosphorylation (OXPHOS).

be deleterious in the context of acute brain injury, leading to neuroglycopenia.^{5,6}

Glucose infusion following brain injury has been tested decades ago on a rat model of brain ischemia, aiming to reverse this substrate-deprived state and restore brain energy metabolism. Surprisingly, it was associated with increased mortality and extension of injury.⁷⁻⁹ Since then, these results have been consistently replicated in various experimental models of ischemic brain injury.¹⁰⁻¹³ In clinical setting, spontaneous high plasma glucose levels have also been associated with increased infarct size, worse clinical outcomes and increased mortality in ischemic stroke and SAH, especially when it happens before or early after ischemia.^{14,15}

With the same idea in mind to control glucose level, insulin therapy was tested, based on the early promising results of intensive insulin therapy in critically ill surgical patients.¹⁶ Many trials have investigated whether insulin-based glycemic control could improve the outcomes of brain-injured patients. Among these, the large GIST-UK trial failed to show a benefit of intensive insulin therapy in patients with acute stroke, although it was likely underpowered due to slow enrollment which justified its early termination.¹⁷ Overall, this trial and other studies pooled in two recent systematic reviews suggest that tight glycemic control causes more symptomatic and asymptomatic hypoglycemic episodes, both associated with poor outcomes.^{18,19}

These challenges in glucose availability, combined with frequent abnormal metabolic states following brain injury, impair its efficient incorporation into brain metabolism and explain the necessity of using alternative fuels to ensure adequate energy supply.

Ketone bodies and lactate as alternative fuels

It is well established that the brain is metabolically flexible and can metabolize other substrates, such as ketone bodies (KB) and lactate.^{20,21} KB are produced in the liver from fatty acids. They represent a major source of energy for the brain in neonates, who rely on breastfeeding for their diet which is rich in lipids. In adults, when glucose availability is limited in situations such as fasting, a very low-carbohydrate diet (ketogenic diet), or prolonged exercise, KB are produced as alternative fuels and used significantly by the brain.²² The three main KB synthesized by the liver are acetoacetate (AcAc), beta-hydroxybutyrate (BHB; AcAc can be converted to BHB, which is the predominant KB in circulation) and acetone, a minor KB that is produced as a byproduct. KB enter the brain via monocarboxylate transporters (MCTs) and provide energy to the

brain in the form of acetyl-CoA entering the tricarboxylic acid cycle to be oxidized, hence reducing reliance on glycolysis for energy production. This alternative energy production turns out to be highly efficient: BHB, one of the main KB, is converted to acetyl-CoA through only three enzymatic reactions, compared to the ten reactions with several rate-limiting enzymes required to convert glucose to acetyl-CoA. It has been shown in non-fasted adult humans infused with [2,4-¹³C₂]-BHB and using ¹H-¹³C polarization transfer spectroscopy that KB can cross the blood-brain barrier and are metabolized by the brain.²² These observations have been confirmed in pre-clinical studies.²³ In euglycemic conditions, KB are predominantly metabolized by neurons and account for 6% of total coenzyme A oxidation in normal conditions.²²

Concerning lactate, experiments suggest that it is commonly used for basal brain metabolism under physiological conditions.²⁰ Notably, Pellerin and Magistretti described a metabolic cooperation between astrocytes and neurons using lactate, termed the astrocyte-neuron lactate shuttle (ANLS).²⁴ In this model, glutamate released as part of synaptic activity is then taken up by astrocytes, one of their major function to avoid excitotoxicity. Concomitantly with glutamate uptake, a Na⁺ influx will occur and its intracellular concentration will rise. As a consequence, Na⁺/K⁺ ATPase activity will increase, leading to enhanced glucose utilization and lactate production by astrocytes. This astrocytic lactate is then transferred to neurons, through the MCTs (isoforms MCT1 and 4 for astrocytes, MCT2 for neurons; same transporters as the ones used for KB). Once in neurons, lactate is converted back to pyruvate and used as an energetic substrate. Since its description, the ANLS has gained popularity and has been validated in experimental models *in vitro*, *ex vivo* and *in vivo*, although remaining controversial.^{25,26} Using infusion of [3-¹³C]lactate and ¹³C magnetic resonance spectroscopy (MRS) it has been shown in rats that lactate efficiently crosses the blood-brain-barrier and then is preferentially a neuronal substrate.²⁷ These observations were confirmed in another study that showed that extracerebral lactate contributes at least two-fold more to replenish the neuronal than the glial pyruvate pools.²⁸ Authors concluded that utilization of exogenous lactate takes place primarily in neurons rather than in astrocytes.²⁷ Moreover, lactate entry into neurons through its transporter MCT2 was shown to be necessary to sustain brain function.²⁹⁻³¹ Under supraphysiological conditions, lactate can represent up to 60% of brain energy supply.

More recently, prevailing lactate uptake by the brain over glucose has been described in a healthy rat model³² and in exercising man,^{20,33,34} likely reflecting

an adaptation of the cerebral metabolic ratio to transient hyperlactatemia and relative glucose shortage. A similar observation has been made in patients with acute brain injury, in which an increase in lactate production occurred during spontaneous glucose depletion despite the absence of brain hypoxia.^{3,35,36} Finally, blocking lactate transport in a rat model of cerebral ischemia altered functional recovery, suggesting an essential role of lactate in basal neuroenergetics.³⁷ This obligatory role was further confirmed in another rat model of neonatal hypoxia-ischemia, in which reductions in brain lesion volume and ROS production were observed when lactate was injected, but not when it was co-administered with oxamate, a lactate dehydrogenase inhibitor therefore avoiding its conversion into pyruvate and its metabolism.³⁸ KB metabolism was also found to be amplified in a model of trauma.³⁹ In a juvenile rat model of TBI infused with [2,4-¹³C₂]-BHB, it has been shown using ¹³C MRS that KB were extensively metabolized, mainly in neurons.³⁹

Thus both KB and lactate can serve as alternative oxidative substrates for the brain and are mainly metabolized by neurons. They both enter through MCTs and deliver energy after their oxidation in the TCA cycle, entering either at the acetylCoA or at the pyruvate level, respectively (Figure 2). In both cases, their metabolism also lead to the conversion of cytosolic NAD⁺ into NADH and thus modify the redox potential of the cell (during the conversion of BHB into AcAc or lactate into pyruvate).

Based on such preclinical evidence, the idea of providing fuel support to brain-injured patients progressively emerged and a few clinical trials started. Bouzat *et al.* were the first to infuse hypertonic lactate in patients with severe TBI undergoing cerebral microdialysis (CMD) monitoring. They observed a significant increase in extracellular cerebral glucose.⁴⁰ Hypertonic lactate therapy was also shown to improve neuroenergetics (CMD glucose concentration) in TBI patients monitored with CMD.⁴¹ Overall, these data suggest that both endogenous and exogenous lactate are associated with an increased cerebral glucose availability. This “excess” glucose may then be redirected for other metabolic purposes, such as the neuronal PPP.⁴²

Taken altogether, challenges in glucose availability after an acute brain injury, combined with the possibility of the brain to metabolize efficiently KB and lactate, offer new perspectives. A direct modulation of neuronal metabolism by these alternative energy substrates could be of major interest in the search for therapeutic avenues in the context of acute brain injury.

Alternative fuels and the pentose-phosphate pathway

Neuronal glycolysis is controlled by the enzymatic complex APC/C-Cdh1 (anaphase-promoting complex/cyclosome-cdc20 homolog 1) that degrades the proglycolytic enzyme 6-phosphofructo-2-kinase/fructose-2,6-bisphosphatase-3 (PFKFB3). The enzyme PFKFB3 is less abundant in neurons than in astrocytes, resulting in a lower glycolytic rate and in the redirection of glucose-6-phosphate through the PPP.^{43,44} Astrocytes, on the other hand, exhibit a greater PFKFB3 activity, resulting in higher glycolytic rate and lactate production. Hence, this compartmentalization suggests that neurons use glucose preferentially for regulation of their redox state rather than for energy requirements.⁴⁵ In stressed states, such after an oxygen-glucose deprived condition, the reperfused neurons showed elevated PFKFB3 expression.⁴⁶ Such changes directed neuronal glucose metabolism from PPP to glycolysis compared to normal neurons, resulting in increased ROS production and apoptosis during reperfusion. Interestingly, both KB and lactate are oxidative substrates and provide acetylCoA or pyruvate, respectively, while contributing to the conversion of cytosolic NAD⁺ into NADH. An increase in the NAD⁺/NADH ratio will inhibit glycolysis and redirect glucose through the PPP, and therefore increase the pool of reduced glutathione. Indeed, it has been shown that both alternative fuels can decrease ROS damages,^{38,47–50} whereas the activity of G6PD, the key enzyme of the first (and irreversible) step of the PPP, was significantly increased when physiological concentrations of L-lactate were applied on neuronal cultures.⁵¹

Lactate and ketone bodies to control neuroinflammation

Emerging data suggest that lactate and KB have anti-inflammatory properties, but the underlying mechanisms remain unclear. Microglia are key regulators of brain homeostasis and act as Janus-faced cells regarding neuroinflammation.⁵² After an ischemic event, microglia are activated and release pro-inflammatory factors, such as TNF- α , IL-1 β , IL-6 and IFN- γ , exhibiting a pro-inflammatory or “M1” microglial phenotype.⁵³ On the other hand, microglia can also promote tissue and vascular remodeling via expression of anti-inflammatory factors such as IL-10, VEGF, TGF- β and BDNF, exhibiting an anti-inflammatory or “M2” phenotype. These microglial phenotypes are tightly linked to their metabolism, and lactate was shown to switch microglial profiles from a glycolytic M1 to an oxidative M2 phenotype, hence promoting microglial energy production.⁵⁴ Lactate has been

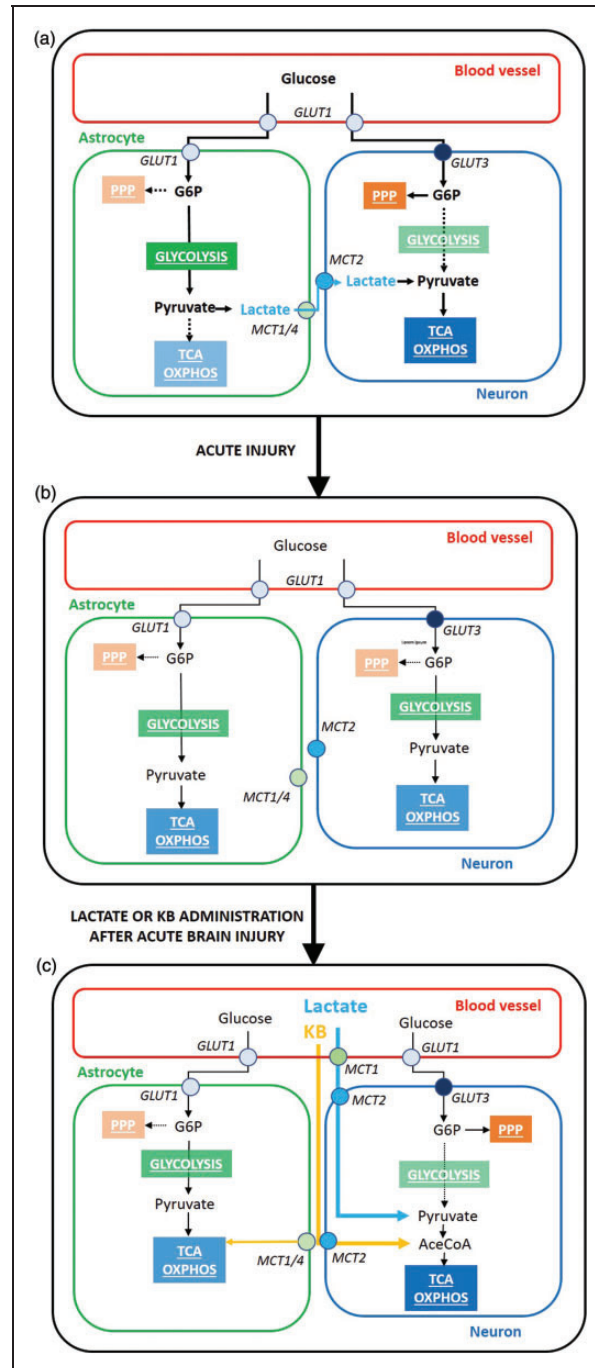


Figure 2. Compartmentalization of cerebral glucose metabolism and consequence in brain injury as well as for neuroprotective strategies. (a) In the brain, glucose from the bloodstream enters both astrocytes and neurons. It is metabolized to pyruvate in the cytoplasm (glycolysis) then supplies ATP via pyruvate oxidation into mitochondria (TCA cycle + OXPHOS). However, astrocytes have a greater glycolytic capacity compared to neurons, which are highly oxidative cells.³⁶ Astrocytes are known to produce high levels of lactate, which can be further transferred to neurons where it can be used as an energy substrate through the TCA and OXPHOS. Glucose can also enter directly into neurons. Low glycolysis in neurons allows glucose conversion into the pentose phosphate pathway PPP to sustain antioxidant protection. (b) In acute brain injury condition, the low level of circulating glucose can not meet cerebral energy needs and the metabolic cooperation between astrocytes and neurons can not be maintained. Glucose is mainly used for oxidative metabolism in both cell types to maintain ATP levels. (c) In order to overcome the energy deficit induced by acute brain injury, energy substitution strategies could be adopted. Lactate or exogenous KB enter directly into brain cells but predominantly in neurons.^{22,28,39} Lactate is then converted into pyruvate, while KB are precursors of acetyl-CoA. Pyruvate and Acetyl-CoA are then oxidized through TCA cycle and OXPHOS to provide energy. Saved glucose can be redirected to the PPP pathway to reestablish antioxidant defenses. GLUT: glucose transporter; KB: ketone bodies; MCT: monocarboxylate transporter; OXPHOS: oxidative phosphorylation; PPP: pentose phosphate pathway; TCA: tricarboxylic acid cycles.

shown to inhibit microglia-mediated neuroinflammation through the Akt-pathway leading to elongation of microglial processes.⁵⁵ BHB was shown to exert similar effects through induction of microglial ramifications and promoting an anti-inflammatory response in a Akt-dependant manner.⁵⁶ There is also evidence that a ketogenic diet reduces ROS generation and suppresses inflammasome activation.⁵⁷ In another study, ketogenic diet and BHB post-treatment reduced infarct volume and suppressed the over-activation of microglia in a mouse model of middle cerebral artery occlusion (MCAO).⁵⁸ Together with glucose redirection through the PPP, these anti-inflammatory effects might constitute an elegant hypothesis to explain some of the clinical benefits observed with alternative fuel supplementation in acute brain injury.

Increase in cerebral blood flow

In an *in vitro* study on rat hippocampal and neocortical slices, cerebral vasomotor tone was influenced by the metabolic state of the tissue. In hypoxic conditions, astrocyte glycolysis and lactate production were increased, causing vasodilation via an attenuation of prostaglandin uptake.^{59,60} In a cohort of patients with severe acute brain injury (SAH and TBI), a lactate infusion increased extracellular glucose levels and cerebral blood flow estimated by transcranial Doppler.⁶¹ Explanation of this mechanism is not well understood. Lactate act as a transmitter of metabolic information by modulating prostaglandin action and cerebral vasodilatation causing cerebral blood flow to increase, regulates the NAD⁺/NADH redox ratio by conversion to pyruvate, and/or activates a G-protein coupled receptor in neurons, astrocytes, and capillaries.^{60,62,63} Interestingly, KB can also increase cerebral blood flow as shown in mice by MRI,⁶⁴ and in human healthy subjects by PET scan⁶⁵ and the Kety-Schmidt technique.⁶⁶

Control of brain edema

Intracranial hypertension (sustained and significant elevation of intracranial pressure) is one of the major challenges faced by neurocritical care clinicians following TBI, massive SAH or hypoxic-ischemic brain injury. To control intracranial pressure (ICP), conventional treatment is based on a stepwise approach mostly described in TBI and involves deep sedation and analgesia, correct positioning of the patient, and hypertonic sodium chloride or mannitol infusions.⁶⁷ If the efficacy of osmotic therapy to control ICP is well established, it is also associated with side effects, among which are hyperchloremic acidosis and fluid overload.⁶⁸ Based on its favorable characteristics (naturally present molecule and no additional chloride),

sodium lactate has been identified as a potential agent to control ICP while avoiding these deleterious effects. It was first tested in a randomized trial in humans against mannitol in the treatment of intracranial hypertension during severe TBI. In this study, lactate was associated with a greater and more prolonged decrease in ICP than mannitol.⁶⁹ This result has since been replicated, and was also shown to prevent intracranial hypertensive episodes and fluid overload.^{40,70,71} This control of brain edema might explain why lactate has been associated with increased tissue oxygenation. In rat models of TBI, exogenous sodium lactate increased cerebral tissue oxygen pressure, possibly via improvement of oxygen diffusion due to reduced tissue edema.^{72,73} The antiedematous effect of sodium lactate might be induced by an increased expression of VRAC, KCC2 and AQP4, channels involved in cell volume regulation and free water movements across membranes, rather than by an osmotic effect.⁷² Concerning KB, their antiedematous effects were examined in rats with transient occlusion of middle cerebral artery. The administration of BHB (30 mg. kg⁻¹.h⁻¹) significantly reduced cerebral infarct area and edema formation.⁷⁴

Prevention of neuronal damage

As described just before, KB have been associated with reductions in lesion volume after middle cerebral artery occlusion,^{58,74} and also after TBI.^{75,76} *In vitro* experiments on cultured cortical neurons showed that a BHB-based treatment prevents neuronal death induced by glucose deprivation or hypoxia by stimulation of the autophagic flux thus preventing autophagosome accumulation and improving neuronal survival.⁷⁷ Moreover, both lactate and KB can limit excitotoxicity, reducing neuronal death as well as lesion volume. In a glutamate excitotoxicity context, pyruvate coming from lactate is metabolized to produce ATP. This ATP in turn activates the PI3K pathway leading to KATP channels opening. This results in hyperpolarization of the neuron, decreasing neuronal excitability and leading to a neuroprotective effect.⁷⁸ The neuroprotective effect of KB, targeting glutamate excitotoxicity, has also been shown in rat neocortical neurons.⁴⁸ In this deleterious context, KB can compete with the Cl⁻ anion and inactivate the vesicular glutamate transporters (VGLUTs), decreasing presynaptic glutamate release.⁷⁹ A summary of *in vivo* studies showing a reduction of the infarct volume after KB administration can be found in Table 2 of the following review.⁸⁰ Lactate was also shown to decrease brain lesion volume in several rat models after acute brain injuries such as neonatal hypoxia-ischemia,^{38,81} TBI and stroke.^{38,81-85}

Improvement of recovery and survival

Both substrates have been associated with improvement of cognitive function. In a rat model of neonatal hypoxia-ischemia, lactate administration was associated with short and long term improvement of motor and cognitive functions.³⁸ Improvement of long term cognitive functions was also observed after lactate injections in rat models of TBI or stroke.⁸⁶⁻⁸⁸ A ketogenic diet or BHB administered post-treatment improved neurological function in a mouse model of MCAO.⁵⁸ Finally, in a recent meta-analysis of 49 studies on murine models of TBI, ketones were associated with an improved survival, particularly in the adult subgroup.⁸⁹ Interestingly, the plasma level of ketosis was the only predictor of the neuroprotective effect. Another meta-analysis of studies on animal models of stroke described a protective effect of ketosis on outcome⁹⁰ while a recent review summarized the different studies showing the neuroprotective effect of KB in acute brain injuries (different models, different species, different ages).⁸⁰

Lactate and ketone bodies as pharmacological agents for acute brain injury

Lactate can be administered either as an intravenous or an intracerebral solution of sodium lactate.⁹¹ Most interventional studies on TBI used intravenous solutions with an osmolality ranging from 500 mmol/L to 1000 mmol/L, the latter being approximately iso-osmolar to 20% mannitol.^{40,41,61,69,92,93} In two studies, investigators injected sodium lactate directly into the brain using a microdialysis technique with 4 and 8 mmol/L microdialysis solutions.^{42,94} Both methods of lactate administration aimed to improve brain function and reduce damage associated with head trauma. Currently, there is no clear consensus on the optimal method of administration of lactate.

Ketone bodies can be administered through various formulations. Ketogenic diets are based on carbohydrate deprivation, either by fasting or by enteral formulas containing medium-chain triglycerides and low amounts of carbohydrates. An exclusive ketogenic

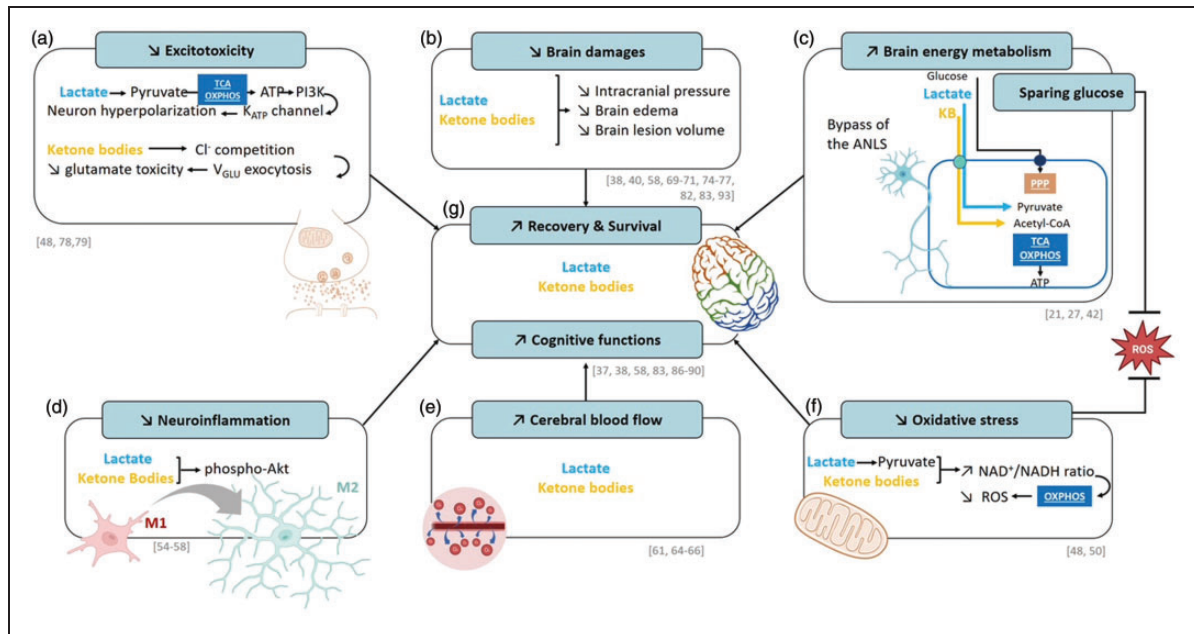


Figure 3. Neuroprotection by exogenous lactate and ketone bodies, as alternative energy substrates, in a context of brain injury. (a) Lactate counteracts glutamate excitotoxicity through the PI3K pathway leading to neuronal hyperpolarization. KB also decrease excitotoxicity through a competition with Cl⁻, responsible of a decrease in vGLU exocytosis. (b) Both exogenous lactate and KB limit brain damages by decreasing intracranial pressure, brain edema as well as brain lesion volume. (c) Lactate and KB increase brain energy metabolism by providing respectively pyruvate and acetyl-CoA to feed the TCA cycle and OXPHOS while preserving glucose for the PPP pathway, helping to fight ROS. (d) Both alternative substrates also switch microglia from a pro-inflammatory M1 to an anti-inflammatory M2 phenotype, through a stimulation of the Akt pathway. (e) Lactate and KB increase cerebral blood flow. (f) Both lactate and KB regulate the oxidative stress by increasing the NAD⁺/NADH ratio. (g) Altogether, beneficial effects of exogenous lactate and KB lead to increase recovery and survival, as well as improvement of cognitive functions. ANLS: astrocyte-neurone lactate shuttle; KB: ketone bodies; OXPHOS: oxidative phosphorylation; PPP: pentose phosphate pathway; ROS: reactive oxygen species; TCA: tricarboxylic acid; vGLU: vesicular glutamate transporters. Related reference numbers are shown in brackets [].

diet such as used to treat epilepsy in children takes 3 to 5 days to reach therapeutic blood levels. These delays render the ketogenic diet of limited interest in acute neurocritical care. Moreover, ketogenic diets are difficult to prepare and compliance is a challenge.⁹⁵ KB may also be infused as intravenous sodium BHB or sodium acetoacetate solutions. Finally, enteral KB are now available in the form of ketone esters (such as 1,3-butanediol monoester of beta-hydroxybutyrate or glyceryl-tris-3-hydroxybutyrate, two precursors of KB). The compound (R)-3-hydroxybutyl (R)-3-hydroxybutyrate is now considered a safe food supplement for humans by the U.S. Food and Drug Administration. Ketone esters induce a rapid and significant ketosis after a single oral intake and as such, represent an interesting therapeutic avenue in acute care.^{21,95}

Conclusion and perspectives

After being considered as unwanted metabolic waste, lactate and KB are now widely viewed as integral parts of brain metabolism. Data accumulating over the last thirty years support their important role in energy production during rest states, as well as alternative substrates in case of glucose shortage or unavailability. They have also been tested as therapeutics in animal models and human cohorts with acute brain injury exhibiting beneficial results concerning excitotoxicity, neuroinflammation, cerebral blood flow control, reduction of brain lesion volume, improvement of cognitive function and survival (Figure 3). KB and lactate share a common mechanism: modulation of brain metabolism. Indeed, when administered, lactate and KB exert a glucose-sparing effect leading to neuronal glycolytic regulation and redirection of glucose through the PPP which helps to fight against ROS produced during oxidative stress (Figure 3). Overall, data in animal models and small human cohorts call for a confirmation of their benefits in large therapeutic trials targeting acute neurocritical care populations. After decades of preclinical research and encouraging new clinical data, we are at the dawn of a new period during which lactate and ketone bodies might become an integral part of neurocritical care therapeutics.

Funding

LH received financial support from FNS 310030_212233 and Donation Eva Zurbrugg.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

ORCID iD

Guillaume Plourde  <https://orcid.org/0000-0002-3641-6281>

References

1. Bouzat P, Sala N, Payen JF, et al. Beyond intracranial pressure: optimization of cerebral blood flow, oxygen, and substrate delivery after traumatic brain injury. *Ann Intensive Care* 2013; 3: 23.
2. Hermanides J, Hong YT, Trivedi M, et al. Metabolic derangements are associated with impaired glucose delivery following traumatic brain injury. *Brain J Brain* 2021; 144: 3492–3504.
3. Patet C, Quintard H, Suys T, et al. Neuroenergetic response to prolonged cerebral glucose depletion after severe brain injury and the role of lactate. *J Neurotrauma* 2015; 32: 1560–1566.
4. Swanson RA, Sagar SM and Sharp FR. Regional brain glycogen stores and metabolism during complete global ischaemia. *Neurol Res* 1989; 11: 24–28.
5. Vespa P, McArthur DL, Stein N, et al. Tight glycemic control increases metabolic distress in traumatic brain injury: a randomized controlled within-subjects trial. *Crit Care Med* 2012; 40: 1923–1929.
6. Magnoni S, Tedesco C, Carbonara M, et al. Relationship between systemic glucose and cerebral glucose is preserved in patients with severe traumatic brain injury, but glucose delivery to the brain may become limited when oxidative metabolism is impaired: implications for glycemic control. *Crit Care Med* 2012; 40: 1785–1791.
7. Pulsinelli WA, Waldman S, Rawlinson D, et al. Moderate hyperglycemia augments ischemic brain damage: a neuropathologic study in the rat. *Neurology* 1982; 32: 1239–1246.
8. Pulsinelli W, Waldman S, Sigsbee B, et al. Experimental hyperglycemia and diabetes mellitus worsen stroke outcome. *Trans Am Neurol Assoc* 1980; 105: 21–24.
9. Siemkiewicz E and Hansen AJ. Brain extracellular ion composition and EEG activity following 10 minutes ischemia in normo- and hyperglycemic rats. *Stroke* 1981; 12: 236–240.
10. Li PA, Kristián T, Shamloo M, et al. Effects of preischemic hyperglycemia on brain damage incurred by rats subjected to 2.5 or 5 minutes of forebrain ischemia. *Stroke* 1996; 27: 1592–1602.
11. Gisselsson L, Smith ML and Siesjö BK. Hyperglycemia and focal brain ischemia. *J Cereb Blood Flow Metab off J Tab* 1999; 19: 288–297.
12. Robbins NM and Swanson RA. Opposing effects of glucose on stroke and reperfusion injury: acidosis, oxidative stress, and energy metabolism. *Stroke* 2014; 45: 1881–1886.
13. MacDougall NJJ and Muir KW. Hyperglycaemia and infarct size in animal models of middle cerebral artery occlusion: systematic review and meta-analysis. *J Cereb Blood Flow Metab off J Tab* 2011; 31: 807–818.
14. Pulsinelli WA, Levy DE, Sigsbee B, et al. Increased damage after ischemic stroke in patients with hyperglycemia with or without established diabetes mellitus. *Am J Med* 1983; 74: 540–544.

15. Badjatia N, Topcuoglu MA, Buonanno FS, et al. Relationship between hyperglycemia and symptomatic vasospasm after subarachnoid hemorrhage. *Crit Care Med* 2005; 33: 1603–1609; quiz 1623.
16. van den Berghe G, Wouters P, Weekers F, et al. Intensive insulin therapy in critically ill patients. *N Engl J Med* 2001; 345: 1359–1367.
17. Gray CS, Hildreth AJ, Sandercock PA, et al. Glucose-potassium-insulin infusions in the management of post-stroke hyperglycaemia: the UK glucose insulin in stroke trial (GIST-UK). *Lancet Neurol* 2007; 6: 397–406.
18. Bellolio MF, Gilmore RM and Ganti L. Insulin for glycaemic control in acute ischaemic stroke. *Cochrane Database Syst Rev* 2014; 2014: CD005346.
19. Hermanides J, Plummer MP, Finnis M, et al. Glycaemic control targets after traumatic brain injury: a systematic review and meta-analysis. *Crit Care Lond Care* 2018; 22: 11.
20. Boumezbeur F, Petersen KF, Cline GW, et al. The contribution of blood lactate to brain energy metabolism in humans measured by dynamic ¹³C nuclear magnetic resonance spectroscopy. *J Neurosci off J Urosci* 2010; 30: 13983–13991.
21. Suissa L, Kotchetkov P, Guignonis J-M, et al. Ingested ketone ester leads to a rapid rise of acetyl-CoA and competes with glucose metabolism in the brain of non-fasted mice. *Int J Mol Sci* 2021; 22: 524.
22. Pan JW, de Graaf RA, Petersen KF, et al. [2,4-¹³C]-beta-hydroxybutyrate metabolism in human brain. *J Cereb Blood Flow Metab off J Tab* 2002; 22: 890–898.
23. Roy M, Beauvieux M-C, Naulin J, et al. Rapid adaptation of rat brain and liver metabolism to a ketogenic diet: an integrated study using (1)H- and (13)C-NMR spectroscopy. *J Cereb Blood Flow Metab off J Tab* 2015; 35: 1154–1162.
24. Pellerin L and Magistretti PJ. Glutamate uptake into astrocytes stimulates aerobic glycolysis: a mechanism coupling neuronal activity to glucose utilization. *Proc Natl Acad Sci U S A* 1994; 91: 10625–10629.
25. Bak LK and Walls AB. CrossTalk opposing view: lack of evidence supporting an astrocyte-to-neuron lactate shuttle coupling neuronal activity to glucose utilisation in the brain. *J Physiol* 2018; 596: 351–353.
26. Barros LF and Weber B. CrossTalk proposal: an important astrocyte-to-neuron lactate shuttle couples neuronal activity to glucose utilisation in the brain. *J Physiol* 2018; 596: 347–350.
27. Bouzier AK, Thiaudiere E, Biran M, et al. The metabolism of [3-¹³C]lactate in the rat brain is specific of a pyruvate carboxylase-deprived compartment. *J Neurochem* 2000; 75: 480–486.
28. Duarte JMN, Girault FM and Gruetter R. Brain energy metabolism measured by (13)C magnetic resonance spectroscopy in vivo upon infusion of [3-(13)C]lactate. *J Neurosci Res* 2015; 93: 1009–1018.
29. Suzuki A, Stern SA, Bozdagi O, et al. Astrocyte-neuron lactate transport is required for long-term memory formation. *Cell* 2011; 144: 810–823.
30. Netzahualcoyotzi C and Pellerin L. Neuronal and astroglial monocarboxylate transporters play key but distinct roles in hippocampus-dependent learning and memory formation. *Prog Neurobiol* 2020; 194: 101888.
31. Roumes H, Jollé C, Blanc J, et al. Lactate transporters in the rat barrel cortex sustain whisker-dependent BOLD fMRI signal and behavioral performance. *Proc Natl Acad Sci* 2021; 118: e2112466118.
32. Wyss MT, Jolivet R, Buck A, et al. In vivo evidence for lactate as a neuronal energy source. *J Neurosci Off J Urosci* 2011 May 18; 31: 7477–7485.
33. Ide K, Schmalbruch IK, Quistorff B, et al. Lactate, glucose and O₂ uptake in human brain during recovery from maximal exercise. *J Physiol* 2000; 522 Pt 1: 159–164.
34. van Hall G, Strømstad M, Rasmussen P, et al. Blood lactate is an important energy source for the human brain. *J Cereb Blood Flow Metab off J Tab* 2009; 29: 1121–1129.
35. Patet C, Suys T, Carteron L, et al. Cerebral lactate metabolism after traumatic brain injury. *Curr Neurol Neurosci Rep* 2016; 16: 31.
36. Oddo M, Levine JM, Frangos S, et al. Brain lactate metabolism in humans with subarachnoid hemorrhage. *Stroke* 2012; 43: 1418–1421.
37. Schurr A, Payne RS, Miller JJ, et al. Blockade of lactate transport exacerbates delayed neuronal damage in a rat model of cerebral ischemia. *Brain Res* 2001; 895: 268–272.
38. Roumes H, Dumont U, Sanchez S, et al. Neuroprotective role of lactate in rat neonatal hypoxia-ischemia. *J Cereb Blood Flow Metab* 2021; 41: 342–358.
39. Scafidi S, Jernberg J, Fiskum G, et al. Metabolism of exogenous [2,4-¹³C]β-Hydroxybutyrate following traumatic brain injury in 21–22-day-old rats: an ex vivo NMR study. *Metabolites* 2022; 12: 710.
40. Bouzat P, Sala N, Suys T, et al. Cerebral metabolic effects of exogenous lactate supplementation on the injured human brain. *Intensive Care Med* 2014; 40: 412–421.
41. Quintard H, Patet C, Zerlauth J-B, et al. Improvement of neuroenergetics by hypertonic lactate therapy in patients with traumatic brain injury is dependent on baseline cerebral lactate/pyruvate ratio. *J Neurotrauma* 2016; 33: 681–687.
42. Jalloh I, Helmy A, Shannon RJ, et al. Lactate uptake by the injured human brain: Evidence from an arteriovenous gradient and cerebral microdialysis study. *J Neurotrauma* 2013; 30: 2031–2037.
43. Almeida A, Jimenez-Blasco D and Bolaños JP. Cross-talk between energy and redox metabolism in astrocyte-neuron functional cooperation. Bolaños JP, editor. *Essays Biochem* 2023; 67: 17–26.
44. Rodriguez-Rodriguez P, Fernandez E and Bolaños JP. Underestimation of the pentose-phosphate pathway in intact primary neurons as revealed by metabolic flux analysis. *J Cereb Blood Flow Metab* 2013; 33: 1843–1845.
45. Bolaños JP. Bioenergetics and redox adaptations of astrocytes to neuronal activity. *J Neurochem* 2016; 139 Suppl 2: 115–125.
46. Li Z, Zhang B, Yao W, et al. APC-Cdh1 regulates neuronal apoptosis through modulating glycolysis and

- pentose-phosphate pathway after oxygen-glucose deprivation and reperfusion. *Cell Mol Neurobiol* 2019; 39: 123–135.
47. Kim DY, Davis LM, Sullivan PG, et al. Ketone bodies are protective against oxidative stress in neocortical neurons. *J Neurochem* 2007; 101: 1316–1326.
 48. Maalouf M, Sullivan PG, Davis L, et al. Ketones inhibit mitochondrial production of reactive oxygen species production following glutamate excitotoxicity by increasing NADH oxidation. *Neuroscience* 2007; 145: 256–264.
 49. Jarrett SG, Milder JB, Liang LP, et al. The ketogenic diet increases mitochondrial glutathione levels. *J Neurochem* 2008; 106: 1044–1051.
 50. Lemire J, Auger C, Mailloux R, et al. Mitochondrial lactate metabolism is involved in antioxidative defense in human astrocytoma cells. *J Neurosci Res* 2014; 92: 464–475.
 51. Pötzsch A, Zocher S, Bernas SN, et al. L-lactate exerts a pro-proliferative effect on adult hippocampal precursor cells in vitro. *iScience* 2021; 24: 102126.
 52. Augusto-Oliveira M, Arrifano GP, Lopes-Araújo A, et al. What do microglia really do in healthy adult brain? *Cells* 2019; 8: 1293.
 53. Ma Y, Wang J, Wang Y, et al. The biphasic function of microglia in ischemic stroke. *Prog Neurobiol* 2017; 157: 247–272.
 54. Monsorno K, Buckinx A and Paolicelli RC. Microglial metabolic flexibility: emerging roles for lactate. *Trends Endocrinol Metab TEM* 2022; 33: 186–195.
 55. Hong H, Su J, Zhang Y, et al. A novel role of lactate: promotion of akt-dependent elongation of microglial process. *Int Immunopharmacol* 2023; 119: 110136.
 56. Huang C, Wang P, Xu X, et al. The ketone body metabolite β -hydroxybutyrate induces an antidepressant-associated ramification of microglia via HDACs inhibition-triggered akt-small RhoGTPase activation. *Glia* 2018; 66: 256–278.
 57. Guo M, Wang X, Zhao Y, et al. Ketogenic diet improves brain ischemic tolerance and inhibits NLRP3 inflammasome activation by preventing Drp1-Mediated mitochondrial fission and endoplasmic reticulum stress. *Front Mol Neurosci* 2018; 11: 86.
 58. Lin C, Wang S, Xie J, et al. Ketogenic diet and β -Hydroxybutyrate alleviate ischemic brain injury in mice via an IRAK1-dependent pathway. *Eur J Pharmacol* 2023; 955: 175933.
 59. Attwell D, Buchan AM, Charpak S, et al. Glial and neuronal control of brain blood flow. *Nature* 2010; 468: 232–243.
 60. Gordon GRJ, Choi HB, Rungta RL, et al. Brain metabolism dictates the polarity of astrocyte control over arterioles. *Nature* 2008; 456: 745–749.
 61. Carteron L, Solari D, Patet C, et al. Hypertonic lactate to improve cerebral perfusion and glucose availability after acute brain injury. *Crit Care Med* 2018; 46: 1649–1655.
 62. Bergersen LH and Gjedde A. Is lactate a volume transmitter of metabolic states of the brain? *Front Neuroenergetics* 2012; 4: 5.
 63. Lauritzen KH, Morland C, Puchades M, et al. Lactate receptor sites link neurotransmission, neurovascular coupling, and brain energy metabolism. *Cereb Cortex N Cortex* 2014; 24: 2784–2795.
 64. Ma D, Wang AC, Parikh I, et al. Ketogenic diet enhances neurovascular function with altered gut microbiome in young healthy mice. *Sci Rep* 2018; 8: 6670.
 65. Svart M, Gormsen LC, Hansen J, et al. Regional cerebral effects of ketone body infusion with 3-hydroxybutyrate in humans: reduced glucose uptake, unchanged oxygen consumption and increased blood flow by positron emission tomography. A randomized, controlled trial. *PLoS One* 2018; 13: e0190556.
 66. Hasselbalch SG, Madsen PL, Hageman LP, et al. Changes in cerebral blood flow and carbohydrate metabolism during acute hyperketonemia. *Am J Physiol* 1996; 270: E746–51.
 67. Carney N, Totten AM, O'Reilly C, et al. Guidelines for the management of severe traumatic brain injury, fourth edition. *Neurosurgery* 2017; 80: 6–15.
 68. Anstey JR, Taccone FS, Udy AA, TBI Collaborative, et al. Early osmotherapy in severe traumatic brain injury: an international multicenter study. *J Neurotrauma* 2020; 37: 178–184.
 69. Ichai C, Armando G, Orban J-C, et al. Sodium lactate versus mannitol in the treatment of intracranial hypertensive episodes in severe traumatic brain-injured patients. *Intensive Care Med* 2009; 35: 471–479.
 70. Bernini A, Miroz J-P, Abed-Maillard S, et al. Hypertonic lactate for the treatment of intracranial hypertension in patients with acute brain injury. *Sci Rep* 2022; 12: 3035.
 71. Ichai C, Orban JC and Fontaine E. Sodium lactate for fluid resuscitation: the preferred solution for the coming decades? *Crit Care Lond Care* 2014; 18: 163.
 72. Duhaut DE, Heurteaux C, Gandin C, et al. The anti-edematous effect of exogenous lactate therapy in traumatic brain injury: a physiological and mechanistic approach. *Neurocrit Care* 2021; 35: 747–755.
 73. Millet A, Cuisinier A, Bouzat P, et al. Hypertonic sodium lactate reverses brain oxygenation and metabolism dysfunction after traumatic brain injury. *Br J Anaesth* 2018; 120: 1295–1303.
 74. Suzuki M, Suzuki M, Kitamura Y, et al. β -Hydroxybutyrate, a cerebral function improving agent, protects rat brain against ischemic damage caused by permanent and transient focal cerebral ischemia. *Jpn J Pharmacol* 2002; 89: 36–43.
 75. Prins ML, Lee SM, Fujima LS, et al. Increased cerebral uptake and oxidation of exogenous betaHB improves ATP following traumatic brain injury in adult rats. *J Neurochem* 2004; 90: 666–672.
 76. Prins ML, Fujima LS and Hovda DA. Age-dependent reduction of cortical contusion volume by ketones after traumatic brain injury. *J Neurosci Res* 2005; 82: 413–420.
 77. Camberos-Luna L, Gerónimo-Olvera C, Montiel T, et al. The ketone body, β -Hydroxybutyrate stimulates the autophagic flux and prevents neuronal death induced

- by glucose deprivation in cortical cultured neurons. *Neurochem Res* 2016; 41: 600–609.
78. Jourdain P, Rothenfusser K, Ben-Adiba C, et al. Dual action of L-Lactate on the activity of NR2B-containing NMDA receptors: from potentiation to neuroprotection. *Sci Rep* 2018; 8: 13472.
 79. Juge N, Gray JA, Omote H, et al. Metabolic control of vesicular glutamate transport and release. *Neuron* 2010; 68: 99–112.
 80. Prins ML. Cerebral metabolic adaptation and ketone metabolism after brain injury. *J Cereb Blood Flow Metab* 2008; 28: 1–16.
 81. Tassinari I, D'Ávila, Andrade MKG, da Rosa LA, et al. Lactate administration reduces brain injury and ameliorates behavioral outcomes following neonatal hypoxia–ischemia. *Neuroscience* 2020; 448: 191–205.
 82. Alessandri B, Schwandt E, Kamada Y, et al. The neuroprotective effect of lactate is not due to improved glutamate uptake after controlled cortical impact in rats. *J Neurotrauma* 2012; 29: 2181–2191.
 83. Berthet C, Lei H, Thevenet J, et al. Neuroprotective role of lactate after cerebral ischemia. *J Cereb Blood Flow Metab off J Tab* 2009; 29: 1780–1789.
 84. Roumes H, Pellerin L and Bouzier-Sore AK. Effet neuroprotecteur du lactate dans l'hypoxie-ischémie cérébrale néonatale. *Med Sci (Paris)* 2020; 36: 973–976.
 85. Castillo X, Rosafio K, Wyss MT, et al. A probable dual mode of action for both L- and D-lactate neuroprotection in cerebral ischemia. *J Cereb Blood Flow Metab* 2015; 35: 1561–1569.
 86. Berthet C, Castillo X, Magistretti PJ, et al. New evidence of neuroprotection by lactate after transient focal cerebral ischaemia: Extended benefit after intracerebroventricular injection and efficacy of intravenous administration. *Cerebrovasc Dis* 2012; 34: 329–335.
 87. Crespy T, Durost M, Fricault P, et al. Hypertonic sodium lactate to alleviate functional deficits following diffuse traumatic brain injury: an osmotic or a Lactate-Related effect? *Neurocrit Care* 2021; 34: 795–803.
 88. Rice AC, Zsoldos R, Chen T, et al. Lactate administration attenuates cognitive deficits following traumatic brain injury. *Brain Res* 2002; 928: 156–159.
 89. Gambardella I, Ascione R, D'Agostino DP, et al. Systematic review – neuroprotection of ketosis in acute injury of the mammalian central nervous system: a meta-analysis. *J Neurochem* 2021; 158: 105–118.
 90. Gibson CL, Murphy AN and Murphy SP. Stroke outcome in the ketogenic state – a systematic review of the animal data. *J Neurochem* 2012; 123 Suppl 2: 52–57.
 91. van Gemert LA, de Galan BE, Wevers RA, et al. Lactate infusion as therapeutic intervention: a scoping review. *Eur J Pediatr* 2022; 181: 2227–2235.
 92. Bisri T, Utomo B and Fuadi I. Exogenous lactate infusion improved neurocognitive function of patients with mild traumatic brain injury. *Asian J Neurosurg* 2016; 11: 151–159.
 93. Ichai C, Payen J-F, Orban J-C, et al. Half-molar sodium lactate infusion to prevent intracranial hypertensive episodes in severe traumatic brain injured patients: a randomized controlled trial. *Intensive Care Med* 2013; 39: 1413–1422.
 94. Gallagher CN, Carpenter KLH, Grice P, et al. The human brain utilizes lactate via the tricarboxylic acid cycle: a ¹³C-labelled microdialysis and high-resolution nuclear magnetic resonance study. *Brain* 2009; 132: 2839–2849.
 95. Clarke K, Tchabanenko K, Pawlosky R, et al. Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol RTP* 2012; 63: 401–408.
 96. Herrero-Mendez A, Almeida A, Fernández E, et al. The bioenergetic and antioxidant status of neurons is controlled by continuous degradation of a key glycolytic enzyme by APC/C-Cdh1. *Nat Cell Biol* 2009; 11: 747–752.