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Immunodeficiency and lymphoma in Jacobsen syndrome

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Hypogammaglobulinemia.

Palabras clave: Síndrome de Jacobsen. Síndrome deleción terminal

Inmunodeficiencia. Linfoma. Hipogammaglobulinemia.

Jacobsen syndrome (JS; ORPHA:2308) is a rare disorder with an estimated prevalence of 1 in

100,000 births and a female to male ratio of 2:1 [1,2]. First described by Jacobsen in 1973, JS

commonly results from partial deletion of the terminal long arm of chromosome 11 (11q

deletion). Clinical manifestations are diverse and are frequently associated with Paris-

Trousseau syndrome, which is characterized by thrombocytopenia and platelet dysfunction [2].

The clinical presentation varies from growth and psychomotor retardation to severe facial

dysmorphia, malformations of the heart, gastrointestinal, genitourinary, ocular, hearing,

skeletal and/or central nervous system. Immunological and hormonal problems may also be

present [2].

Here we report the case of a patient with dysmorphic features, mental impairment, Paris-

Trousseau syndrome and transient immunodeficiency of childhood, who lived with a false

diagnosis for many years.

Our patient is the second child of a healthy non-consanguineous Swiss couple, with no relevant

family history. He was born at thirty-two weeks following a pregnancy complicated by

intrauterine growth restriction. In the first year, he presented with failure to thrive, delayed grow

and thrombocytopenia in laboratory studies. He only started walking at the age of two and his

language and speech development were significantly delayed, with consecutive intellectual

disability requiring special educational support. He has short stature, dysmorphic features

including bilateral ptosis, strabismus, long philtrum, thin lips, bilateral clinodactyly of the 5th

finger and a limited elbow mobility that required surgical correction. Intravenous

immunoglobulins replacement therapy (IRT, 0.4 g/kg/monthly) was given for recurrent

infectious tonsillitis, otitis and bronchitis that required regular courses of antibiotic treatment

from the age of eight until the age of twelve. Thereafter, no relevant infections were observed.

At the age of forty-six he presented with dysphagia and tonsillitis resistant to antibiotic

treatment. A biopsy of the right tonsil showed a non-otherwise specified (NOS) high-grade B

cell lymphoma (CD20+ CD79a+, CD10+ and BCL6+, EBER-, CD30-, IRF4-, CD5- and

PDL1), with a proliferation rate of the E3 ubiquitin-protein ligase MIB1 of 100%. PDL1+

tingible body macrophages without necrosis were found. MYC, Bcl-2 or Bcl-6 rearrangements

were absent. Serological analyses for active viral infections herpes and varicella zoster, EBV,

CMV, parvovirus B19, HIV, and hepatitis B and C were negative.

He received six cycles of R-EPOCH chemotherapy (rituximab 570 mg, etoposide phosphate 75

mg/m², prednisone 60 mg/m², vincristine sulfate 0.4 mg/m², cyclophosphamide 750 mg/m² and

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doxorubicin hydrochloride 10mgm²), four cycles of intrathecal methotrexate (12mg) and two

cycles of high dose intravenous methotrexate (3000 mg/m²) over six months. At follow-up, a

PET-CT scan confirmed a successful recovery from the lymphoma, but showed residual

bronchiectasis, pulmonary infiltrates and ground glass opacities in the lower lobe of the left

lung and the apex of the right lung. The patient suffered from recurring upper and lower

respiratory tract infections requiring frequent antibiotic treatments.

The biological work-up revealed thrombocytopenia, hypogammaglobinemia, lymphopenia

with low B-cell, CD4+ and CD4+gamma-delta T-cells (table, see supplementary file).

Response to pneumococcal polysaccharide vaccination was absent. Monthly IRT (0.6 g/kg) was

initiated: IgG through-levels normalized (10.9 g/l) after three months, and the patient reported

no more infections. He is currently receiving IRT, and because of neutropenia and lymphopenia,

he is receiving prophylaxis with atovaquone.

A diagnosis of Noonan syndrome has been assumed since infancy due to his dysmorphic

features, short stature and thrombocytopenia, but this had never been confirmed by genetic

testing. With the patient's and his legal guardian's consent, an array-CGH analysis was

performed using Human Genome CGH Microarray Kit G3 180 (Agilent Technologies, Palo

Alto, USA) with approximately 13KB overall median probe spacing. Labelling and

hybridization were performed following the protocols provided by the manufacturer. Graphical

overview was obtained using the Agilent Genomic Workbench 7.0.4.0. and data analysis was

done with UCSC Genome Browser Human Genome build19.

The array-CGH analysis (Figure 1) revealed a heterozygous terminal loss of approximately

10.7Mb on chromosome 11, in the region q24.2q25 between positions 124,170,725 and

134,927,114bp, encompassing 52 Online Mendelian Inheritance in Man (OMIM) genes,

including as ETS1 (ETS Proto-oncogene, transcription factor), NRGN (Neurogranin,

OMIM602350), with 18 OMIM morbid genes identified, such as FLI1 (Friend leukemia virus

integration 1, OMIM193067). Parental samples were not available.

The terminal loss on the long arm of chromosome 11, including at least five genes,

BSX, NRGN, ETS-1, FLI-1, and RICS (ARHGAP32), is compatible with JS (OMIM 147791).

JS has recently been recognized as a primary immunodeficiency [3-6]. It is often associated

with recurrent respiratory, urinary and ENT infections [1]. JS presents with an antibody

deficiency and impaired response to pneumococcal polysaccharide vaccination, or features of

combined immunodeficiency. Late-onset of clinical symptoms of immunodeficiency have been

reported. Antibiotic prophylaxis and IRT may be necessary to control recurring infection. Genes

involved in immune regulation are suppressed in the 11q deletion syndrome. The deletion on

chromosome 11 in our patient comprised 52 OMIM genes including 11 genes of pathological

significance such as the ETS1, NRGN and FLI-1; a proto-oncogene involved in platelet

functions whose deletion is associated with Paris-Trousseau syndrome. ETS-1 knockout in

mice show significant defects in T, B, and NK cell development. Based on the patient's medical

history, including recurrent infections requiring antibiotic treatment and IRT from age eight to

twelve, we assume that the patient already suffered from immunodeficiency during childhood.

We further hypothesize that chemotherapy led to an exacerbation of this immunodeficiency.

Unfortunately, laboratory values prior to chemotherapy treatment were not available.

While there are no reports available on the association between JS and neoplasia in the

literature, 11q deletions have been associated with myeloid and lymphoid neoplasia, especially

Burkitt-like (MYC-negative) tumors [7]. ETS-1 may play a role in malignant transformation of

hematopoietic neoplasms, including B-cell malignancies [8]. The occurrence of these 2 unusual

diagnoses (JS and B-cell lymphoma) could be incidental, but an association between

chromosome 11q deletion and malignancies is plausible [7,9,10].

We believe that JS patients should be screened for immunodeficiency at diagnosis and during

follow-up to prevent recurring infections. A high degree of vigilance is required due to the

elevated risk of Burkitt-like lymphomas in patients 11q deletion.

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FIGURE

Figure 1. Array-CGH analysis showing a heterozygous terminal loss of approximately 10.7Mb on chromosome 11, in the region q24.2q25.

