



Article scientifique

Article

2022

Accepted version

Open Access

This is an author manuscript post-peer-reviewing (accepted version) of the original publication. The layout of the published version may differ .

2022 EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in adults with autoimmune inflammatory rheumatic diseases

Fragoulis, George E; Nikiphorou, Elena; Dey, Mrinalini; Zhao, Sizheng Steven; Courvoisier, Delphine; Arnaud, Laurent; Atzeni, Fabiola; Behrens, Georg MN; Bijlsma, Johannes WJ; Böhm, Peter; Constantinou, Costas A; Garcia-Diaz, Silvia; Kapetanovic, Meliha Crnkic; Lauper, & Kim [and 10 more]

How to cite

FRAGOULIS, George E et al. 2022 EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in adults with autoimmune inflammatory rheumatic diseases. In: Annals of the rheumatic diseases, 2022, vol. 82, n° 6, p. 742–753. doi: 10.1136/ard-2022-223335

This publication URL: <https://archive-ouverte.unige.ch/unige:170889>

Publication DOI: [10.1136/ard-2022-223335](https://doi.org/10.1136/ard-2022-223335)



King's Research Portal

DOI:

[10.1136/ard-2022-223335](https://doi.org/10.1136/ard-2022-223335)

Document Version

Peer reviewed version

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Fragoulis, G. E., Nikiphorou, E., Dey, M., Zhao, S. S., Courvoisier, D. S., Arnaud, L., Atzeni, F., Behrens, G. M., Bijlsma, J. W., Böhm, P., Constantinou, C. A., Garcia-Diaz, S., Kapetanovic, M. C., Lauper, K., Luís, M., Morel, J., Nagy, G., Poleverino, E., van Rompay, J., ... Hyrich, K. L. (2022). 2022 EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in adults with autoimmune inflammatory rheumatic diseases. *Annals of the Rheumatic Diseases*, [223335]. <https://doi.org/10.1136/ard-2022-223335>

Citing this paper

Please note that where the full-text provided on King's Research Portal is the Author Accepted Manuscript or Post-Print version this may differ from the final Published version. If citing, it is advised that you check and use the publisher's definitive version for pagination, volume/issue, and date of publication details. And where the final published version is provided on the Research Portal, if citing you are again advised to check the publisher's website for any subsequent corrections.

General rights

Copyright and moral rights for the publications made accessible in the Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognize and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the Research Portal

Take down policy

If you believe that this document breaches copyright please contact librarypure@kcl.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.

2022 EULAR Recommendations for screening and prophylaxis of chronic and opportunistic infections in adults with autoimmune inflammatory rheumatic diseases

George E Fragoulis^{1,2}, Elena Nikiphorou^{3,4}, Mrinalini Dey^{5,6}, Sizheng Steven Zhao⁷, Delphine S Courvosier⁸, Laurent Arnaud⁹, Fabiola Atzeni¹⁰, Georg MN Behrens¹¹, Johannes WJ Bijlsma¹², Peter Böhm¹³, Costas A Constantinou¹⁴, Silvia Garcia-Diaz¹⁵, Meliha Kapetanovic¹⁶, Kim Lauper^{7,8}, Marianna Luis¹⁷, Jacques Morel¹⁸, Gyorgy Nagy^{19,20,21}, Eva Poleverino²², Jef van Rompay²³, Marco Sebastiani²⁴, Anja Strangfeld²⁵, Annette de Thurah^{26,27}, James Galloway^{3,4*}, Kimme Hyrich^{7,28*}

1. Joint Academic Rheumatology program, First Department of Propaedeutic and Internal Medicine, National and Kapodistrian University of Athens, Athens, Greece
2. University of Glasgow, Institute of Infection, Immunity and Inflammation
3. Rheumatology Department, King's College Hospital, London, UK.
4. Centre for Rheumatic Diseases, King's College London, London, UK
5. Institute of Life Course and Medical Sciences, University of Liverpool, Brownlow Hill, Liverpool, L69 3BX, UK.
6. Department of Rheumatology, Countess of Chester Hospital NHS Foundation Trust, Liverpool Road, Chester, CH2 1UL, UK.
7. Centre for Epidemiology Versus Arthritis, Division of Musculoskeletal and Dermatological Science, School of Biological Sciences, Faculty of Biological Medicine and Health, The University of Manchester, Manchester Academic Health Science Centre, Manchester, UK.
8. Division of Rheumatology, University Hospitals of Geneva, Switzerland
9. Department of Rheumatology, National Reference Center for Autoimmune Diseases (RESO), Hôpitaux Universitaires de Strasbourg, INSERM UMR-S 1109, Strasbourg, France.
10. Rheumatology Unit, Department of Experimental and Internal Medicine, University of Messina, Messina, Italy

11. Department for Rheumatology and Immunology, Hannover Medical School, 30625 Hannover, Germany
12. Dept of Rheumatology & Clinical Immunology, University Medical Center Utrecht, Utrecht, The Netherlands
13. German League against Rheumatism, Bonn, Germany
14. Department of Internal Medicine, Nicosia General Hospital, Nicosia, Cyprus, and Unit for Surveillance and Control of Communicable Diseases, Medical and Public Health Services, Ministry of Health, Republic of Cyprus
15. Rheumatology Department, Complex Hospitalari Moises Broggi, Barcelona, Spain.
16. Lund University, Skåne University Hospital, Department of Clinical Sciences Lund, Rheumatology, Lund, Sweden
17. Department of Rheumatology, Coimbra Hospital and University Centre, Coimbra, Portugal and Faculty of Medicine, University of Coimbra, Coimbra, Portugal
18. Department of Rheumatology, Montpellier University Hospital and University of Montpellier, PhyMedExp, Université de Montpellier, INSERM, CNRS, Montpellier, France
19. Department of Rheumatology and Clinical Immunology, Department of Internal Medicine and Oncology, Semmelweis University, Budapest, Hungary
20. Heart and Vascular Center, Semmelweis University, Budapest, Hungary
21. Department of Genetics, Cell- and Immunobiology, Semmelweis University, Budapest, Hungary
22. Pneumology Department, Hospital Universitari Vall d'Hebron, Vall d'Hebron Institut de Recerca (VHIR), Vall d'Hebron Barcelona Hospital Campus, Barcelona, Spain.
23. EULAR People with Arthritis / Rheumatism across Europe (PARE), Belgium patient partner program
24. Rheumatology Unit, University of Modena and Reggio Emilia, Modena, Italy
25. Epidemiology and Health Services Research, German Rheumatism Research Centre (DRFZ) Berlin and Charite University Medicine Berlin, Berlin, Germany.
26. Department of Rheumatology, Aarhus University Hospital, Aarhus, Denmark

27. Department of Clinical Medicine, Aarhus University, Aarhus, Denmark

28. National Institute of Health Research Manchester Biomedical Research Centre, Manchester University
NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester, United Kingdom

Corresponding author: Kimme Hyrich, Stopford Building, The University of Manchester, Oxford Rd,
Manchester, M13 9PT, United Kingdom, e-mail: Kimme.Hyrich@manchester.ac.uk

Keywords: chronic infections, opportunistic infections, tuberculosis, hepatitis B, hepatitis C, Pneumocystis
jirovecii, human immunodeficiency virus, varicella zoster virus, screening, prophylaxis.

Abstract

Objectives: To develop EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in patients with autoimmune inflammatory rheumatic diseases (AIIRD).

Methods: An international Task Force (TF) (22 members/15 countries) formulated recommendations, supported by systematic literature review findings. Level of evidence and grade of recommendation were assigned for each recommendation. Level of agreement was provided anonymously by each TF member.

Results: Four overarching principles (OAP) and eight recommendations were developed. The OAPs highlight the need for infections to be discussed with patients and with other medical specialties, in accordance with national regulations. In addition to b/tsDMARDs for which screening for latent tuberculosis (TB) should be performed, screening could be considered also before csDMARDs, glucocorticoids and immunosuppressants. Interferon gamma release assay should be preferred over tuberculin skin test, where available. Hepatitis B (HBV) anti-viral treatment should be guided by HBV status defined prior to starting anti-rheumatic drugs. All patients positive for Hepatitis-C-RNA should be referred for antiviral treatment. Also, patients who are non-immune to varicella zoster virus should be informed about the availability of post-exposure prophylaxis should they contact with this pathogen. Prophylaxis against *Pneumocystis jirovecii* seems to be beneficial in patients treated with daily doses > 15-30mg of prednisolone or equivalent for > 2-4 weeks..

Conclusions: These recommendations provide guidance on the screening and prevention of chronic and opportunistic infections. Their adoption in clinical practice is recommended to standardize and optimize care to reduce the burden of opportunistic infections in people living with AIIRD.

Word count: 4753

Introduction

Opportunistic and chronic infections, i.e. those which present more commonly or more severely in people who are immunocompromised¹, are encountered in the setting of Autoimmune Inflammatory Rheumatic Diseases (AIIRD) and are often associated with immunosuppressive and immunomodulatory treatments

used for these diseases. Although it is recognised that screening procedures and prophylactic measures should be followed, clinical practice is largely heterogeneous and relevant recommendations are often lacking or are disparately located across the literature. There is, therefore, a need for collating evidence for different AIIRD and treatment regimens to be used as a single point of reference in routine clinical practice.^{2,3}

Setting a single set of guidelines for infection screening and prophylaxis is challenging, as recommendations and procedures cannot be unified across all infections and organisms due to differences in area of residence, type of AIIRD and associated risk, the anti-rheumatic treatment received, and other factors that may present additional layers of complexity, such as age and comorbidities.⁴⁻⁶ Our goal was to formulate a set of recommendations, taking these challenges into account, to inform rheumatologists and health care providers in their decision-making when caring for people living with AIIRD, to ensure that these infections can be identified and adequately managed.

A EULAR Task Force (TF) has been formed, comprised of health care professionals and patients across different disciplines and countries, to develop the first EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in patients living with AIIRD based on the best available scientific evidence. This manuscript presents the work of this TF and the final set of recommendations.

Methods

The EULAR standardized operating procedures (SOP)⁷ were followed throughout the undertaking of this work. The project was approved by the EULAR executive committee (No: CLI 118). The steering committee included a main convenor (KH) and a co-convenor (JG), one methodologist (EN), a co-methodologist (DC) and a main fellow (GF). Two co-fellows (MD and SZ) supported the undertaking of the systematic literature review (SLR) (under submission), especially during validation steps (see below). TF members were selected based on their experience in the field of infections in the setting of AIIRD, considering also gender and regional equity. The final TF consisted of 22 people (including steering committee members) from 15 different European countries. Two patient research partners, two health care professionals in rheumatology, two infectious disease doctors with an interest in rheumatology and one pulmonologist were included in addition to rheumatologists/epidemiologists (including two Emerging EULAR Network [EMEUNET] members).

In preparation of the first TF meeting, the steering group identified research questions of interest and relevance, leading to a scoping review (available on request) by the fellow (GF). The scoping review

provided an overview of the existing literature on chronic and opportunistic infections in AIIRD. During the first TF meeting which was held virtually in September 2020, the results of the scoping review were presented and the research questions for the main SLR were discussed and modified as deemed appropriate by the TF. Additionally, there was review and discussion on the pathogens that would be included in the subsequent SLR (presented in Supplementary material 1), based on the findings of the scoping review as well as expert opinion of TF members including the two infectious disease doctors who reviewed separately the list of microbes.

Afterwards, the steering committee transformed the research questions (Supplementary material 1) into epidemiological questions that were addressed via the SLR. The latter was registered in PROSPERO (No: CRD42021244732) and was performed as *per guidance* provided in the Cochrane Handbook.⁸ The SLR results were reported according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines.⁹

The SLR was performed for studies published from inception up to the 5th December 2021. However, new studies that were published after this date and up until the date of the second TF meeting (18th January 2022) were also considered, where these provided additional evidence relevant to the research questions. The SLR focused on studies examining the efficacy of screening and prophylaxis for chronic and opportunistic infections. Results as well as details about the methodology of the SLR are presented separately. The results of the SLR were presented in the second virtual meeting (split over December 2021 and January 2022) during which the overarching principles (OAP) and the recommendations were formulated and voted upon. Recommendations and OAPs were accepted if $\geq 75\%$ of the members agreed in a first round of anonymized voting; if this agreement was not reached, the recommendation/OAP were reworded with a voting cut-off of $\geq 67\%$. If this was not achieved, voting in favour by $>50\%$ of the TF members was required as part of a third and final round of voting, after rephrasing. As per EULAR guidance,⁷ the Oxford Evidence Based Medicine categorization was followed for applying level of evidence (LoE) and grade of recommendation (GoR).¹⁰ A research agenda was formed, based on the identified unmet need and gaps in the literature found via the SLR and in discussions between TF members. Finally, after the second meeting, TF members provided their level of agreement (LoA) which each OAP and recommendation from 0 (=no agreement) to 10 (=full agreement), via an anonymized online survey.

Results

These recommendations address the screening procedures and prevention measures that should be followed in people living with AIIRD, treated (or about to be treated) with anti-rheumatic drugs. After the identification of the pathogens that were covered in the respective SLR, extensive discussions took place (during the second meeting) about the nomenclature that should be followed for the various anti-rheumatic drugs used. The TF reached consensus (agreed by 88% of the TF members) on the use of a four-category system as follows: 1. b- and ts-DMARDs: all biologic and targeted synthetic DMARDs (except apremilast), 2. csDMARDs: methotrexate, leflunomide. Sulfasalazine and hydroxychloroquine were exempted from this category, and the TF members agreed to name them specifically, if needed, as it was thought that they only have a mild immunomodulatory/immunosuppressive effect. 3. other immunosuppressants: cyclophosphamide, mycophenolate mofetil, azathioprine, cyclosporin, tacrolimus. 4. glucocorticoids. These categories were adopted, with some modifications from recently published expert opinion and other consensus papers. It is recognized that the rheumatology community should discuss and reach a final consensus about the terminology used to describe these drugs.¹¹⁻¹³ The term “anti-rheumatic treatment/drugs” is also used in this manuscript, encompassing all the above-mentioned categories. The TF meetings resulted in the formulation of four OAPs and eight recommendations (Table 1).

Overarching principles

OAPs form the basis on which the recommendations were built. They reflect the rationale behind the development of this set of recommendations and they highlight key concepts in the management of AIIRD. In total, four OAPs that apply across all recommendations were formulated and met with high consensus by the TF (Table 1).

A. The risk of chronic and opportunistic infections should be considered and discussed with all patients with AIIRD prior to treatment with csDMARDs, tsDMARDs, bDMARDs, immunosuppressants and/or glucocorticoids and reassessed periodically.

Chronic and opportunistic infections are an important aspect of AIIRD and a significant cause of morbidity and mortality.^{14 15} This principle was regarded as the cornerstone of all formulated recommendations. Anti-rheumatic treatment is a widely accepted risk for infections and thus the respective risk should be explained and discussed with patients, including how these risks can be minimised. The association of high disease activity with increased infection rates should also be considered.^{16 17} Shared-decision making is

increasingly recognized as an important component of good clinical care in the management of people living with AIIRD,¹⁸⁻²¹ who should also be educated to identify promptly signs and symptoms of infections and how to seek relevant medical attention. Considering also that escalation or change in treatment might be necessary and late reactivation of latent infections is possible, the respective risk should be re-assessed and discussed periodically.

B. Collaboration between rheumatologists and other specialists including but not limited to infectious disease doctors, gastroenterologists, hepatologists and pulmonologists is important.

Rheumatologists carry primary responsibility when it comes to the treatment of people living with AIIRD and should work in close collaboration with other specialties when planning prevention or management of chronic and opportunistic infections in patients receiving anti-rheumatic drugs. This is an important component of multi-disciplinary care and particularly relevant in the setting of these recommendations. Given that tuberculosis (TB) and hepatitis are among the most commonly discussed infections in people with AIIRD, teamwork with pulmonologists and hepatologists/gastroenterologist, respectively, is important. Other specialties, including infectious disease doctors, radiologists, haematologists and microbiologists also have a crucial role in guiding the screening and prophylaxis of chronic and opportunistic infections in AIIRD patients.

C. Individual risk factors should be considered in the decision for screening and prophylaxis of chronic and opportunistic infections and reassessed periodically.

An individualised approach has been identified as a key principle of this set of recommendations, since several factors are known to increase the susceptibility for specific preventable infections.^{4-6 22-24} These include, but are not limited to, age, comorbidities (e.g. lung disease), co-treatment with other medications and travelling/living in endemic areas. Given that these parameters can change, and that escalation in the treatment of AIIRD is not unusual (OAP A), the presence of risk factors for chronic and opportunistic infections should be reassessed periodically. From this point of view, medical history including previous infections, lifestyle (e.g. frequent travelling), habits (e.g. smoking), vaccination status and previous countries of residence should be taken into account.

D. National guidelines and recommendations, amongst other country/region-level factors pertaining to endemic infectious diseases, should be considered.

It was recognized by the TF members that there are significant variations in the strategies followed across different regions/countries. This might reflect differences in the geoepidemiology of certain pathogens,

as well as in factors related to cost and/or availability. To give an example, TB is more prevalent in specific areas of the world and/or resistance of *Mycobacterium tuberculosis* varies across countries²⁵, reflecting in the use of different therapeutic regimes/schemes for prophylaxis against latent TB reactivation. From this point of view, the TF thought it appropriate to have as an OAP that national/regional recommendations should always be taken into consideration in addition to these recommendations.

Recommendations

1. Screening for latent tuberculosis is recommended in patients prior to starting bDMARDs or tsDMARDs. Screening should also be considered in patients with increased risk for latent tuberculosis prior to starting csDMARDs, immunosuppressants and/or glucocorticoids (according to dose and duration).

Screening for latent TB before starting bDMARDs is included in screening programs of most national and international rheumatology associations, while the same applies for tsDMARDs, although there is less evidence.²⁶⁻³¹ On the other hand, there is some evidence that AIIRD patients under treatment with csDMARDs and/or glucocorticoids have also increased risk for latent TB reactivation.^{28 32-36} The minimum dose/duration of glucocorticoids above which latent TB screening should be performed, is unknown. A number of studies and other guidelines have suggested that screening should be considered particularly in those patients likely to receive >15 mg of prednisolone (or equivalent)/day for longer periods of time (e.g >4 weeks)^{33 34 37 38}. Additionally, screening for latent TB before commencement of these drugs should be considered in patients who also have accompanying TB risk factors like alcohol abuse, smoking, living with people with TB, living in endemic countries and others.^{22 39} Finally, despite being suggested that cyclophosphamide might associate with TB development in some AIIRD,^{22 40} evidence specifically addressing the impact of immunosuppressants is lacking. Recommendation for immunosuppressants at the time of drafting these recommendations, is only based on expert opinion.

2. Screening for latent tuberculosis should follow national and/or international guidelines and would typically include a chest X-ray, and interferon-gamma release assay (IGRA) over tuberculin skin test (TST) where available.

Evidence suggests that IGRA performs better than TST in the diagnosis of latent TB and is less affected by treatment with glucocorticoids, DMARDs or immunosuppressants.⁴¹⁻⁵¹ From this point of view, IGRA should be preferred over TST for TB screening. Given the low agreement between TST and IGRA,^{43 45 52-71} performing both tests can also be considered in cases of high suspicion for latent TB and/or in high-endemic countries.^{58 70 72} Concordance between different IGRAs (Quantiferon® and EliSPOT) is good thus one is not recommended over the other.^{47 73-75} Additionally, although there is no robust evidence for the usefulness of chest-X-Ray, the TF considered it appropriate that this should be included in the TB-screening procedures, especially as a negative IGRA or TST cannot exclude active TB or rule out latent TB.⁷⁶ Finally, as discussed in the SLR informing current recommendations, conversion (from negative to positive) of TST or IGRA after treatment with bDMARDs has been reported.^{63 70 77-89} Therefore, periodic re-screening could be considered, especially if risk factors exist or develop over time.^{22 39} There are no robust data to define how often re-screening should be performed and/or if there is a need to re-screen patients who switch bDMARDs or tsDMARDs; this issue has been added in the research agenda. As stated, given the regional differences in TB-burden and also issues (e.g. cost) that might affect the availability of some investigations (e.g. Quantiferon®), national and international guidelines should also be followed, where available.

3. Choice and timing of latent tuberculosis therapy should be guided by national and/or international guidelines. Special attention should be given to interactions with drugs commonly used to treat AIIRD.

Various therapeutic schemes have been used for the treatment of latent TB. These include isoniazid for 6-12 months, combination of rifampicin/isoniazid for 3-4 months, rifampicin for 4 months and once-weekly therapy of isoniazid plus rifapentine.^{28 72 77 83 90-107} Given differences in the TB-burden and drug resistances among regions/countries, the TF members advise to adhere to relevant national guidelines.

Interactions between drugs used to treat AIIRD and those used as treatment for latent TB should be considered. Monitoring of liver function tests (LFT) is necessary in patients co-treated with isoniazid and hepatotoxic drugs like methotrexate and leflunomide.^{96 108 109} Additionally, pharmacokinetics of JAK-inhibitors and glucocorticoids might be affected by co-administration with rifampicin.^{110 111}

4. All patients being considered for treatment with csDMARDs, bDMARDs, tsDMARDs, immunosuppressants and glucocorticoids (according to dose and duration) should be screened for HBV.

The risk of HBV reactivation (appearance/rise in HBV-DNA or conversion from HBsAg-negative to HBsAg-positive)¹¹² depends on the HBV-status (unexposed, vaccinated, carrier [i.e. HBsAg-positive] and resolved-HBV [anti-HBcore-positive and HBsAg-negative]) and this should be determined before the treatment for AIIRD is commenced. HBV-status would also help identify patients at risk (e.g. from their occupation) who should be vaccinated.¹¹³ Due to the complex nature of this recommendation, the TF decided to include a figure outlining the suggested procedures according to the HBV status of the patient (Figure 1).

Evidence suggests that HBV carriers (HBsAg-positive) would benefit from prophylactic treatment and thus it is advised that they should be referred to hepatologist for anti-viral prophylactic treatment.^{114 115} As outlined in the SLR informing these recommendations, data are less robust for drugs¹¹⁶⁻¹²⁵ other than bDMARDs.¹²⁶⁻¹³⁶ However, for non-bDMARDs users, referral to a hepatologist for consideration of anti-viral prophylaxis is also recommended. The exact dose and duration of glucocorticoids that would increase HBV reactivation risk cannot be inferred from existing studies. Patients receiving at least 10 mg of prednisolone or equivalent for ≥ 4 weeks are regarded by the American Gastroenterology Association¹³⁷ as a high-risk group for HBV reactivation, also supported by expert opinion.^{138 139}

For patients who have resolved-HBV (anti-HBcore-positive and HBsAg-negative), risk for HBV reactivation is lower.^{119 121-123 140-148} Baseline measurement of LFT and HBV-DNA levels and then regular (e.g every 3-6 months) monitoring of LFT and HBV-DNA levels over universal prophylaxis is advised.^{132 149-152} Referral to a hepatologist is also recommended for all patients, but is imperative for those with detectable HBV-DNA. Special attention should be given to patients considered as high-risk for HBV reactivation. These are mainly patients treated with rituximab; some investigators as well as rheumatology and/or hepatology societies have suggested that these patients should be referred to a hepatologist for consideration of prophylactic treatment irrespective of HBV-DNA levels.¹⁵³⁻¹⁵⁷ Of note, compared to people with high titres of anti-HBs antibodies, those with low titres have also been linked with greater risk of reactivation.¹⁵⁸⁻¹⁶³ In terms of prophylaxis, the TF did not suggest any anti-viral drug in favour of the other, as this is a decision that should be made by the treating hepatologist. There are no data to support a recommendation about the timing of anti-viral treatment, but it is reasonable to start ideally before or at least simultaneously with the treatment administered for AIIRD and continuing for least 6-12 months after discontinuation of anti-rheumatic treatment, as has been proposed in recommendations from rheumatology and hepatology/gastroenterology societies.^{137 153 155 157 164 165} This proposed time window for prophylaxis continuation might be longer for patients treated with rituximab.^{137 153 157 165} Given the lack of data, the TF

did not make a specific recommendation related to this. Instead, a relevant research agenda item has been agreed (see below).

5. Screening for chronic hepatitis C should be considered in patients prior to starting csDMARDs, bDMARDs, tsDMARDs, immunosuppressants and glucocorticoids (according to dose and duration). Screening is recommended for patients with elevated alanine aminotransferase (ALT) or those with known risk factors.

Most of the studies examining hepatitis C (HCV) reactivation pertain to treatment with bDMARDs, particularly TNF inhibitors, and show that HCV reactivation does occur, albeit in a low number of patients.¹⁶⁶⁻¹⁷⁰ Of note, most of these studies were published before newer, more effective drugs against HCV (e.g. direct acting anti-virals) were widely available. In the interest of public health, the TF suggests that screening should be considered in AIIRD patients before starting treatment. Considering also cost-effectiveness and geographical variations, the threshold for screening should be lower for patients with concurrent HCV risk factors (e.g. intravenous use of drugs) and/or abnormal liver function tests, especially ALT. No data exist regarding HCV screening and glucocorticoids or immunosuppressants. Therefore, recommendation for these drug categories is based on expert opinion. Screening for HCV includes anti-HCV antibodies and if these are present, measurement of HCV-RNA levels.^{164 171 172} Patients with detectable HCV-RNA should be referred for consideration of antiviral treatment. In these patients, regular monitoring with liver function tests and viral load is also advised.^{166 170 173-175}

6. Screening for HIV is recommended prior to treatment with bDMARDs and should be considered prior to treatment with csDMARDs, tsDMARDs, immunosuppressants and glucocorticoids (according to dose and duration).

No robust data exist for the safety of treatment with DMARDs, immunosuppressants or glucocorticoids in patients with Human immunodeficiency virus (HIV); however, the TF supported that screening for HIV should be undertaken prior to treatment with bDMARDs, with appropriate HIV care and treatment given where indicated. Taking also into account the importance of addressing public health and depending on cost-effectiveness and national guidelines, screening of HIV could be performed before commencing other anti-rheumatic drugs as suggested in other recommendations for specific AIIRD or drugs.^{176 177}

7. All patients commencing csDMARDs, bDMARDs, tsDMARDs, immunosuppressants and/or glucocorticoids (according to dose and duration) who are non-immune to VZV should be informed about post-exposure prophylaxis following contact with VZV.

In the TF meeting, it was discussed whether AIIRD patients should have serological screening for Varicella zoster virus (VZV) immunity. Acknowledging that status of VZV-immunity can be affected by various factors, including national regulations, access to testing, as well as previous vaccination or infection history, it was considered appropriate not to formulate a specific recommendation on this issue; however, the TF advocates the importance of establishing VZV-immunity status through a detailed past medical history of previous exposure e.g. chickenpox. Mainly based on published expert opinion^{178 179} the TF agreed that those identified as non-immune or where there is doubt about their immunity status, should be informed in advance about post-exposure prophylaxis and offered prophylaxis after contact with a person with chickenpox or shingles, according to local guidelines. There is no evidence about the level of immunosuppression/immunomodulation (type of treatment) above which, patients would have a benefit from post-exposure prophylaxis. This has been noted in the research agenda.

Prophylaxis with anti-virals against reactivation of Herpes zoster infection (shingles), as has been suggested by some in the literature (largely expert opinion)¹⁷⁹⁻¹⁸¹, could not be recommended routinely at this stage. It has been suggested that this might benefit AIIRD patients with a history of recurrent Herpes zoster infections; however, the TF considered that there was not enough evidence to support such a recommendation at this stage.

8. Prophylaxis against *Pneumocystis jirovecii* pneumonia (PCP) should be considered in patients with AIIRD in whom high doses of glucocorticoids are used, especially in combination with immunosuppressants* and depending on the risk-benefit ratio.

Prophylaxis for *Pneumocystis jirovecii* pneumonia (PCP) has been mostly examined in AIIRD patients treated with glucocorticoids. Although the minimum dose and duration of glucocorticoid treatment above which prophylaxis is recommended is not defined, evidence suggests that in daily doses > 15-30mg of prednisolone or equivalent for > 2-4 weeks, prophylaxis is beneficial.¹⁸²⁻¹⁸⁶ Most studies do not focus on a specific AIIRD. Therefore, it was not possible to make recommendations for PCP prophylaxis in individual diseases although the risk for PCP infection might be significantly different.¹⁸⁷ Data specifically addressing

the contribution of other anti-rheumatic drugs in PCP development are limited.^{188 189} On the other hand, it has been shown that co-administration of immunosuppressants with glucocorticoids^{184 185 190} increase the risk for PCP. Other features including persistent lymphopenia,^{5 6 184 185} older age and pre-existing lung disease are also considered risk factors for PCP.⁴⁻⁶

The most commonly used prophylaxis scheme is trimethoprim/sulfamethoxazole (TMP-SMX) 480mg/day (single-strength) or 960mg three times a week; of note, there is limited evidence that reduced doses (e.g. half-strength, daily) are equally effective and are associated with less adverse events.¹⁹¹⁻¹⁹⁵ It should be noted that adverse events related to TMP-SMX (e.g. nausea, headache, rash) are common, affecting about 20% of patients.¹⁹⁶ Concerns for higher adverse event rates have been expressed for individuals treated with methotrexate (in specific relation to the combination of TMP and MTX and the risk of cytopenia) or in patients with systemic lupus erythematosus (SLE).^{187 197}

Alternative prophylactic medications include atovaquone, dapsone or nebulized pentamidine. Although there is some disagreement in the literature,¹⁹⁸ it seems that they are equally effective compared to TMP-SMX¹⁹⁹⁻²⁰¹; however their usage is limited by factors like cost or need for hospital administration.²

Discussion

This is the first set of EULAR recommendations on the screening and prophylaxis of opportunistic and chronic infections in AIIRD. The four OAPs comprise the cornerstones of the 8 recommendations produced. The latter are presented and grouped *per infectious agent* (rather than per underlying rheumatic disease or by individual anti-rheumatic treatments) as the steering group and the members of the TF concluded that this was the best way to present the evidence in the respective SLR and subsequently formulate the recommendations. They should be considered as a whole for each patient.

During the development of these recommendations, we faced several challenges mainly pertaining to the variations across different types of AIIRD or anti-rheumatic drugs used. Initially, we had to decide which pathogens should be included in these recommendations. As discussed, our scoping review identified the bulk of these microorganisms, and the TF members made their additions based on their expertise. Contribution of the two infectious disease doctors who participated in this TF and reviewed the list of studies regarding microorganisms was crucial. We also reviewed an authoritative consensus about

opportunistic infections reporting during clinical trials and post-marketing surveillance of biologic therapies in immune mediated diseases ¹ and found it to be consistent with the pathogens that were included in our SLR. Of note, infection with SARS-CoV2 was not included in these recommendations, as it is covered by EULAR recommendations dedicated to this topic ²⁰².

Some infections are traditionally linked with a specific drug class (e.g. TB with TNF-inhibitors) which creates a risk of underestimating the importance of screening before commencing treatment with other drug categories (i.e., csDMARDs and glucocorticoids in the example of TB). Stronger recommendations for specific treatments could not always be made as there is a lack of data for many of the commonly used drugs in rheumatology. This includes newer medications such as the JAK-inhibitors but also well-established immunosuppressants, such as cyclophosphamide. In these cases, level and grade of recommendations were low and the respective unmet needs are captured in the research agenda.

To add another level of complexity, there is heterogeneity on clinical grounds about the screening and prevention strategies followed currently across different AIIRD. For example, prophylactic treatment for PCP with TMP-SMX is recommended in patients with ANCA-associated vasculitis ²⁰³ but not in patients with SLE, as the evidence about the latter is limited thus far. ²⁰⁴ As regards treatment with glucocorticoids, risk for specific infections like TB or HBV reactivation differs in relation to dose and duration of treatment. Therefore, where there was evidence available, specific doses/duration of glucocorticoids are proposed in this set of recommendations as a cut-off, in accordance with guidance from other societies. ^{37 137}

Finally, some pathogens are more prevalent in specific areas of the world, so special attention should be paid in these cases. Extensive discussions took place during the TF meetings about whether a separate recommendation should be included for rarer pathogens like *Histoplasma* spp., *Coccidioides* spp., *Strongyloides* spp. and others which are more prevalent in specific geographical areas. As discussed in the respective SLR, relevant evidence was scarce, despite several expert opinion articles. Eighty-two percent of the members voted that no recommendation can be formulated at this stage for these less common organisms. On the other hand, TF members agreed, as has been shown ²⁰⁵, that people living with AIIRD benefit when provided with general dietary and environmental advice to reduce their risk of infection from specific pathogens (e.g. *Listeria* spp., *Salmonella* spp.) whilst receiving treatment with bDMARDs, tsDMARDs, immunosuppressants and high-doses of glucocorticoids. Besides, patients commencing any antirheumatic therapies should be counselled about infection risk as part of self-management. ¹⁸ Furthermore, increased awareness for atypical or rarer infections (e.g. *Histoplasma* spp.) is proposed for patients living or travelling from high-endemic areas. ^{206 207}

Considering differences between countries and consistently with other EULAR recommendations²⁰ and EULAR SOP⁷, cost-effectiveness was also taken into account in the discussions that took place during the TF meetings, although such formal assessments were not conducted. As captured in the OAPs of this set of recommendation, national regulations, where they exist, should also be considered as a guidance for screening/therapeutic decisions. OAPs were phrased to stress that decision and were made on a case-by-case basis, considering concurrent risk factors (e.g. treatment with other medications, comorbidities). Importantly, screening and prophylactic procedures should be reassessed periodically. The importance of the multidisciplinary approach is also highlighted. Even though rheumatologists should always be in close collaboration and refer where appropriate to other professions, the TF underscores the central role of the rheumatologist in the management of chronic and opportunistic infections arising in the context of AIIRD and relating to the anti-rheumatic treatment received. For example, in HBV reactivation, rheumatologists should be able to understand the meaning of the various HBV screening tests and refer the patient on as appropriate. Our TF included clinicians from other disciplines (e.g infectious diseases, pulmonology) and although recommendations/guidelines from other non-rheumatology societies were not included specifically in our SLR, their views were taken into account.^{37 137 155 157}

In these recommendations, despite discussing prevention strategies, we did not include or discuss studies about vaccination, as this is covered by another set of EULAR recommendations²⁰⁸; however, screening strategies proposed herein might identify individuals who are candidates for vaccinations.

During the TF meetings, it was discussed that the TF members in collaboration with EULAR, will help towards the implementation of this set of recommendation in clinical practice. As outlined in the EULAR SOP,⁷ there are various implementation strategies, including audits and inclusion of recommendations in quality indicators. It is expected that apart from EULAR and EMEUNET, the TF members will help in the dissemination of this set of recommendations, in the first instance via their national rheumatology societies. Apart from rheumatologists and health-policy makers, HPRs should be also aware of these recommendations given their active role in the education and monitoring of people living with AIIRD.²⁰⁹ It is also important that patient associations and people living with AIIRD, who are encouraged to play an active role in shared decision making and their care pathway, are also aware of these recommendations.¹⁸ We believe that implementation of these recommendations will lead to better outcomes for patients, as it has been shown, for example, that rates of TB were significantly decreased after screening recommendations were issued at a national level.¹⁰²

For some infectious diseases (e.g. fungal infections) data are still scarce. Most of these are recognized in the work presented here and in the respective SLR and are captured in the research agenda. Hopefully these issues will be the subject of future research and will be answered in time.

In summary, this is the first set of EULAR recommendations addressing the need for guidance about screening and prophylaxis in people living with AIIRD. Variations relating to treatment, geographical and other differences were taken into account. We believe that these recommendations will be a useful aid for decision making for people living in many countries and working in different health care systems.

Research agenda

A research agenda was considered during and after the second TF meeting. Items collected for the research agenda are shown in Box 1

Box 1

General:

Does the risk of opportunistic and chronic infections differ between the different classes of DMARDs or immunosuppressive drugs?

What is the dose and duration of glucocorticoids above which the risk of opportunistic and chronic infections starts to increase compared to those patients not receiving glucocorticoids? Does this differ by pathogen?

How often should people with AIIRD receiving anti-rheumatic therapies be re-screened for chronic and opportunistic infections?

Is screening and prophylaxis for opportunistic and chronic infections in people with AIIRD receiving anti-rheumatic therapies cost-effective?

Tuberculosis

Should patients starting immunosuppressants (e.g cyclophosphamide) be screened routinely for latent TB?

Should patients starting antirheumatic therapies be screened for non-tuberculous mycobacteria? What is the most effective way to screen for these infections?

How often should patients who have already been tested for tuberculosis, be re-screened? In relation to that, is there a need to re-screen patients who switch bDMARDs or ts-DMARDs?

Hepatitis

When should hepatitis anti-viral treatment be started in people living with AIIRD commencing anti-rheumatic treatment found to be at risk of hepatitis reactivation?

For how long should hepatitis anti-viral prophylaxis be continued in patients at risk for hepatitis reactivation after anti-rheumatic treatment is stopped?

Should patients with chronic or resolved hepatitis B also be screened for hepatitis D?

Other viruses

Is it safe to treat people living with HIV with anti-rheumatic treatments?

When should anti-viral prophylaxis be considered in people with AIIRD who have recurrent Herpes Zoster infections?

Is post-exposure prophylaxis for patients non-immune to VZV who are exposed to VZV beneficial?

Should patients with AIIRD starting anti-rheumatic therapy be screened for cytomegalovirus (CMV)?

Pneumocystis jirovecii pneumonia (PCP)

Does the risk of PCP differ according to underlying AIIRD (e.g. giant cell arteritis, SLE, ANCA-associated vasculitis, etc)?

What is the added risk of PCP in patients treated with combination glucocorticoids/immunosuppressive therapies compared to those receiving glucocorticoids along?

What is the safest and most effective regimen for PCP prophylaxis?

How long should patients at risk for PCP receive prophylaxis?

Other pathogens

Does avoidance of certain foods (e.g. unpasteurised cheese) reduce the risk of opportunistic and severe infections in patients with AIIRD receiving anti-rheumatic treatments?

Should people with AIIRD starting anti-rheumatic therapies living in endemic areas be screened for *Leishmania*, *Histoplasma* or *Coccidioides*?

Should people with AIIRD starting anti-rheumatic therapies be screened for fungal infections?

Table 1. The EULAR recommendations for screening and prophylaxis of chronic and opportunistic infections in adults with autoimmune inflammatory rheumatic diseases

Overarching principles	LoE	GoR	LoA mean (SD)
A. The risk of chronic and opportunistic infections should be considered and discussed with all patients with AIIRD prior to treatment with csDMARDs, tsDMARDs, bDMARDs, immunosuppressants and/or glucocorticoids and reassessed periodically.	NA	NA	9.5 (1.0)

B. Collaboration between rheumatologists and other specialists including but not limited to infectious disease doctors, gastroenterologists, hepatologists and pulmonologists is important.	NA	NA	9.6 (0.8)
C. Individual risk factors should be considered in the decision for screening and prophylaxis of chronic and opportunistic infections and reassessed periodically.	NA	NA	9.8 (0.7)
D. National guidelines and recommendations, amongst other country/region-level factors pertaining to endemic infectious diseases, should be considered.	NA	NA	9.7 (0.8)

Recommendations

1. Screening for latent tuberculosis is recommended in patients prior to starting bDMARDs or tsDMARDs*. Screening should also be considered in patients with increased risk for latent tuberculosis prior to starting csDMARDs, immunosuppressants* and/or glucocorticoids (according to dose and duration).	2b 5*	B D*	9.5 (0.9)
2. Screening for latent tuberculosis should follow national and/or international guidelines and would typically include a chest X-ray* and Interferon-gamma release assay (IGRA) over tuberculin skin test (TST) where available.	2b 5*	B D*	9.5 (0.8)
3. Choice and timing of latent tuberculosis therapy should be guided by national and/or international guidelines. Special attention should be given to interactions with drugs commonly used to treat AIIRD.	5	D	9.3 (1.4)
4. All patients being considered for treatment with csDMARDs, bDMARDs, tsDMARDs*, immunosuppressants* and glucocorticoids (according to dose and duration) should be screened for HBV.	2a 2b*	C C*	9.1 (1.3)
5. Screening for chronic hepatitis C should be considered in patients prior to starting csDMARDs, bDMARDs, tsDMARDs*, immunosuppressants and glucocorticoids* (according to dose and duration). Screening is recommended for patients with elevated alanine aminotransferase (ALT) or those with known risk factors.	2b 5*	C D*	9.0 (1.3)
6. Screening for HIV is recommended prior to treatment with bDMARDs and should be considered prior to treatment with csDMARDs, tsDMARDs, immunosuppressants and glucocorticoids (according to dose and duration).	5	D	8.9 (1.6)
7. All patients commencing csDMARDs, bDMARDs, tsDMARDs, immunosuppressants and/or glucocorticoids (according to dose and duration) who are non-immune to VZV should be informed about post-exposure prophylaxis following contact with VZV.	5	D	8.9 (1.5)
8. Prophylaxis against PCP should be considered in patients with AIIRD in whom high doses of glucocorticoids are used, especially in combination with immunosuppressants* and depending on the risk-benefit ratio.	2b 5*	B D*	9.2 (1.1)

AIIRD: autoimmune inflammatory rheumatic diseases, bDMARDs: biologic DMARDs, csDMARDs: conventional synthetic disease modifying anti-rheumatic drugs, GoR: grade of recommendation, HBV: hepatitis B virus, HIV: human immunodeficiency virus, LoA: level of agreement, LoE: level of evidence, tsDMARDs: targeted synthetic DMARDs, NA: not applicable, PCP: pneumocystis pneumonia, SD (standard deviation), VZV: varicella zoster virus. *denotes separate LoE and GoR, where this is different from the rest of the statement

Figure legends

Figure-1

Typical screening for hepatitis B virus (HBV) status include HBsAg, anti-HBcore and anti-HBs. HBsAg-positive patients (HBV carriers) would benefit from prophylactic treatment and thus it is advised that they should be referred to hepatologist for anti-viral prophylactic treatment. For those who are anti-HBcore-positive and HBsAg-negative (resolved HBV), measurement of HBV-DNA and liver function tests at baseline and then regular monitoring is advised. If HBV reactivation is suspected, based on these tests, referral to hepatologist for anti-viral treatment is recommended. For high-risk patients (e.g. commencing treatment with anti-CD20 regimes) prophylactic treatment, irrespective of DNA levels might be considered.

‡ positive anti-HBs without positive HBsAg or anti-HBcore is consistent with prior vaccination. If all three (HBsAg, anti-HBcore, anti-HBs) are negative, means no previous exposure to HBV

* Consider referral for anti-viral prophylaxis for those commencing rituximab, having also low titers of anti-HBs. Risk is assessed on an individual basis

] HBV-reactivation: rise or appearance of HBV-DNA, or conversion from HBsAg-negative to HBsAg-positive

periodic: there are no data to specify the exact time at which re-screening for HBV-reactivation should be performed. However, every 3-6 months is the standard for many national guidelines. Risk factors and cost should also be considered

§ referral to hepatologists is also recommended

Abbreviation List

AIIRD: Autoimmune inflammatory rheumatic diseases

ALT: alanine aminotransferase

bDMARDs: biologic disease modifying antirheumatic drugs

csDMARDs: conventional synthetic disease modifying antirheumatic drugs

HBV: hepatitis B virus

HCV: hepatitis C virus

HIV: human immunodeficiency virus

IGRA: Interferon gamma release assay

LFT: liver function tests

OAP: overarching principles

PCP: Pneumocystis jirovecii pneumonia

SLE: systemic lupus erythematosus

SLR: systematic literature review

TB: tuberculosis

TF: Task force

TNF: tumor necrosis factor

TMP/SXZ: trimethoprim/sulfamethoxazole

tsDMARDs: targeted synthetic disease modifying antirheumatic drugs

TST: tuberculin skin test

VZV: varicella zoster virus

Contributors: All authors contributed and finally approved the current manuscript

Funding: The systematic literature review was funded as part of the EULAR Quality of Care Committee (CI118) project for the 2022 EULAR recommendations on the screening and prophylaxis of chronic and opportunistic infections. KLH is also supported by the NIHR Manchester Biomedical Research Centre.

Conflict of interests

GEF: Consulting fees/honoraria: Pfizer, Abbvie, Novartis, UCB, AENorasis, Janssen, Pharmaserv-Lilly

EN: speaker fees/honoraria: Celltrion, Pfizer, Sanofi, Gilead, Galapagos, AbbVie, Lilly, Fresenius. Research funding: Pfizer, Lilly.

MD: none

SSZ: consulting fees: UCB

DC: None

LA: has received consulting fees and/or research funding from Astra-Zeneca, GSK, Pfizer

FA: none

GMNB: has received consulting fees/honoraria as speaker from Gilead, ViiV Healthcare, Janssen, MSD, Roche, Moderna unrelated to this work

JWJB: none

PB: none

CAC: none

SGD: none

MK: none

KL: has received consulting/speaker fees from Pfizer, Viatris, Celltrion outside of the submitted work

ML: none

JM: none

GN: consulting fees/honoraria as speaker from AbbVie, Amgen, Boehringer Ingelheim, Janssen, Miltenyi Biotech, Lilly, Pfizer, Roche unrelated to this work

EP: consulting fees/honoraria as speaker from Bayer, Menarini, Grifols, Zambon, Pfizer, Chiesi, Teva, Shire, Shionogi, Inmed, Boehringer-Ingelheim, unrelated to this work.

JvR: none

MS: consulting fees: BMS, Boehringer-Ingheleim, Eli Lilly, Celltrion, Amgen, Pfizer, Janssen Cilag.

AS: has received consulting fees/honoraria as speaker from AbbVie, Amgen, BMS, Celltrion, Janssen, Lilly, MSD, Pfizer, Roche unrelated to this work

AdT: none

JG: speaker fees / honoraria: Abbvie, Galapagos, Gilead, Janssen, Lilly, Novartis, Pfizer, UCB. Research funding: Abbvie, Astrazeneca, Galapagos, Gilead, Gritstone, Janssen, Moderna, Novovax, Pfizer

KH: honoraria from Abbvie; grant income from Pfizer and BMS

References

1. Winthrop KL, Novosad SA, Baddley JW, et al. Opportunistic infections and biologic therapies in immune-mediated inflammatory diseases: consensus recommendations for infection reporting during clinical trials and postmarketing surveillance. *Ann Rheum Dis* 2015; **74**:2107-16.
2. Stamp LK, Hurst M. Is there a role for consensus guidelines for *P. jirovecii* pneumonia prophylaxis in immunosuppressed patients with rheumatic diseases? *J Rheumatol* 2010; **37**:686-8.
3. Park JW, Curtis JR, Lee EB. Response to: 'Can we prescribe TMP/SMX prophylaxis without any concerns equally for all patients with rheumatic disease?' by Suyama and Okada. *Ann Rheum Dis* 2019; **78**:e18.
4. Mecoli CA, Danoff SK. Pneumocystis jirovecii Pneumonia and Other Infections in Idiopathic Inflammatory Myositis. *Curr Rheumatol Rep* 2020; **22**:7.
5. Hsu HC, Chang YS, Hou TY, et al. Pneumocystis jirovecii pneumonia in autoimmune rheumatic diseases: a nationwide population-based study. *Clin Rheumatol* 2021; **40**:3755-63.
6. Mori S, Sugimoto M. Pneumocystis jirovecii Pneumonia in Rheumatoid Arthritis Patients: Risks and Prophylaxis Recommendations. *Clin Med Insights Circ Respir Pulm Med* 2015; **9**:29-40.
7. van der Heijde D, Aletaha D, Carmona L, et al. 2014 Update of the EULAR standardised operating procedures for EULAR-endorsed recommendations. *Ann Rheum Dis* 2015; **74**:8-13.
8. Cochrane Handbook for Systematic Reviews of Interventions. 6.3 ed: Cochrane.
9. Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *PLoS Med* 2009; **6**:e1000097.
10. Oxford centre for evidence-based Medicine—Levels of evidence March 2009 [accessed March 2022].
11. Isaacs JD, Burmester GR. Smart battles: immunosuppression versus immunomodulation in the inflammatory RMDs. *Ann Rheum Dis* 2020; **79**:991-93.
12. Machado PM, Lawson-Tovey S, Strangfeld A, et al. Safety of vaccination against SARS-CoV-2 in people with rheumatic and musculoskeletal diseases: results from the EULAR Coronavirus Vaccine (COVAX) physician-reported registry. *Ann Rheum Dis* 2021;
13. Strangfeld A, Schafer M, Gianfrancesco MA, et al. Factors associated with COVID-19-related death in people with rheumatic diseases: results from the COVID-19 Global Rheumatology Alliance physician-reported registry. *Ann Rheum Dis* 2021; **80**:930-42.
14. Fragoulis GE, Sipsas NV. When rheumatology and infectious disease come together. *Ther Adv Musculoskelet Dis* 2019; **11**:1759720X19868901.

15. Hsu CY, Ko CH, Wang JL, et al. Comparing the burdens of opportunistic infections among patients with systemic rheumatic diseases: a nationally representative cohort study. *Arthritis Res Ther* 2019; **21**:211.
16. Accortt NA, Lesperance T, Liu M, et al. Impact of Sustained Remission on the Risk of Serious Infection in Patients With Rheumatoid Arthritis. *Arthritis Care Res (Hoboken)* 2018; **70**:679-84.
17. Au K, Reed G, Curtis JR, et al. High disease activity is associated with an increased risk of infection in patients with rheumatoid arthritis. *Ann Rheum Dis* 2011; **70**:785-91.
18. Nikiphorou E, Santos EJF, Marques A, et al. 2021 EULAR recommendations for the implementation of self-management strategies in patients with inflammatory arthritis. *Ann Rheum Dis* 2021; **80**:1278-85.
19. Ritschl V, Stamm TA, Aletaha D, et al. 2020 EULAR points to consider for the prevention, screening, assessment and management of non-adherence to treatment in people with rheumatic and musculoskeletal diseases for use in clinical practice. *Ann Rheum Dis* 2020;
20. Smolen JS, Landewe RBM, Bijlsma JWJ, et al. EULAR recommendations for the management of rheumatoid arthritis with synthetic and biological disease-modifying antirheumatic drugs: 2019 update. *Ann Rheum Dis* 2020; **79**:685-99.
21. Toupin-April K, Decary S, de Wit M, et al. Endorsement of the OMERACT core domain set for shared decision making interventions in rheumatology trials: Results from a multi-stepped consensus-building approach. *Semin Arthritis Rheum* 2021; **51**:593-600.
22. Xiao X, Da G, Xie X, et al. Tuberculosis in patients with systemic lupus erythematosus-a 37-year longitudinal survey-based study. *J Intern Med* 2020;
23. Curtis JR, Xie F, Yang S, et al. Risk for Herpes Zoster in Tofacitinib-Treated Rheumatoid Arthritis Patients With and Without Concomitant Methotrexate and Glucocorticoids. *Arthritis Care Res (Hoboken)* 2019; **71**:1249-54.
24. Lertnawapan R, Totemchokchayakarn K, Nantiruj K, et al. Risk factors of Pneumocystis jirovecii pneumonia in patients with systemic lupus erythematosus. *Rheumatol Int* 2009; **29**:491-6.
25. Ektefaie Y, Dixit A, Freschi L, et al. Globally diverse Mycobacterium tuberculosis resistance acquisition: a retrospective geographical and temporal analysis of whole genome sequences. *Lancet Microbe* 2021; **2**:e96-e104.
26. Evangelatos G, Koulouri V, Iliopoulos A, et al. Tuberculosis and targeted synthetic or biologic DMARDs, beyond tumor necrosis factor inhibitors. *Ther Adv Musculoskelet Dis* 2020; **12**:1759720X20930116.
27. Winthrop KL, Harigai M, Genovese MC, et al. Infections in baricitinib clinical trials for patients with active rheumatoid arthritis. *Ann Rheum Dis* 2020; **79**:1290-97.
28. Winthrop KL, Park SH, Gul A, et al. Tuberculosis and other opportunistic infections in tofacitinib-treated patients with rheumatoid arthritis. *Ann Rheum Dis* 2016; **75**:1133-8.
29. Ji X, Hu L, Wang Y, et al. Risk of tuberculosis in patients with rheumatoid arthritis treated with biological and targeted drugs: meta-analysis of randomized clinical trials. *Chin Med J (Engl)* 2022; **135**:409-15.
30. Cantini F, Blandizzi C, Niccoli L, et al. Systematic review on tuberculosis risk in patients with rheumatoid arthritis receiving inhibitors of Janus Kinases. *Expert Opin Drug Saf* 2020; **19**:861-72.
31. Cohen SB, van Vollenhoven RF, Winthrop KL, et al. Safety profile of upadacitinib in rheumatoid arthritis: integrated analysis from the SELECT phase III clinical programme. *Ann Rheum Dis* 2020;
32. Brassard P, Lowe AM, Bernatsky S, et al. Rheumatoid arthritis, its treatments, and the risk of tuberculosis in Quebec, Canada. *Arthritis Rheum* 2009; **61**:300-4.
33. Long W, Cai F, Wang X, et al. High risk of activation of latent tuberculosis infection in rheumatic disease patients. *Infect Dis (Lond)* 2020; **52**:80-86.

34. Jick SS, Lieberman ES, Rahman MU, et al. Glucocorticoid use, other associated factors, and the risk of tuberculosis. *Arthritis Rheum* 2006; **55**:19-26.
35. Fragoulis GE, Constantinou CA, Sipsas NV, et al. Tuberculosis in inflammatory arthritis. Are biologics the only culprits? *Lancet Rheumatology* 2019; **(Accepted for publication)**
36. Brode SK, Jamieson FB, Ng R, et al. Increased risk of mycobacterial infections associated with anti-rheumatic medications. *Thorax* 2015; **70**:677-82.
37. Targeted tuberculin testing and treatment of latent tuberculosis infection. This official statement of the American Thoracic Society was adopted by the ATS Board of Directors, July 1999. This is a Joint Statement of the American Thoracic Society (ATS) and the Centers for Disease Control and Prevention (CDC). This statement was endorsed by the Council of the Infectious Diseases Society of America. (IDSA), September 1999, and the sections of this statement. *Am J Respir Crit Care Med* 2000; **161**:S221-47.
38. Prevention CfDCa, National Center for HIV/AIDS VH, STD, and TB Prevention, Elimination DoT. Latent tuberculosis infection: A guide for primary health 2020 [Publication Number 22-0468]. Available from: <https://www.cdc.gov/tb/publications/ltbi/default.htm> accessed 13/7/2022.
39. Cantini F, Niccoli L, Capone A, et al. Risk of tuberculosis reactivation associated with traditional disease modifying anti-rheumatic drugs and non-anti-tumor necrosis factor biologics in patients with rheumatic disorders and suggestion for clinical practice. *Expert Opinion on Drug Safety* 2019; **18**:415-25.
40. Balbi GGM, Machado-Ribeiro F, Marques CDL, et al. The interplay between tuberculosis and systemic lupus erythematosus. *Curr Opin Rheumatol* 2018; **30**:395-402.
41. Ruan Q, Zhang S, Ai J, et al. Screening of latent tuberculosis infection by interferon-gamma release assays in rheumatic patients: a systemic review and meta-analysis. *Clin Rheumatol* 2016; **35**:417-25.
42. Hsia EC, Schluger N, Cush JJ, et al. Interferon- γ release assay versus tuberculin skin test prior to treatment with golimumab, a human anti-tumor necrosis factor antibody, in patients with rheumatoid arthritis, psoriatic arthritis, or ankylosing spondylitis. *Arthritis Rheum* 2012; **64**:2068-77.
43. Maeda T, Banno S, Maeda S, et al. Comparison of QuantiFERON-TB Gold and the tuberculin skin test for detecting previous tuberculosis infection evaluated by chest CT findings in Japanese rheumatoid arthritis patients. *J Infect Chemother* 2011; **17**:842-8.
44. Matulis G, Jüni P, Villiger PM, et al. Detection of latent tuberculosis in immunosuppressed patients with autoimmune diseases: performance of a Mycobacterium tuberculosis antigen-specific interferon gamma assay. *Ann Rheum Dis* 2008; **67**:84-90.
45. Ruan Q, Zhang S, Ai J, et al. Screening of latent tuberculosis infection by interferon- γ release assays in rheumatic patients: a systemic review and meta-analysis. *Clin Rheumatol* 2016; **35**:417-25.
46. Sargın G, Şentürk T, Ceylan E, et al. TST, QuantiFERON-TB Gold test and T-SPOT.TB test for detecting latent tuberculosis infection in patients with rheumatic disease prior to anti-TNF therapy. *Tuberk Toraks* 2018; **66**:136-43.
47. Vassilopoulos D, Tsikrika S, Hatzara C, et al. Comparison of two gamma interferon release assays and tuberculin skin testing for tuberculosis screening in a cohort of patients with rheumatic diseases starting anti-tumor necrosis factor therapy. *Clin Vaccine Immunol* 2011; **18**:2102-8.
48. Jiang B, Ding H, Zhou L, et al. Evaluation of interferon-gamma release assay (T-SPOT.TB[™]) for diagnosis of tuberculosis infection in rheumatic disease patients. *Int J Rheum Dis* 2016; **19**:38-42.
49. Lee H, Park HY, Jeon K, et al. QuantiFERON-TB Gold In-Tube assay for screening arthritis patients for latent tuberculosis infection before starting anti-tumor necrosis factor treatment. *PLoS One* 2015; **10**:e0119260.

50. Marques CDL, Duarte ALBP, Barros de Lorena VM, et al. Attenuated response to PPD in the diagnosis of latent tuberculosis infection in patients with rheumatoid arthritis. *Revista Brasileira de Reumatologia* 2009; **49**:121-31.
51. B elard E, Semb S, Ruhwald M, et al. Prednisolone treatment affects the performance of the QuantiFERON gold in-tube test and the tuberculin skin test in patients with autoimmune disorders screened for latent tuberculosis infection. *Inflamm Bowel Dis* 2011; **17**:2340-9.
52. Cho H, Kim YW, Suh CH, et al. Concordance between the tuberculin skin test and interferon gamma release assay (IGRA) for diagnosing latent tuberculosis infection in patients with systemic lupus erythematosus and patient characteristics associated with an indeterminate IGRA. *Lupus* 2016; **25**:1341-8.
53. Escalante P, Kooda KJ, Khan R, et al. Diagnosis of latent tuberculosis infection with T-SPOT(®).TB in a predominantly immigrant population with rheumatologic disorders. *Lung* 2015; **193**:3-11.
54. Girlanda S, Mantegani P, Baldissera E, et al. ELISPOT-IFN-gamma assay instead of tuberculin skin test for detecting latent Mycobacterium tuberculosis infection in rheumatic patients candidate to anti-TNF-alpha treatment. *Clin Rheumatol* 2010; **29**:1135-41.
55. Gogus F, G unendi Z, Karakus R, et al. Comparison of tuberculin skin test and QuantiFERON-TB gold in tube test in patients with chronic inflammatory diseases living in a tuberculosis endemic population. *Clin Exp Med* 2010; **10**:173-7.
56. Hanta I, Ozbek S, Kuleci S, et al. Detection of latent tuberculosis infection in rheumatologic diseases before anti-TNF  therapy: tuberculin skin test versus IFN-  assay. *Rheumatol Int* 2012; **32**:3599-603.
57. Inanc N, Aydin SZ, Karakurt S, et al. Agreement between Quantiferon-TB gold test and tuberculin skin test in the identification of latent tuberculosis infection in patients with rheumatoid arthritis and ankylosing spondylitis. *J Rheumatol* 2009; **36**:2675-81.
58. Kim JH, Cho SK, Han M, et al. Factors influencing discrepancies between the QuantiFERON-TB gold in tube test and the tuberculin skin test in Korean patients with rheumatic diseases. *Semin Arthritis Rheum* 2013; **42**:424-32.
59. Klein M, Jarosov  K, Forejtov  S, et al. Quantiferon TB Gold and tuberculin skin tests for the detection of latent tuberculosis infection in patients treated with tumour necrosis factor alpha blocking agents. *Clin Exp Rheumatol* 2013; **31**:111-7.
60. Lee JH, Sohn HS, Chun JH, et al. Poor agreement between QuantiFERON-TB Gold test and tuberculin skin test results for the diagnosis of latent tuberculosis infection in rheumatoid arthritis patients and healthy controls. *Korean Journal of Internal Medicine* 2014; **29**:76-84.
61. M nguez S, Latorre I, Mateo L, et al. Interferon-gamma release assays in the detection of latent tuberculosis infection in patients with inflammatory arthritis scheduled for anti-tumour necrosis factor treatment. *Clin Rheumatol* 2012; **31**:785-94.
62. Paluch-Oles J, Magrys A, Koziol-Montewka M, et al. Identification of latent tuberculosis infection in rheumatic patients under consideration for treatment with anti-TNF-alpha agents. *Archives of Medical Science* 2013; **9**:112-17.
63. Park JH, Seo GY, Lee JS, et al. Positive conversion of tuberculin skin test and performance of interferon release assay to detect hidden tuberculosis infection during anti-tumor necrosis factor agent trial. *J Rheumatol* 2009; **36**:2158-63.
64. Pyo J, Cho SK, Kim D, et al. Systemic review: agreement between the latent tuberculosis screening tests among patients with rheumatic diseases. *Korean J Intern Med* 2018; **33**:1241-51.
65. Scrivo R, Sauzullo I, Mengoni F, et al. Mycobacterial interferon-gamma release variations during longterm treatment with tumor necrosis factor blockers: Lack of correlation with clinical outcome. *Journal of Rheumatology* 2013; **40**:157-65.

66. Tang I, So H, Luk L, et al. Comparison of single and dual latent tuberculosis screening strategies before biologic and targeted therapy in patients with rheumatic diseases: a retrospective cohort study. *Hong Kong Med J* 2020; **26**:111-19.
67. Vassilopoulos D. Should we routinely treat patients with autoimmune/rheumatic diseases and chronic hepatitis B virus infection starting biologic therapies with antiviral agents? Yes. *Eur J Intern Med* 2011; **22**:572-5.
68. Vassilopoulos D, Stamoulis N, Hadziyannis E, et al. Usefulness of enzyme-linked immunospot assay (Elispot) compared to tuberculin skin testing for latent tuberculosis screening in rheumatic patients scheduled for anti-tumor necrosis factor treatment. *J Rheumatol* 2008; **35**:1271-6.
69. Wu X, Chen P, Wei W, et al. Diagnostic value of the interferon-gamma release assay for tuberculosis infection in patients with Behcet's disease. *BMC Infectious Diseases* 2019; **19**
70. Xie X, Chen JW, Li F, et al. A T-cell-based enzyme-linked immunospot assay for tuberculosis screening in Chinese patients with rheumatic diseases receiving infliximab therapy. *Clin Exp Med* 2011; **11**:155-61.
71. So H, Yuen CS, Yip RM. Comparison of a commercial interferon-gamma release assay and tuberculin skin test for the detection of latent tuberculosis infection in Hong Kong arthritis patients who are candidates for biologic agents. *Hong Kong Med J* 2017; **23**:246-50.
72. Malaviya AN, Thakaran R, Rawat R, et al. Real life experience of a screening strategy for latent tuberculosis before treatment with biologicals in Indian patients with rheumatic diseases. *Indian Journal of Rheumatology* 2018; **13**:233-39.
73. Iwagaito S, Naniwa T, Maeda S, et al. A comparative analysis of two interferon- γ releasing assays to detect past tuberculosis infections in Japanese rheumatoid arthritis patients. *Mod Rheumatol* 2016; **26**:690-5.
74. Martin J, Walsh C, Gibbs A, et al. Comparison of interferon {gamma} release assays and conventional screening tests before tumour necrosis factor {alpha} blockade in patients with inflammatory arthritis. *Ann Rheum Dis* 2010; **69**:181-5.
75. Melath S, Ismajli M, Smith R, et al. Screening for latent TB in patients with rheumatic disorders prior to biologic agents in a 'high-risk' TB population: comparison of two interferon gamma release assays. *Rheumatol Int* 2014; **34**:149-50.
76. Kang J, Jeong DH, Yoo B, et al. The usefulness of routine chest radiograph examinations in patients treated with TNF inhibitors for inflammatory arthritis in South Korea. *Respir Med* 2018; **143**:109-15.
77. Bonfiglioli KR, Ribeiro ACM, Moraes JCB, et al. LTBI screening in rheumatoid arthritis patients prior to anti-TNF treatment in an endemic area. *International Journal of Tuberculosis and Lung Disease* 2014; **18**:905-11.
78. Busquets-Pérez N, Ponce A, Ortiz-Santamaria V, et al. How many patients with rheumatic diseases and TNF inhibitors treatment have latent tuberculosis? *Reumatol Clin* 2017; **13**:282-86.
79. Cerda OL, de Los Angeles Correa M, Granel A, et al. Tuberculin test conversion in patients with chronic inflammatory arthritis receiving biological therapy. *Eur J Rheumatol* 2019; **6**:19-22.
80. Chen DY, Shen GH, Hsieh TY, et al. Effectiveness of the combination of a whole-blood interferon-gamma assay and the tuberculin skin test in detecting latent tuberculosis infection in rheumatoid arthritis patients receiving adalimumab therapy. *Arthritis Rheum* 2008; **59**:800-6.
81. Goel N, Torralba K, Downey C, et al. Screening for acquired latent tuberculosis in rheumatoid arthritis patients on tumor necrosis factor inhibition therapy in Southern California. *Clin Rheumatol* 2020; **39**:2291-97.
82. Hatzara C, Hadziyannis E, Kandili A, et al. Frequent conversion of tuberculosis screening tests during anti-tumour necrosis factor therapy in patients with rheumatic diseases. *Ann Rheum Dis* 2015; **74**:1848-53.

83. He D, Bai F, Zhang S, et al. High incidence of tuberculosis infection in rheumatic diseases and impact for chemoprophylactic prevention of tuberculosis activation during biologics therapy. *Clin Vaccine Immunol* 2013; **20**:842-7.
84. Hejazi ME, Ahmadzadeh A, Khabbazi A, et al. Tuberculin skin test conversion in patients under treatment with anti-tumor necrotizing factor alpha agents. *BMC Infect Dis* 2020; **20**:464.
85. Kim HW, Kwon OC, Han SH, et al. Positive conversion of interferon- γ release assay in patients with rheumatic diseases treated with biologics. *Rheumatol Int* 2020; **40**:471-79.
86. Scrivo R, Sauzullo I, Mengoni F, et al. Mycobacterial interferon- γ release variations during longterm treatment with tumor necrosis factor blockers: lack of correlation with clinical outcome. *J Rheumatol* 2013; **40**:157-65.
87. Son CN, Jun JB, Kim JH, et al. Follow-up testing of interferon-gamma release assays are useful in ankylosing spondylitis patients receiving anti-tumor necrosis factor alpha for latent tuberculosis infection. *J Korean Med Sci* 2014; **29**:1090-3.
88. Thomas K, Hadziyannis E, Hatzara C, et al. Conversion and Reversion Rates of Tuberculosis Screening Assays in Patients With Rheumatic Diseases and Negative Baseline Screening Under Long-Term Biologic Treatment. *Pathog Immun* 2020; **5**:34-51.
89. Cuomo G, D'Abrosca V, Iacono D, et al. The conversion rate of tuberculosis screening tests during biological therapies in patients with rheumatoid arthritis. *Clin Rheumatol* 2017; **36**:457-61.
90. Aggarwal R, Manadan AM, Poliyedath A, et al. Safety of etanercept in patients at high risk for mycobacterial tuberculosis infections. *J Rheumatol* 2009; **36**:914-7.
91. Carmona L, Gómez-Reino JJ, Rodríguez-Valverde V, et al. Effectiveness of recommendations to prevent reactivation of latent tuberculosis infection in patients treated with tumor necrosis factor antagonists. *Arthritis Rheum* 2005; **52**:1766-72.
92. Hernandez-Cruz B, Sifuentes-Osornio J, Ponce-De-Leon Rosales S, et al. Mycobacterium tuberculosis infection in patients with systemic rheumatic diseases. A case-series. *Clinical and Experimental Rheumatology* 1999; **17**:289-96.
93. Hsia EC, Cush JJ, Matteson EL, et al. Comprehensive tuberculosis screening program in patients with inflammatory arthritides treated with golimumab, a human anti-tumor necrosis factor antibody, in Phase III clinical trials. *Arthritis Care Res (Hoboken)* 2013; **65**:309-13.
94. Kurt OK, Kurt B, Talay F, et al. Intermediate to long-term follow-up results of INH chemoprophylaxis prior to anti-TNF-alpha therapy in a high-risk area for tuberculosis. *Wien Klin Wochenschr* 2013; **125**:616-20.
95. Valls V, Ena J. Short-course treatment of latent tuberculosis infection in patients with rheumatic conditions proposed for anti-TNF therapy. *Clin Rheumatol* 2015; **34**:29-34.
96. Bray MG, Poulain C, Dougados M, et al. Frequency and tolerance of antituberculosis treatment according to national guidelines for prevention of risk of tuberculosis due to tumor necrosis factor blocker treatment. *Joint Bone Spine* 2010; **77**:135-41.
97. Hazlewood GS, Naimark D, Gardam M, et al. Prophylaxis for latent tuberculosis infection prior to anti-tumor necrosis factor therapy in low-risk elderly patients with rheumatoid arthritis: A decision analysis. *Arthritis Care and Research* 2013; **65**:1722-31.
98. Shen Y, Ma HF, Luo D, et al. The T-SPOT.TB assay used for screening and monitoring of latent tuberculosis infection in patients with Behçet's disease pre- and post-anti-TNF treatment: A retrospective study. *J Chin Med Assoc* 2019; **82**:375-80.
99. Sichletidis L, Settas L, Spyrtatos D, et al. Tuberculosis in patients receiving anti-TNF agents despite chemoprophylaxis. *Int J Tuberc Lung Dis* 2006; **10**:1127-32.
100. Thomas K, Vassilopoulos D. Infections in Patients with Rheumatoid Arthritis in the Era of Targeted Synthetic Therapies. *Mediterr J Rheumatol* 2020; **31**:129-36.

101. Watanabe A, Matsumoto T, Igari H, et al. Risk of developing active tuberculosis in rheumatoid arthritis patients on adalimumab in Japan. *Int J Tuberc Lung Dis* 2016; **20**:101-8.
102. Gómez-Reino JJ, Carmona L, Angel Descalzo M. Risk of tuberculosis in patients treated with tumor necrosis factor antagonists due to incomplete prevention of reactivation of latent infection. *Arthritis Rheum* 2007; **57**:756-61.
103. Chen YM, Liao TL, Chen HH, et al. Three months of once-weekly isoniazid plus rifapentine (3HP) in treating latent tuberculosis infection is feasible in patients with rheumatoid arthritis. *Ann Rheum Dis* 2018; **77**:1688-89.
104. Gaitonde S, Pathan E, Sule A, et al. Efficacy of isoniazid prophylaxis in patients with systemic lupus erythematosus receiving long term steroid treatment. *Ann Rheum Dis* 2002; **61**:251-3.
105. Shobha V, Chandrashekar S, Rao V, et al. Biologics and risk of tuberculosis in autoimmune rheumatic diseases: A real-world clinical experience from India. *Int J Rheum Dis* 2019; **22**:280-87.
106. Shobha V, Rao V, Desai A, et al. Prescribing patterns and safety of biologics in immune-mediated rheumatic diseases: Karnataka biologics cohort study group experience. *Indian Journal of Rheumatology* 2019; **14**:17-20.
107. Song YJ, Cho SK, Kim H, et al. Risk of Tuberculosis Development in Patients with Rheumatoid Arthritis Receiving Targeted Therapy: a Prospective Single Center Cohort Study. *J Korean Med Sci* 2021; **36**:e70.
108. Bourré-Tessier J, Arino-Torregrosa M, Choquette D. Increased incidence of liver enzymes abnormalities in patients treated with isoniazid in combination with disease modifying and/or biologic agents. *Clin Rheumatol* 2014; **33**:1049-53.
109. Vanhoof J, Landewe S, Van Wijngaerden E, et al. High incidence of hepatotoxicity of isoniazid treatment for tuberculosis chemoprophylaxis in patients with rheumatoid arthritis treated with methotrexate or sulfasalazine and anti-tumour necrosis factor inhibitors. *Ann Rheum Dis* 2003; **62**:1241-2.
110. Nam SH, Oh JS, Hong S, et al. Early discontinuation of tofacitinib in patients with rheumatoid arthritis co-treated with rifampin for latent tuberculosis. *Joint Bone Spine* 2020; **87**:475-79.
111. McAllister WA, Thompson PJ, Al-Habet SM, et al. Rifampicin reduces effectiveness and bioavailability of prednisolone. *Br Med J (Clin Res Ed)* 1983; **286**:923-5.
112. Myint A, Tong MJ, Beaven SW. Reactivation of Hepatitis B Virus: A Review of Clinical Guidelines. *Clin Liver Dis (Hoboken)* 2020; **15**:162-67.
113. Rondaan C, Furer V, Heijstek MW, et al. Efficacy, immunogenicity and safety of vaccination in adult patients with autoimmune inflammatory rheumatic diseases: a systematic literature review for the 2019 update of EULAR recommendations. *RMD Open* 2019; **5**:e001035.
114. Lin TC, Yoshida K, Tedeschi SK, et al. Risk of Hepatitis B Virus Reactivation in Patients With Inflammatory Arthritis Receiving Disease-Modifying Antirheumatic Drugs: A Systematic Review and Meta-Analysis. *Arthritis Care Res (Hoboken)* 2018; **70**:724-31.
115. Su J, Long L, Zou K. Antiviral prophylaxis for preventing reactivation of hepatitis B virus in rheumatic patients: a systematic review and meta-analysis. *Clin Rheumatol* 2018; **37**:3201-14.
116. Kalyoncu U, Emmungil H, Onat AM, et al. Current antiviral practice and course of Hepatitis B virus infection in inflammatory arthritis: a multicentric observational study (A + HBV study). *Eur J Rheumatol* 2015; **2**:149-54.
117. Kalyoncu U, Yonem O, Calguneri M, et al. Prophylactic use of lamivudine with chronic immunosuppressive therapy for rheumatologic disorders. *Rheumatol Int* 2009; **29**:777-80.
118. Lin WT, Chen YM, Chen DY, et al. Increased risk of hepatitis B virus reactivation in systemic lupus erythematosus patients receiving immunosuppressants: a retrospective cohort study. *Lupus* 2018; **27**:66-75.

119. Matsuzaki T, Eguchi K, Nagao N, et al. Hepatitis B virus reactivation in patients with rheumatoid arthritis: A single-center study. *Modern Rheumatology* 2018; **28**:808-13.
120. Mo YQ, Liang AQ, Ma JD, et al. Discontinuation of antiviral prophylaxis correlates with high prevalence of hepatitis B virus (HBV) reactivation in rheumatoid arthritis patients with HBV carrier state: a real-world clinical practice. *BMC Musculoskelet Disord* 2014; **15**:449.
121. Tan J, Zhou J, Zhao P, et al. Prospective study of HBV reactivation risk in rheumatoid arthritis patients who received conventional disease-modifying antirheumatic drugs. *Clin Rheumatol* 2012; **31**:1169-75.
122. Chen MH, Wu CS, Chen MH, et al. High Risk of Viral Reactivation in Hepatitis B Patients with Systemic Lupus Erythematosus. *Int J Mol Sci* 2021; **22**
123. Ming-Xu H, Chen M, Cai Y, et al. Clinical outcomes of low-dose leflunomide for rheumatoid arthritis complicated with Hepatitis B virus carriage and safety observation. *Pak J Med Sci* 2015; **31**:320-4.
124. Chen MH, Chen MH, Liu CY, et al. Hepatitis B Virus Reactivation in Rheumatoid Arthritis Patients Undergoing Biologics Treatment. *J Infect Dis* 2017; **215**:566-73.
125. Jeong W, Choe J, Song BC, et al. Effect of Low-Dose Corticosteroid Use on HBV Reactivation in HBsAg-positive Rheumatoid Arthritis Patients. *The Open Rheumatology Journal* 2021; **15**:39-46.
126. Chen LF, Mo YQ, Jing J, et al. Short-course tocilizumab increases risk of hepatitis B virus reactivation in patients with rheumatoid arthritis: a prospective clinical observation. *Int J Rheum Dis* 2017; **20**:859-69.
127. Giardina AR, Ferraro D, Ciccia F, et al. No detection of occult HBV-DNA in patients with various rheumatic diseases treated with anti-TNF agents: a two-year prospective study. *Clin Exp Rheumatol* 2013; **31**:25-30.
128. Kuo MH, Tseng CW, Lu MC, et al. Risk of Hepatitis B Virus Reactivation in Rheumatoid Arthritis Patients Undergoing Tocilizumab-Containing Treatment. *Dig Dis Sci* 2021:1-9.
129. Lan JL, Chen YM, Hsieh TY, et al. Kinetics of viral loads and risk of hepatitis B virus reactivation in hepatitis B core antibody-positive rheumatoid arthritis patients undergoing anti-tumour necrosis factor alpha therapy. *Ann Rheum Dis* 2011; **70**:1719-25.
130. Padovan M, Filippini M, Tincani A, et al. Safety of Abatacept in Rheumatoid Arthritis With Serologic Evidence of Past or Present Hepatitis B Virus Infection. *Arthritis Care Res (Hoboken)* 2016; **68**:738-43.
131. Vassilopoulos D, Apostolopoulou A, Hadziyannis E, et al. Long-term safety of anti-TNF treatment in patients with rheumatic diseases and chronic or resolved hepatitis B virus infection. *Ann Rheum Dis* 2010; **69**:1352-5.
132. Ye H, Zhang XW, Mu R, et al. Anti-TNF therapy in patients with HBV infection--analysis of 87 patients with inflammatory arthritis. *Clin Rheumatol* 2014; **33**:119-23.
133. Zingarelli S, Frassi M, Bazzani C, et al. Use of tumor necrosis factor-alpha-blocking agents in hepatitis B virus-positive patients: reports of 3 cases and review of the literature. *J Rheumatol* 2009; **36**:1188-94.
134. Ryu HH, Lee EY, Shin K, et al. Hepatitis B virus reactivation in rheumatoid arthritis and ankylosing spondylitis patients treated with anti-TNFalpha agents: a retrospective analysis of 49 cases. *Clin Rheumatol* 2012; **31**:931-6.
135. Wang ST, Tseng CW, Hsu CW, et al. Reactivation of hepatitis B virus infection in patients with rheumatoid arthritis receiving tofacitinib. *Int J Rheum Dis* 2021; **24**:1362-69.
136. Chen YM, Huang WN, Wu YD, et al. Reactivation of hepatitis B virus infection in patients with rheumatoid arthritis receiving tofacitinib: a real-world study. *Ann Rheum Dis* 2018; **77**:780-82.
137. Reddy KR, Beavers KL, Hammond SP, et al. American Gastroenterological Association Institute guideline on the prevention and treatment of hepatitis B virus reactivation during immunosuppressive drug therapy. *Gastroenterology* 2015; **148**:215-9; quiz e16-7.

138. Koutsianas C, Thomas K, Vassilopoulos D. Reactivation of hepatitis B virus infection in rheumatic diseases: risk and management considerations. *Ther Adv Musculoskelet Dis* 2020; **12**:1759720x20912646.
139. Sasadeusz J, Grigg A, Hughes PD, et al. Screening and Prophylaxis to Prevent Hepatitis B Reactivation: Other Populations and Newer Agents. *Clin Liver Dis* 2019; **23**:521-34.
140. Barone M, Notarnicola A, Lopalco G, et al. Safety of long-term biologic therapy in rheumatologic patients with a previously resolved hepatitis B viral infection. *Hepatology* 2015; **62**:40-6.
141. Fukuda W, Hanyu T, Katayama M, et al. Incidence of hepatitis B virus reactivation in patients with resolved infection on immunosuppressive therapy for rheumatic disease: a multicentre, prospective, observational study in Japan. *Ann Rheum Dis* 2017; **76**:1051-56.
142. Fukuda W, Hanyu T, Katayama M, et al. Risk stratification and clinical course of hepatitis B virus reactivation in rheumatoid arthritis patients with resolved infection: final report of a multicenter prospective observational study at Japanese Red Cross Hospital. *Arthritis Res Ther* 2019; **21**:255.
143. Laohapand C, Arromdee E, Tanwandee T. Long-term use of methotrexate does not result in hepatitis B reactivation in rheumatologic patients. *Hepatology International* 2015; **15**
144. Schwaneck EC, Krone M, Kreissl-Kemmer S, et al. Management of anti-HBc-positive patients with rheumatic diseases treated with disease-modifying antirheumatic drugs-a single-center analysis of 2054 patients. *Clin Rheumatol* 2018; **37**:2963-70.
145. Cantini F, Boccia S, Goletti D, et al. HBV Reactivation in Patients Treated with Antitumor Necrosis Factor-Alpha (TNF-alpha) Agents for Rheumatic and Dermatologic Conditions: A Systematic Review and Meta-Analysis. *Int J Rheumatol* 2014; **2014**:926836.
146. Mori S. Do low titers of antibody against hepatitis B surface antigen carry a risk of viral reactivation during immunosuppressive therapy for rheumatic diseases? *Journal of Rheumatology* 2012; **39**:1292-93.
147. Chen MH, Chen MH, Chou CT, et al. Low but Long-lasting Risk of Reversal of Seroconversion in Patients With Rheumatoid Arthritis Receiving Immunosuppressive Therapy. *Clin Gastroenterol Hepatol* 2020; **18**:2573-81 e1.
148. Urata Y, Uesato R, Tanaka D, et al. Prevalence of reactivation of hepatitis B virus replication in rheumatoid arthritis patients. *Mod Rheumatol* 2011; **21**:16-23.
149. Caporali R, Bobbio-Pallavicini F, Atzeni F, et al. Safety of tumor necrosis factor alpha blockers in hepatitis B virus occult carriers (hepatitis B surface antigen negative/anti-hepatitis B core antigen positive) with rheumatic diseases. *Arthritis Care Res (Hoboken)* 2010; **62**:749-54.
150. Charpin C, Guis S, Colson P, et al. Safety of TNF-blocking agents in rheumatic patients with serology suggesting past hepatitis B state: results from a cohort of 21 patients. *Arthritis Res Ther* 2009; **11**:R179.
151. Giannitti C, Lopalco G, Vitale A, et al. Long-term safety of anti-TNF agents on the liver of patients with spondyloarthritis and potential occult hepatitis B viral infection: an observational multicentre study. *Clin Exp Rheumatol* 2017; **35**:93-97.
152. Kim YJ, Bae SC, Sung YK, et al. Possible reactivation of potential hepatitis B virus occult infection by tumor necrosis factor-alpha blocker in the treatment of rheumatic diseases. *J Rheumatol* 2010; **37**:346-50.
153. Lau CS, Chia F, Dans L, et al. 2018 update of the APLAR recommendations for treatment of rheumatoid arthritis. *Int J Rheum Dis* 2019; **22**:357-75.
154. Fraenkel L, Bathon JM, England BR, et al. 2021 American College of Rheumatology Guideline for the Treatment of Rheumatoid Arthritis. *Arthritis Rheumatol* 2021; **73**:1108-23.
155. European Association for the Study of the Liver. Electronic address eee, European Association for the Study of the L. EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection. *J Hepatol* 2017; **67**:370-98.

156. Mok CC. Hepatitis B and C infection in patients undergoing biologic and targeted therapies for rheumatic diseases. *Best Pract Res Clin Rheumatol* 2018; **32**:767-80.
157. Terrault NA, Lok ASF, McMahon BJ, et al. Update on prevention, diagnosis, and treatment of chronic hepatitis B: AASLD 2018 hepatitis B guidance. *Hepatology* 2018; **67**:1560-99.
158. Chen YM, Chen HH, Huang WN, et al. Reactivation of hepatitis B virus infection following rituximab treatment in HBsAg-negative, HBcAb-positive rheumatoid arthritis patients: A long-term, real-world observation. *Int J Rheum Dis* 2019; **22**:1145-51.
159. Kuo MH, Tseng CW, Lee CH, et al. Moderate Risk of Hepatitis B Virus Reactivation in HBsAg(-)/HBcAb(+) Carriers Receiving Rituximab for Rheumatoid Arthritis. *Sci Rep* 2020; **10**:2456.
160. Watanabe T, Fukae J, Fukaya S, et al. Incidence and risk factors for reactivation from resolved hepatitis B virus in rheumatoid arthritis patients treated with biological disease-modifying antirheumatic drugs. *Int J Rheum Dis* 2019; **22**:574-82.
161. Tien YC, Yen HH, Li CF, et al. Changes in hepatitis B virus surface antibody titer and risk of hepatitis B reactivation in HBsAg-negative/HBcAb-positive patients undergoing biologic therapy for rheumatic diseases: a prospective cohort study. *Arthritis Res Ther* 2018; **20**:246.
162. Chen MH, Lee IC, Chen MH, et al. Abatacept is second to rituximab at risk of HBsAg reverse seroconversion in patients with rheumatic disease. *Ann Rheum Dis* 2021; **80**:1393-99.
163. Kato M, Atsumi T, Kurita T, et al. Hepatitis B virus reactivation by immunosuppressive therapy in patients with autoimmune diseases: risk analysis in Hepatitis B surface antigen-negative cases. *J Rheumatol* 2011; **38**:2209-14.
164. Karadag O, Kasifoglu T, Ozer B, et al. Viral hepatitis screening guideline before biological drug use in rheumatic patients. *Eur J Rheumatol* 2016; **3**:25-28.
165. Sebastiani M, Atzeni F, Milazzo L, et al. Italian consensus Guidelines for the management of hepatitis B virus infections in patients with rheumatoid arthritis. *Joint Bone Spine* 2017; **84**:525-30.
166. Brunasso AM, Puntoni M, Gulia A, et al. Safety of anti-tumour necrosis factor agents in patients with chronic hepatitis C infection: a systematic review. *Rheumatology (Oxford)* 2011; **50**:1700-11.
167. Costa L, Caso F, Atteno M, et al. Long-term safety of anti-TNF- α in PsA patients with concomitant HCV infection: a retrospective observational multicenter study on 15 patients. *Clin Rheumatol* 2014; **33**:273-6.
168. Parke FA, Reveille JD. Anti-tumor necrosis factor agents for rheumatoid arthritis in the setting of chronic hepatitis C infection. *Arthritis Rheum* 2004; **51**:800-4.
169. Peterson JR, Hsu FC, Simkin PA, et al. Effect of tumour necrosis factor alpha antagonists on serum transaminases and viraemia in patients with rheumatoid arthritis and chronic hepatitis C infection. *Ann Rheum Dis* 2003; **62**:1078-82.
170. Gandhi NP, Manadan AM, Block JA. Retrospective Study of Patients on Etanercept Therapy for Rheumatic Diseases in Patients With Chronic Hepatitis C Virus. *J Clin Rheumatol* 2017; **23**:252-57.
171. Holroyd CR, Seth R, Bukhari M, et al. The British Society for Rheumatology biologic DMARD safety guidelines in inflammatory arthritis. *Rheumatology (Oxford)* 2019; **58**:e3-e42.
172. Sebastiani M, Milazzo L, Atzeni F, et al. Italian consensus recommendations for the management of hepatitis C infection in patients with rheumatoid arthritis. *Mod Rheumatol* 2019; **29**:895-902.
173. Chen MH, Chen MH, Tsai CY, et al. Incidence and antiviral response of hepatitis C virus reactivation in lupus patients undergoing immunosuppressive therapy. *Lupus* 2015; **24**:1029-36.
174. Lin KM, Cheng TT, Lin JC, et al. Tumor necrosis factor- α antagonist therapy for concomitant rheumatoid arthritis and hepatitis C virus infection: a case series study. *Clin Rheumatol* 2015; **34**:1039-46.
175. Caso F, Cantarini L, Morisco F, et al. Current evidence in the field of the management with TNF- α inhibitors in psoriatic arthritis and concomitant hepatitis C virus infection. *Expert Opin Biol Ther* 2015; **15**:641-50.

176. Mosca M, Tani C, Aringer M, et al. European League Against Rheumatism recommendations for monitoring patients with systemic lupus erythematosus in clinical practice and in observational studies. *Ann Rheum Dis* 2010; **69**:1269-74.
177. Visser K, Katchamart W, Loza E, et al. Multinational evidence-based recommendations for the use of methotrexate in rheumatic disorders with a focus on rheumatoid arthritis: integrating systematic literature research and expert opinion of a broad international panel of rheumatologists in the 3E Initiative. *Ann Rheum Dis* 2009; **68**:1086-93.
178. Cates M, Donati M, Gillet S, et al. Managing varicella zoster virus contact and infection in patients on anti-rheumatic therapy. *Rheumatology (Oxford)* 2018; **57**:596-605.
179. Winthrop KL, Tanaka Y, Lee EB, et al. Prevention and management of herpes zoster in patients with rheumatoid arthritis and psoriatic arthritis: a clinical review. *Clin Exp Rheumatol* 2022; **40**:162-72.
180. Guillevin L. Infections in vasculitis. *Best Pract Res Clin Rheumatol* 2013; **27**:19-31.
181. Lortholary O, Fernandez-Ruiz M, Baddley JW, et al. Infectious complications of rheumatoid arthritis and psoriatic arthritis during targeted and biological therapies: a viewpoint in 2020. *Ann Rheum Dis* 2020; **79**:1532-43.
182. Honda N, Tagashira Y, Kawai S, et al. Reduction of *Pneumocystis jirovecii* pneumonia and bloodstream infections by trimethoprim-sulfamethoxazole prophylaxis in patients with rheumatic diseases. *Scand J Rheumatol* 2021:1-7.
183. Park JW, Curtis JR, Kim MJ, et al. *Pneumocystis pneumonia* in patients with rheumatic diseases receiving prolonged, non-high-dose steroids-clinical implication of primary prophylaxis using trimethoprim-sulfamethoxazole. *Arthritis Res Ther* 2019; **21**:207.
184. Park JW, Curtis JR, Moon J, et al. Prophylactic effect of trimethoprim-sulfamethoxazole for *pneumocystis pneumonia* in patients with rheumatic diseases exposed to prolonged high-dose glucocorticoids. *Ann Rheum Dis* 2018; **77**:644-49.
185. Ogawa J, Harigai M, Nagasaka K, et al. Prediction of and prophylaxis against *Pneumocystis pneumonia* in patients with connective tissue diseases undergoing medium- or high-dose corticosteroid therapy. *Mod Rheumatol* 2005; **15**:91-6.
186. Vananuvat P, Suwannalai P, Sungkanuparph S, et al. Primary prophylaxis for *Pneumocystis jirovecii pneumonia* in patients with connective tissue diseases. *Semin Arthritis Rheum* 2011; **41**:497-502.
187. Wolfe RM, Peacock JE, Jr. *Pneumocystis Pneumonia* and the Rheumatologist: Which Patients Are At Risk and How Can PCP Be Prevented? *Curr Rheumatol Rep* 2017; **19**:35.
188. Katsuyama T, Saito K, Kubo S, et al. Prophylaxis for *Pneumocystis pneumonia* in patients with rheumatoid arthritis treated with biologics, based on risk factors found in a retrospective study. *Arthritis Res Ther* 2014; **16**:R43.
189. Meuli K, Chapman P, O'Donnell J, et al. Audit of *pneumocystis pneumonia* in patients seen by the Christchurch Hospital rheumatology service over a 5-year period. *Intern Med J* 2007; **37**:687-92.
190. Vela Casasempere P, Ruiz Torregrosa P, Garcia Sevilla R. *Pneumocystis jirovecii* in immunocompromised patients with rheumatic diseases. *Reumatol Clin (Engl Ed)* 2021; **17**:290-96.
191. Harada T, Kato R, Sueda Y, et al. The efficacy and safety of reduced-dose sulfamethoxazole-trimethoprim for chemoprophylaxis of *Pneumocystis pneumonia* in patients with rheumatic diseases. *Mod Rheumatol* 2020:1-7.
192. Takenaka K, Komiya Y, Ota M, et al. A dose-escalation regimen of trimethoprim-sulfamethoxazole is tolerable for prophylaxis against *Pneumocystis jirovecii pneumonia* in rheumatic diseases. *Mod Rheumatol* 2013; **23**:752-8.
193. Utsunomiya M, Dobashi H, Odani T, et al. An open-label, randomized controlled trial of sulfamethoxazole-trimethoprim for *Pneumocystis* prophylaxis: Results of 52-week follow-up. *Rheumatology Advances in Practice* 2020; **4**

194. Utsunomiya M, Dobashi H, Odani T, et al. Optimal regimens of sulfamethoxazole-trimethoprim for chemoprophylaxis of *Pneumocystis pneumonia* in patients with systemic rheumatic diseases: results from a non-blinded, randomized controlled trial. *Arthritis Res Ther* 2017; **19**:7.
195. Suyama Y, Okada M, Rokutanda R, et al. Safety and efficacy of upfront graded administration of trimethoprim-sulfamethoxazole in systemic lupus erythematosus: A retrospective cohort study. *Mod Rheumatol* 2016; **26**:557-61.
196. Wallace ZS, Choi H, Stone JH. Risk of severe infection following rituximab and the efficacy of antimicrobial prophylaxis. *Ann Rheum Dis* 2020; **79**:e40.
197. Suyama Y, Okada M. Can we prescribe TMP/SMX prophylaxis without any concerns equally for all patients with rheumatic disease? *Ann Rheum Dis* 2019; **78**:e17.
198. Sonomoto K, Tanaka H, Nguyen TM, et al. Prophylaxis against pneumocystis pneumonia in rheumatoid arthritis patients treated with b/tsDMARDs: Insights from 3,787 cases in FIRST registry. *Rheumatology (Oxford)* 2021;
199. Jinno S, Akashi K, Onishi A, et al. Comparative effectiveness of trimethoprim-sulfamethoxazole versus atovaquone for the prophylaxis of pneumocystis pneumonia in patients with connective tissue diseases receiving prolonged high-dose glucocorticoids. *Rheumatol Int* 2021;
200. Kitazawa T, Seo K, Yoshino Y, et al. Efficacies of atovaquone, pentamidine, and trimethoprim/sulfamethoxazole for the prevention of *Pneumocystis jirovecii* pneumonia in patients with connective tissue diseases. *J Infect Chemother* 2019; **25**:351-54.
201. Schmajuk G, Jafri K, Evans M, et al. *Pneumocystis jirovecii* pneumonia (PJP) prophylaxis patterns among patients with rheumatic diseases receiving high-risk immunosuppressant drugs. *Semin Arthritis Rheum* 2019; **48**:1087-92.
202. Landewe RBM, Kroon FPB, Alunno A, et al. EULAR recommendations for the management and vaccination of people with rheumatic and musculoskeletal diseases in the context of SARS-CoV-2: the November 2021 update. *Ann Rheum Dis* 2022;
203. Yates M, Watts RA, Bajema IM, et al. EULAR/ERA-EDTA recommendations for the management of ANCA-associated vasculitis. *Ann Rheum Dis* 2016; **75**:1583-94.
204. Fanouriakis A, Kostopoulou M, Alunno A, et al. 2019 update of the EULAR recommendations for the management of systemic lupus erythematosus. *Ann Rheum Dis* 2019; **78**:736-45.
205. Davies R, Dixon WG, Watson KD, et al. Influence of anti-TNF patient warning regarding avoidance of high risk foods on rates of listeria and salmonella infections in the UK. *Ann Rheum Dis* 2013; **72**:461-2.
206. Bradshaw MJ, Cho TA, Chow FC. Central Nervous System Infections Associated with Immunosuppressive Therapy for Rheumatic Disease. *Rheum Dis Clin North Am* 2017; **43**:607-19.
207. Orenstein R, Matteson EL. Opportunistic infections associated with TNF-alpha treatment. *Future Rheumatology* 2007; **2**:567-76.
208. Furer V, Rondaan C, Heijstek MW, et al. 2019 update of EULAR recommendations for vaccination in adult patients with autoimmune inflammatory rheumatic diseases. *Ann Rheum Dis* 2020; **79**:39-52.
209. Edelaar L, Nikiphorou E, Fragoulis GE, et al. 2019 EULAR recommendations for the generic core competences of health professionals in rheumatology. *Ann Rheum Dis* 2020; **79**:53-60.

