



Article  
scientifique

Revue de la  
littérature

1982

Published  
version

Open  
Access

This is the published version of the publication, made available in accordance with the publisher's policy.

---

## Highly active nitro-aromatic antiparasitic drugs

---

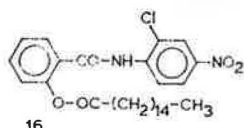
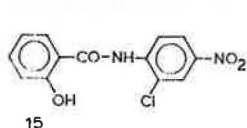
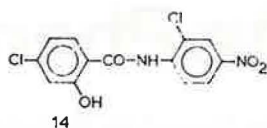
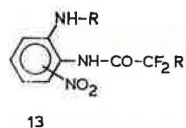
Jefford, Charles; Cadby, Peter Alan; Smith, L.C.; Pipe, David Frédéric

### How to cite

JEFFORD, Charles et al. Highly active nitro-aromatic antiparasitic drugs. In: Die Pharmazie, 1982, vol. 37, n° 6, p. 395–402.

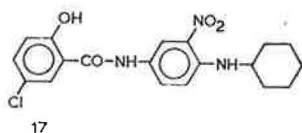
This publication URL: <https://archive-ouverte.unige.ch/unige:168563>

## 395



A large number of nitroanilides such as 13 [6] show anthelmintic properties and these are most pronounced for nitroanilides of halogenated benzoic acids. For instance, niclosamide (14) is the drug of choice for the treatment of tapeworm infections [17, 51], being effective against *Taenia saginata*, *T. solium*, *H. nana* and *Dipyllobothrium latum* as well as the intestinal flukes *Heterophyes heterophyes* and *Metagonimus yokogawai* [51]. It is interesting to note that the dechlorinated niclosamide (15) is quite inactive unless compounded with the surfactant, sodium dioctylsulphosuccinate, which gave it powerful anthelmintic activity against a large number of parasites with exceptional activity against *H. nana* [10]. Similarly, the introduction of a long-chain fatty acid ester group such as the palmitoyl derivative 16 greatly enhances its anthelmintic activity.

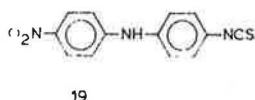
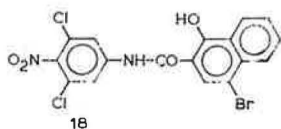
In an attempt to elucidate a structure/activity relationship in this large class, Singh and co-workers [96] synthesized a number of niclosamide analogues and found that the cyclohexylamino derivative 17 was the best cestodicidal agent with a particularly high therapeutic index (166.6) against *H. nana*. On the



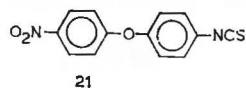
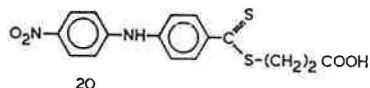
basis of further structural studies [97], he concluded that cestodicidal activity is not only dependent on the higher electron density of the un-nitrated ring, but is also related to the steric interactions between substituents at the 3- and 4-positions of that ring.

Other more profound structural modifications (eg. 18) [32] are also found to increase anthelmintic activities in certain cases.

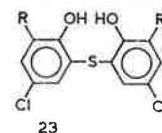
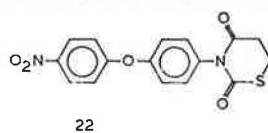
Heteroatom-bridged bi-aryls constitute another sub-class of anthelmintic nitrobenzenes. The best known N-bridged anthelmintic is the thiocyanate 19 (amoscanate: C 9333-Go/CGP-4540) which has activity against cestodes such as *Hymenolepis diminuta* in rats and *H. nana* in mice [68], nema-



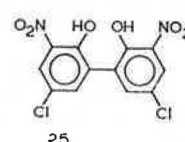
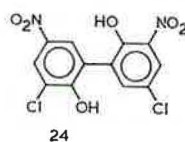
todes such as the hookworms *Ancylostoma duodenale* and *Necator americanus* [106], the whipworm *Trichuris trichiura* [106], the roundworm *Ascaris lumbricoides* [106] and *Nematostomoides dubius* [28, 92]. Schistosomicidal activity of this drug has been demonstrated against *Schistosoma haematobium*, *S. mansoni* in mice [99] and *S. japonicum* in dogs [99], mice [99, 118] and in rabbits and *in vitro* [118]. Furthermore, effective destruction of both micro- and macro-filariae (*Litosomoides carinii* [99, 100], *Dipetalonema viteae* [99, 100] and *Brugia pahangi* [89, 90]) has been reported in mongolian jirds. Preliminary studies of structural analogues of amoscanate (19) such as its thio-esters 20 indicate anthelmintic activity in mice [108].



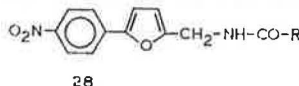
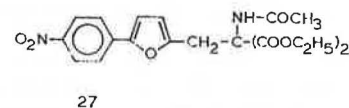
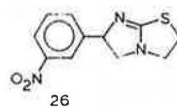
The O-bridged analogue of amoscanate is the thiocyanate ether 21 (nitroscanate) showing a broad spectrum of anthelmintic activity in domestic animals [16]. It is also used to eradicate nematodes and cestodes in dogs [15]. Recent reports [83] indicate that this drug eradicates the microfilariae of *Brugia malayi* and *Dipetalonema viteae* in the multimammate rat and the microfilariae of *Litosomoides carinii* in the cotton rat as well as both macro- and microfilariae of *D. viteae* (in jirds) and *L. carinii* (jirds and multimammate rats) [35]. Other O-bridged analogues 22 have shown anthelmintic properties [107] as have S-bridged dinitrophenols 23 [105].



Directly linked bi-aryls are also known to have anthelmintic activity such as the cattle fasciolide niclofolan 24 [65]. Its isomer meniclopholan 25 eradicates trematodes especially *Paragonimus*

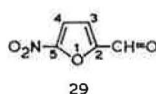


uterobilateralis from humans [111]. Some nitrobenzenes directly linked to heterocycles are also anthelmintic such as nitramisole (26) which is microfilaricidal to *B. malayi*, *D. viteae* and *L. carinii* in multimammate rats [83], the nitrophenylfuran 27 which eradicates *S. obvelata* from mice [78] and 28 which at 100 mg/kg b.i.d. results in 80–90% reduction of *Ascaris* serum in mice [112].



## 2.2. Nitrofurans

The second major class of anthelmintic nitro-aromatics comprises the nitrofurans. It is worthy of note that *without exception* the compounds investigated have always been derivatives of 5-nitrofuran-2-aldehyde (29). Of the various substitution

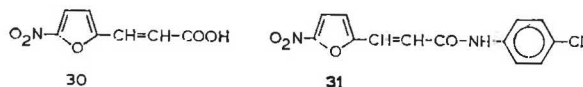


patterns possible, this 2,5-disubstitution is the most readily achieved by classical synthetic methods. (This may well be a major shortcoming of chemotherapeutic studies on the nitrofurans. Clearly, the synthesis of 3- and 4-substituted compounds, using recent methods, would provide at the very



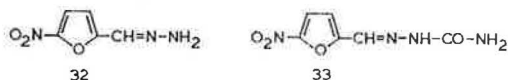
least important insights into structure/activity relationships in this area.)

A much investigated sub-class of nitrofurans has been the derivatives of 5-nitro-2-furanacrylic acid (30) and an extensive study of the substituted amides, anilides and phthalides of this acid have shown that while these compounds are inactive *in vitro*, many exhibit good anthelmintic activity *in vivo* [58]. The most useful of these proved to be the chloroanilide 31

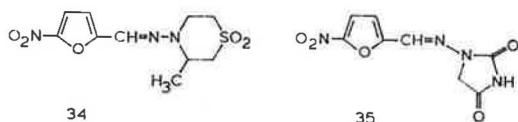


which was active against *Ancylostoma duodenale* and *Trichocephalus trichinosis* without undesirable side-effects [58]. A further 107 amides derived from 30 were synthesized and tested for their action against *Schistosoma japonicum*. Of the 33 compounds with activity, the two most active were the N-isopropyl and N-cyclohexyl derivatives [119]. Other amides have also been found to cure infections of *Schistosoma mansoni* in mice [50]. n-Alkyl esters of 30 have also shown exceptional anthelmintic activity and eradicate *Syphacia obvelata* from animals [87].

Perhaps the most commonly investigated types of nitrofuran are the derivatives of nitrofurfuralhydrazone (32). One of the simplest of these is nitrofural (33; nifurcin, furacin) which is active against the agent of Chagas' disease, *Trypanosoma cruzi* [91].

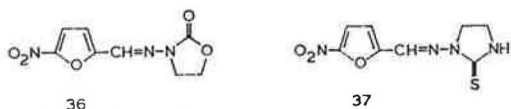


Another widely used hydrazone is nifurtimox (34; Lampit®) which shows promise as a trypanocide and filaricide [27, 111]. This drug eradicates *T. cruzi* [42] as well as offering protection against infection by this parasite [21] and is also active *in vitro*



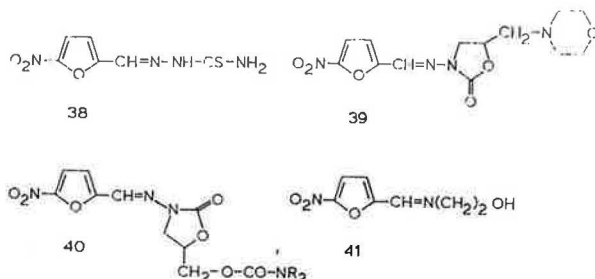
and *in vivo* against *T. rhodesiense* and *T. lewisi* [113]. It has been noted that nifurtimox decreased the number of parasites in the liver and spleen as well as the bloodstream of *T. cruzi* [110]. Both macro and microfilariæ of *Leishmaniasis* *carinii* are destroyed by nifurtimox in the multimammate rat [27, 83], the Chinese hamster [83] and cotton rats [83] as are both forms of *Dipetalonema viteae* in rodents [27], whereas only microfilariæ of *L. carinii* are destroyed in the jird [83]. Nitrofurantoin (35) finds use as a veterinary anthelmintic [58] and shows promise as a filaricide [27] in destroying micro and macrofilariæ of *L. carinii* in the multimammate rat [27, 83] and jird [83] but only macrofilariæ of *Brugia malayi* and *Dipetalonema viteae* in multimammate rats [83]. Unfortunately, no activity is observed against either form of the agent of river blindness, *Onchocerca volvulus* [27]. Pyridoxine hydrochloride has been added to nitrofurantoin to increase the compatibility of the gastrointestinal tract for the latter [40].

An analogous hydrazone, furazolidone (36) also shows promising filaricidal activity by destroying microfilariæ of *B. malayi* in the multimammate rat, macrofilariæ of *L. carinii* in jirds as well as both forms of *L. carinii* and *D. viteae* in multi-

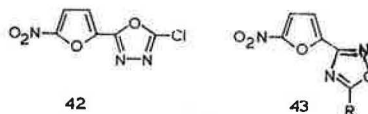


mammate rats and *L. carinii* in hamsters and cotton rats [83]. This drug also immobilizes the trypanosomes *T. cruzi* *in vitro* [38] and *T. gambiense* in mice [77]. Thiofuradene (37) has been

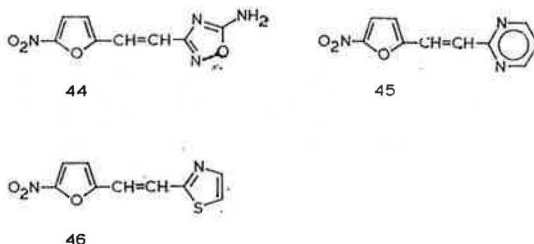
found to be a useful veterinary anthelmintic against the roundworm *Ascaridia galli* [66]. Benazone VII (38) is effective against *T. cruzi* *in vivo* and *in vitro* [35] and the macrofilariæ of *L. carinii* [83]. Furaltadone (39) and the related carbamate 40 [50] both display trypanocidal activity.



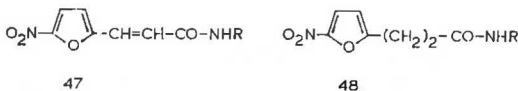
Another sub-class of active nitrofurans is formed by the Schiff-base derivatives of the aldehyde 29 of which 41 has known trypanocidal properties [38]. The same methylene-imino function is also shared by the more complicated heterocyclic anthelmintics 42 [44] and 43 [18].



Nitrofurans linked to another heterocycle by a trans-vinyl bridge can also show anthelmintic activity and of these SQ 18,506 (44) has been the most studied, displaying proven activity against the blood fluke *Schistosoma mansoni* in mice [84] and Chagas' disease (*T. cruzi*) [94, 95]. The activity of 44 is superior to that of nifurtimox 34 against *T. cruzi* and to that of simplified analogues 45 and 46 [48].

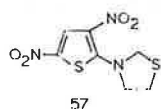
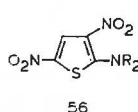
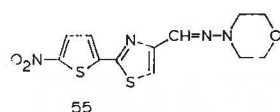
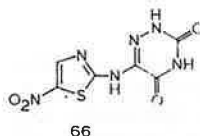
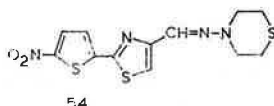
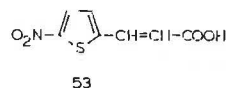
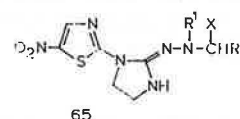
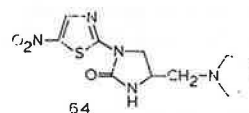
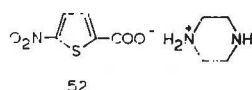
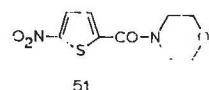
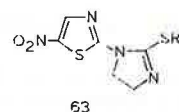
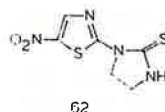
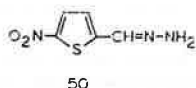
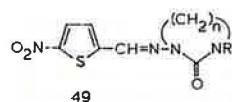


It is clear then that anthelmintic nitrofurans can possess a wide variety of side-chain structures. In all cases, however, the furfuryl carbon atom is always unsaturated and it has been suggested that the  $sp^2$  (planar) nature of this carbon atom is critical for activity. It is found, in keeping with this prediction, that the acrylamides 47 show good anthelmintic activity, but their saturated analogues 48 are inactive [14].



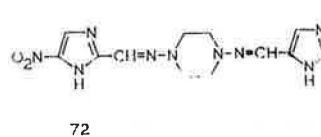
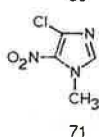
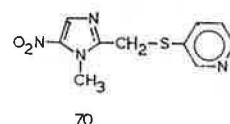
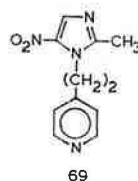
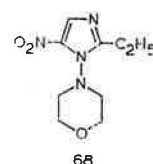
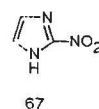
### 2.3. Nitrothiophenes

By complete analogy with the nitrofurans, a wide range of nitrothiophenes has also shown anthelmintic activity. A number of heterocyclic hydrazones 49 have been examined for activity against *Schistosoma mansoni* in mice and in general the greatest activity has been found for side-chains containing a six-membered heterocycle (49:  $n = 3$ ) with a small apolar N-substituent (49: R = H,  $C_2H_5$ , etc.) [49]. Similarly, studies with mice infected with *Syphacia obvelata* have shown that nitrothiophenes of quite diverse structure (eg. 50–53) show high activity [22]. Nitrothiophenes with thiazole side-chains have also been examined for trypanocidal activity in mice [109].

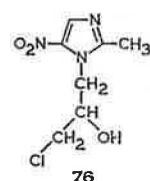
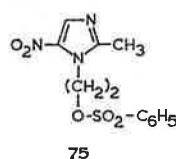
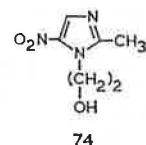
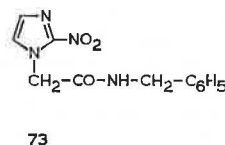


## 2.5. Nitroimidazoles

A large number of 5- and 2-nitroimidazoles have been developed as antitrichomonal and antiamoebic agents. These include azomycin (67), nimorazole (68), panidazole (69), pirinidazole (70), chloronidazole (71) and Hoe 316 (72). Unfortunately, they have not yet been closely examined for anthelmintic properties [34, 90, 111]. Those which demonstrate anthelmintic activity



can be divided into two sub-classes depending on whether they bear the major substituent on N-1 or on C-2. Of the N-substituted nitroimidazoles, Ro 7-1051 (73) is effective against Chagas' disease (T. cruzi) in mice [2, 42, 80] and *in vitro* [42, 80]. Metronidazole (74) shows only limited trypanocidal



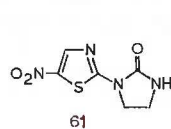
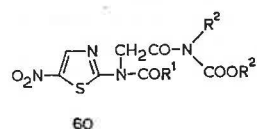
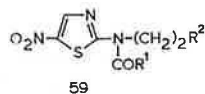
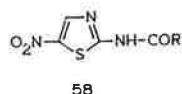
dal activity against T. cruzi [42], but shows promise as macrofilaricide against L. carinii [83] and B. pahangi [111]. Tinidazole (75) is active against B. pahangi [111] and ornidazole (76) is a possible microfilaricide of L. carinii [83].

Strangely, the best agent against T. rhodensiense 54 was one of the worst against T. cruzi whereas the best against the latter trypanosome 55 was one of the worst against T. rhodensiense [109].

Dinitrothiophenes 56 have recently been shown to have schistosomicidal activity [47]. Hoffmann-La Roche have recently prepared one of these, Ro 11-0761 (57) which is claimed to be effective against the schistosomes S. mansoni, S. haematobium and S. japonicum in a variety of laboratory animals [98].

## 2.4. Nitrothiazoles

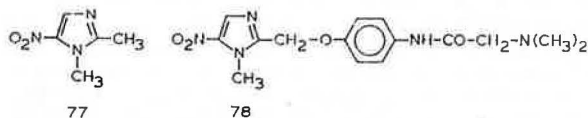
Unlike the nitrofurans, most of the active nitrothiazoles have a nitrogen substituent in the 2-position and the simplest of these are the amides aminotriazole (58, R = CH3), active against S. mansoni [5] and its aryl analogues (58: R = Ar) [23]. More complicated amides such as 59 (R1 = C6H7, NH2, R2 = CONH2, CH) [52] and 60 (R1 = H, CH3, R2 = CH3, C2H5, etc.) [63] are active against S. mansoni and Trichomonas vaginalis.



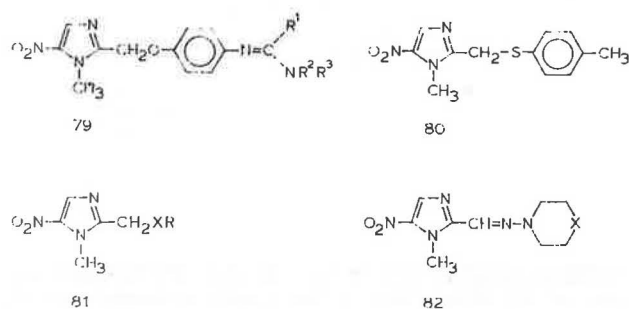
A large number of anthelmintic nitrothiazoles have a nitrogen heterocycle as 2-substituent. The best known of these is niridazole (61) which is of particular interest because of its microfilaricidal activity, especially against Onchocerca volvulus in humans [27, 102] and because of its utility as a schistosomicide [90] against S. haematobium and S. mansoni in humans [51]. It is also used to treat infections of guinea worm (Dracunculus medinensis) [51].

Several derivatives of niridazole (61) have been investigated and the thione 62 and thionamine 63 are also found to show activity in animals [102]. Condensation of niridazole with various reagents gives rise to 64 [3] and 65 [4] which have been found to show anthelmintic activity. The compound 66 which has a nitrogen substituent in the 2-position substituted by a nitrogen heterocycle has been shown to kill 80% of Schistosoma mansoni at a dose of 35 mg/kg per d [71].

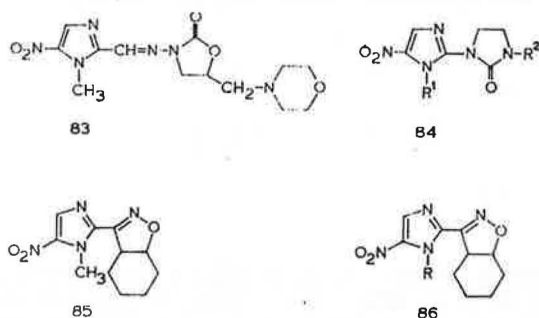
Dimetridazole (77) which is equally substituted in both positions, destroys macrofilariae of *L. carinii* in the multimammate rat [83]. Of the nitroimidazoles bearing their major substituent on C-2, the glycylamide (78) was found to be the most active



of 105 aryl ethers tested against *Trypanosoma brucei* in mice [116]. Anthelmintic activity has been claimed for an analogous ether (79) [7], while the thioether 80 has been shown to be markedly active against trypanomastigotes amastigotes of *T. cruzi* even during the manifest phase of Chagas' disease in mice [81]. In one case, 190 compounds of type 81 ( $X = N, S$ ) were tested for nematocidal activity, but few showed any activity [115]. In another case, 135 derivatives of 81 were tested, but only a few showed an *in vitro* effect on *Trypanosoma cruzi* and *T. brucei* and there required high doses [114]. On

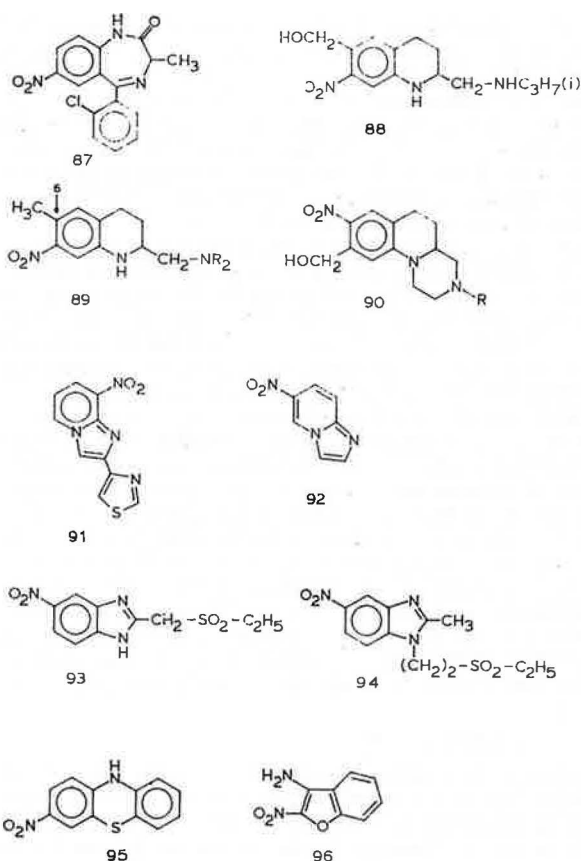


a more positive note, the hydrazones (82;  $X = O, SO_2$ , etc.) showed better activity against *T. cruzi* in mice than did the corresponding furans, thiophenes and thiazoles [76]. An analogous compound, moxnidazole (83) inactivates *D. viteae* and *L. carinii* in rats [83]. There have also been several reports of antischistosomal activity of the imidazolidinones (84) ( $R^1 = \text{alkyl, alkoxy, } R^2 = \text{carboxy, carbothioxy or sulphony}$ ) [57, 73, 74]. Short term treatment with the isoxazolone (85) and analogues has been shown to be effective against acute and chronic infections induced with a wide range of *T. cruzi* substrains [55, 69]. Use of berenil and the isoxazolone (86;  $R = H$  or alkyl) at a dose of 40 mg/kg i.p. cured 10/10 mice which had been infected with *T. brucei* (TREU 66711) for 21 d [54].



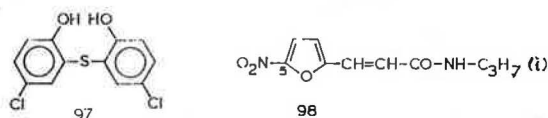
## 2.6. Polycyclic Nitro-aromatics

The most promising anthelmintics of this class of nitro-aromatics are the clonazepam (87; Ro 11-3128) which is schistosomicidal (*S. mansoni*, *S. haematobium*, but not *S. japonicum*) in various laboratory animals [103], and oxamniquine (88; Mansil®) which is the best current anthelmintic for *Schistosoma mansoni* [1, 103]. Other schistosomicides related to oxamniquine are the 6-methyl analogue 89 [81, 37] and the tricyclic analogue 90 [9]. The imidazole pyridines 91 and 92 are active against *Trichostrongyles in vitro* [33] and the benzimidazoles 93 and 94 show activity against *Nematospirides dubius* in mice [46].



## 3. The Importance of the Nitro Group

Although there are exceptions [59], as a general rule the presence of a nitro substituent is the one immutable structural characteristic of nitro-aromatic anthelmintics that is required for activity. For instance, bithional (97) is a useful antimicrobial and fungicide, but shows only weak anthelmintic activity [93]. On the other hand, its dinitro derivative 23 is a powerful trematocide [105]. When other nitro-aromatic anthelmintics such as niclosamide (14) are reduced to their corresponding amino compounds, it is found that their activity is either diminished or lost entirely [101]. In other cases, analogues



of nitro-aromatics, where the nitro group is replaced by another electron-withdrawing substituent (such as the 5-cyano and 5-carboxy analogues of the isopropylamide 98 [85] and the 5-chloro derivative of 93 [46]) are found to be inactive.

Other polycyclic nitro-aromatic anthelmintics are the phenothiazine 95 [11] and the benzofuran 96 [82].

There are, of course, many effective anthelmintics possessing no nitrosubstituent [1, 17, 27, 34, 51, 90, 111]. Nonetheless, it does appear that the nitro-aromatics constitute a special class in which the nitro group is essential for anthelmintic activity.

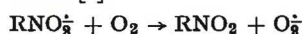
Very little is known of the mode of action of these drugs. It is evident that different anthelmintics produce diverse effects on different helminths. Niclosamide (14) for instance interferes with the respiration and blocks glucose uptake by tapeworms [51]. It appears to uncouple oxidative phosphorylation in both mammalian and taenoid mitochondria, but is therapeutically effective because the helminths depend on anaerobic metabolism of carbohydrates as their major source of energy [51].

The action of niridazole (61) on schistosomes is quite different. It appears to arrest egg-shell formation and to interfere with reproductive processes in the females. It also interferes with



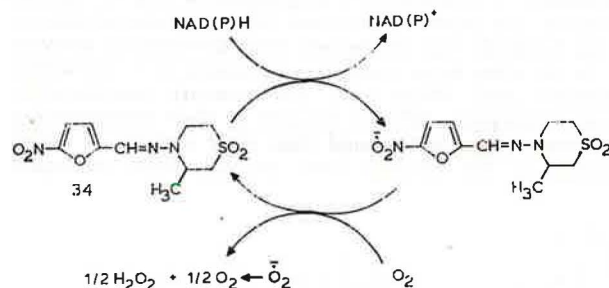
carbohydrate metabolism by inhibiting glucose adsorption and accelerating glycogen breakdown [27, 51]. On the other hand, the nitroimidazole 73 was found to inhibit protein and RNA synthesis in *Trypanosoma cruzi*, but had a negligible effect on DNA-synthesis or respiration [80]. In *T. cruzi* SQ 18506 (44) inhibited protein formation at the lowest effective concentration and may, therefore, be the primary mechanism of action [41]. Nitrophenolic anthelmintics are thought to interfere with phosphorylation processes in helminths [88].

It has been suggested that *in vivo* reduction of the nitro group to nitroso or hydroxylamino groups may be important [85] and the reduction potential of the nitro group should be an index for determining anthelmintic activity [60]. In character with anaerobic metabolic mechanisms, a key step in the uptake of nitro compounds could be their reduction by flavodoxins and ferredoxins [70]. This would have the dual effect of generating a gradient that drives the selective uptake of these drugs by helminths, and may also create metabolites that are more cytotoxic than the unreduced nitro compounds [70]. Alternatively, the partially reduced nitro-aromatics could generate other toxic species by further reactions. It has been suggested that radical anions (one electron reduction products  $\text{RNO}_2^{\cdot-}$ ) of nitro-aromatic and nitro-heterocyclic compounds are essential intermediates in the enzymatic reduction of these compounds to hydroxylamines or amines [61, 62, 63]. Oxygen inhibits nitroreductase activity in e. g. hepatic microsomal incubations and measurements of the rate of  $\text{O}_2$  uptake have provided evidence [61] that the inhibition results from the electron transfer reaction [1]:



Electron transfer to a nitro group and the consequent interference with the electron transport system has been postulated as the mechanism of action in nitrofurans [39]. The generation of superoxide ion has been postulated as the toxic principle in nitrofurans such as nitrofurantoin (35) [24, 79]. Addition of nifurtimox (34) to cultures of *Trypanosoma cruzi* has been shown to produce both superoxide ion and hydrogen peroxide (Scheme) [29].

#### Scheme



With NADH as reducing agent nifurtimox (34) induced release of superoxide anion, at a nifurtimox concentration closely parallel to the concentration reached in blood serum with a therapeutic dose of 15 mg/kg [30]. It is consistent with these theories of nitro reduction that enzyme preparations from cestodes catalyze the *in vitro* reduction of nitrophenol anthelmintics to the corresponding amino compounds [31].

#### 4. Structure and Activity

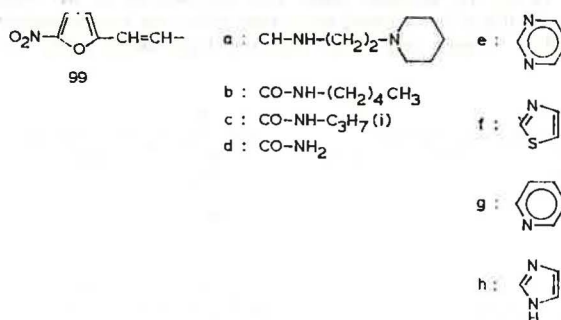
The interpretation of structure/activity relationships of the nitroaromatic anthelmintics is complicated by several factors. There are numerous cases where the activity of an anthelmintic varies enormously depending on the nature of the helminth. As yet, there is no anthelmintic which is therapeutically effective for the complete spectrum of helminth afflictions. Indeed, most of the drugs examined in this review are only active against one class of helminth (eg. schistosomes) or just one species (eg. *S. mansoni*) or worse still, one strain (eg. *S. mansoni* — St. Lucia). This is further complicated by host specificity. For example, certain drugs are filaricidal in one type of laboratory animal, but not in others [83]. Finally, interpretation of the published structure/activity studies is complicated by incomplete and unsystematic structural variation and by technical problems prohibiting consistent dosages, etc.

In the case of nitro-heterocyclic compounds however, there appears to be one simplifying factor. There is good reason to believe that these compounds may be considered as having *firstly*, a nitro-heterocyclic nucleus to which is attributable the toxicity of the drug and *secondly*, a sidechain which may simply modify the physico-chemical parameters affecting the absorption of the drug by host and parasite [76]. In general, these two functions may be considered as distinct and independent, although there may be certain limitations such as the requirement for an unsaturated furfuryl carbon atom in the side-chain of nitrofurans which would effect the reduction potential of the nitro group [76]. We have therefore analyzed the relationships of anthelmintic activity of each structural component separately.

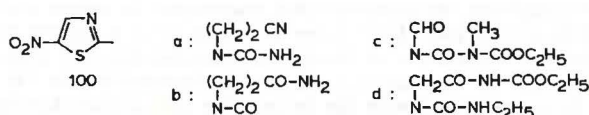
#### 4.1. The Effect of the Structure of the Side Chain on Activity

##### 4.1.1. Schistosomicides

Studies on *S. mansoni* in mice have shown that SQ 18,506 (44) shows the highest activity of the nitrofurans [85] while the related amides (99: a-d)<sup>1</sup> show decreasing activity as the N-

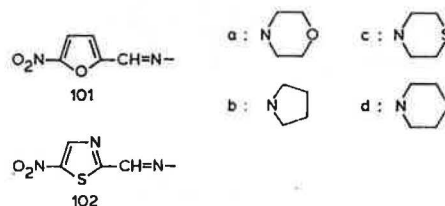


substituent becomes simpler (a → d) [50]. Heterocyclic analogues of SQ 18,506 (99e-h) have schistosomicidal activity that decreases in the order 99e = 99f > 99g > 99h [48]. No thiazoles were as effective as niridazole (61) and the activity of open-chain analogues 100 a-d against *S. mansoni* in mice decreases in alphabetical order 100 a → d [52, 53].

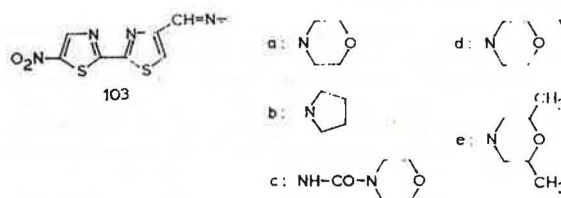


##### 4.1.2. Trypanocides

SQ 18,506 (44) is also the best nitrofuran for eradicating *T. cruzi* in mice, being better than nifurtimox (34) [42, 56], which in turn is better than the series 101 a-d which shows decreasing activity in alphabetical order [76]. Similarly, nitro-



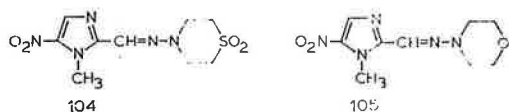
thiazines having the same side-chains (102a-d) show decreasing activity in the order 102a = 102c → 102b [76]. Trypanocidal activity of various nitrothiophene thiazoles 103<sup>2</sup>, however, depends on the species of protozoan tested. For example, in



<sup>1</sup> CO instead of CH in 99 a

<sup>2</sup> Nitrothiophen instead of nitrothiazol in 103

mice infected with *T. cruzi*, activity declines in alphabetical order 103a → e whereas for *T. rhodesiense* the order is 103d > 103b > 103a > 103e > 103c [109]. With nitroimidazoles, it was found that the aryl ether (78) was more effective against *T. brucei* in mice than Tinidazole (75) which in turn was more effective than Metronidazole (74) [116]. Against *T. cruzi* in mice, the sulphoxide 104 was superior to both its morpholine analogue 105 and nifurtimox (34) [76].



#### 4.1.3. Filaricides

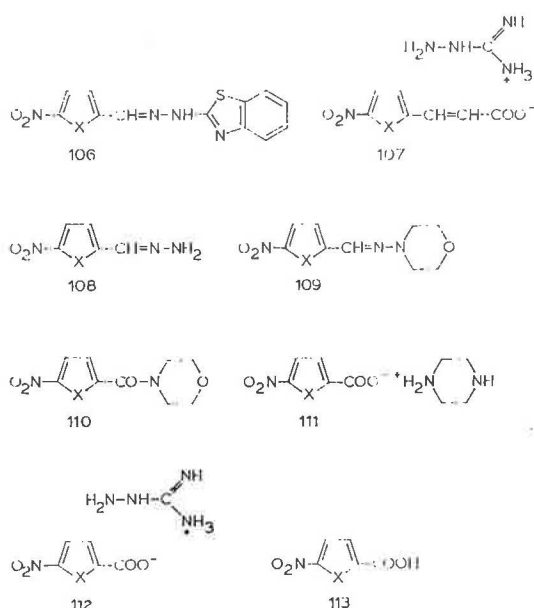
No quantitative or ranking studies have been undertaken. In the search for a macrofilaricide which has no effect on microfilariae, various nitro-heterocyclic compounds have been examined for their effect on *Litomosoides carinii* in a variety of laboratory animals [83], but no clear conclusions can be drawn as

- results depend on the nature of the experimental protocol and the type of laboratory animal and
- experimental screening with animal parasites is not always relevant to the treatment of human parasitic diseases.

Furazolidone (36) kills adult worms in cotton rats and mongolian jirds, but some tests reveal simultaneous microfilaricidal activity. The same drug kills both stages of *L. carinii* in multimammate rats and chinese hamsters. Similarly, nifurtimox (34) kills both forms in hamsters and rats although some experiments reveal only macrofilaricidal activity. Nitrofurantoin (35) proves to be both micro- and macrofilaricidal in all studies whereas nitrofurantoin (33) and the nitroimidazoles metronidazole (74) and dimetridazole (77) kill only the adult forms in some animals but are completely inactive on other test animals [83].

#### 4.1.4. Nematocides

Examination of the effect of nitrofurans and thiophenes on *Syphacia obvelata* in mice reveals different side-chain ranking for the two heterocyclic nuclei [22]. Of the compounds tested for both heterocycles, the following rankings were found: for 106 (X = O: best nitrofuran, X = S: inactive), 107 (X = O: second nitrofuran, X = S: eighth nitrothiophene), 108 (X = O: inactive, X = S: best nitrothiophene), 109 (X = O: third,



X = S: inactive, 110 (X = O: inactive, X = S: second), 111 (X = O: sixth, X = S: third), 112 (X = O: fourth, X = S: seventh), 113 (X = O: fourth, X = S: inactive), etc.

#### 4.2. The Effect of the Nitro-substituted Heterocycle on Activity

Only a few drug-screening studies have been performed in which different nitroheterocycles having the same side-chain have been compared (2-R): 5-Nitrofurans, 5-Nitrothiophenes, 5-Nitrothiazoles, 5-Nitroimidazoles, 2-Nitroimidazoles.

##### 4.2.1. Schistosomicides

It has been reported that the antischistosomal activity of various nitrofurans acrylamides 47 is lost when the nitrofuran group is replaced by a nitrothiazine [60]. Other studies on mice infected with *Schistosoma mansoni* have shown that in general the nitrothiophenes are better than corresponding nitrofurans [59].

##### 4.2.2. Trypanocides

Although one study has ranked the nitrofurans, SQ 18,606 (44) and nifurtimox (34) as being more active against *T. cruzi* than the N-substituted imidazoles Ro 7-1051 (73: 2-nitro) and metronidazole (74: 5-nitro) [52], the only study in which all compounds shared the same side-chains showed that 5-nitroimidazoles were better than nitrofurans, nitrothiophenes and nitrothiazines in that order [76]. In general, 5-nitroimidazoles are more effective when they bear the side-chain on C-2 rather than N-1. It also appears that 2-nitroimidazoles may be more effective than either type of 5-nitroimidazole. In any case, the 2-nitroimidazoles have a different physiological mode of action than their 5-nitro counterparts [80].

##### 4.2.3. Nematocides

In general, nitrofurans have more effect than correspondingly substituted nitrothiophenes against *Syphacia obvelata* in mice [22].

Acknowledgement: One of us (D.P.) is grateful for financial support from the filariasis Scientific Working Group of the UNDP/World Bank/WHO Special Programme for Research and Training in Tropical Diseases.

#### References

- 1 Adolphe, M. (Ed.), Advanced Pharmacology and Therapy, Proc. 7th Int. Congr. Pharmacology, Pergamon, Oxford 1979
- 2 Andrade, S. G., and R. M. Figueira, Rev. Inst. Med. Trop. Sao Paulo 19, 335 (1977)
- 3 Ascher, G., and H. Reinshagen, Ger. Offen 2 603 820 (to Sandoz-Patent GmbH), Aug. 26, 1976
- 4 Ascher, G., and H. Reinshagen, Ger. Offen 2 604 118 (to Sandoz-Patent GmbH), Aug. 26, 1976
- 5 Ashton, C. C., A. B. Kupferberg and N. Millman, Antibiotics and Chemotherapy 5, 540 (1955)
- 6 Austrian P. 341 507 (to Eli Lilly & Co), Feb. 10, 1978
- 7 Austrian P. 351 021 (1979)
- 8 Baxter, C. A. R., and H. C. Richards, J. med. Chem. 14, 1033 (1971)
- 9 Baxter, C. A. R., and H. C. Richards, ibid. 15, 351 (1972)
- 10 Bekhti, A. F., E. A. Amvrosieva, S. I. Kharitonova, I. A. Krotov, D. G. Bayandina and I. S. Budanova, USSR P. 563 414, June 30, 1977
- 11 Belg. P. 611 115 (to Rhone-Poulenc SA), June 4, 1962
- 12 Belg. P. 878 013; C. A. 92, 215 057 j (1980)
- 13 Beretta, C., G. Gasparini, A. Locatelli and R. Faustini, Veterinaria [Milan] 21, 3 (1972)
- 14 Bueding, E., and J. Fischer, Molecular Pharmacol. 6, 532 (1970)
- 15 Boray, J. C., Proc. 54th Ann. Conf. Aust. Vet. Assoc., p. 123, Perth 1977
- 16 Boray, J. C., M. B. Strong, J. R. Allison, M. von Orelli, G. Sarasin and W. Gfeller, Austral. Vet. J. 55, 45 (1979)
- 17 Botero, R. D., Ann. Review Pharmacol. Toxicol. 18, 1 (1978)
- 18 Brit. P. 1 025 439 (to Abbott Laboratories), 1966
- 19 Brit. P. 1 449 540 (to Sarabashi Research Centre), Sept. 15, 1976
- 20 Cappello, B., V. Boido and F. Sparatore, Boll. ital. Biol. sperim. 53, 801 (1977)
- 21 Cardoni, R. L., M. T. Rimoldi and M. M. E. Deb Racco, Exp. Parasitol. 43, 370 (1977)
- 22 Cavier, R., J. Cenac, R. Royer and L. René, Chim. therap. 5, 270 (1970)
- 23 Cavier, R., P. Gayral, J. Guillaume and J. M. Clavel, Europ. J. med. Chem. chim. Therap. 13, 639 (1978)
- 24 Combs, Jr. G. F., Proc.-Cornell Nutr. Conf. Food Manuf. 9 (1979)
- 25 Davis, M., J. M. S. Lucas, J. Rosenbaum and D. E. Wright, Nature [London] 211, 882 (1966)
- 26 DeCarneri, I., and C. Vita, in: R. Cavier and F. Hawkins, (Eds.) International Encyclopedia of Pharmacology and Therapeutics, Sn 64 vol. 1, p. 9, Pergamon, Oxford 1973
- 27 DeGraw, J. I., and W. T. Colwell, Chemotherapy of Filariasis. A Review and Analysis for Future Research Directions, SWG Report on Filariasis, WHO, 1978



- 28 Dilloo, P., L. Baurokova, K. Dimitrov, D. Drumev, V. Dzhurov, V. Bratanov, I. Torlakov, A. Petkov and N. Naidenov, *MBI, Med. Biol. Inf.* 3, 33 (1979)
- 29 Docampo, R., and A. O. M. Stoppani, *Arch. Biochem. Biophysics* 197, 317 (1979)
- 30 Docampo, R., and A. O. M. Stoppani, *Medicina* 40 (Suppl. 1), 10 (1980)
- 31 Douch, P. G. C., and L. L. Buchanan, *Xenobiotica* 9, 467 (1979)
- 32 Dubey, S. K., A. K. Singh, H. Singh, S. Sharma, R. N. Lyer, J. C. Katiyar, P. Goel and A. B. Sen, *J. med. Chem.* 21, 1178 (1978)
- 33 Fisher, M. H., and A. Lusi, *ibid.* 15, 982 (1972)
- 34 Fisher, M. H., and C. C. Wang, in: F. H. Clarke (Ed.), *Annual Reports in Medicinal Chemistry*, vol. 13, p. 130, Academic Press, New York 1978
- 35 Follinova, P., L. Ebringer, A. Jurasek and A. Rybar, *Acta Fac. Rerum Nat. Univ. Comenianae, Microbiol.* 6, 35 (1978)
- 36 Forster, R., G. Pringle, D. F. King and J. Paris, *Ann. Trop. Med. Parasitol.* 63, 95 (1969)
- 37 Forster, R., E. T. Mesmer, B. L. Cheetman and D. F. King, *ibid.* 65, 221 (1971)
- 38 Freeman, F., P. L. Wilson and B. H. Kazan, *Exp. Parasitol.* 38, 181 (1975)
- 39 Gounot, A. M., C. Brevet and S. Videz, *Ann. Pharm.* 37, 421 (1979)
- 40 Groenig, R., *Dtsch. Apotheker-Ztg.* 119, 2086 (1979)
- 41 Gugliotta, J. L., H. B. Tanowitz, M. Wittner and R. Sociro, *Exp. Parasitol.* 49, 216 (1980)
- 42 Gutteridge, W. E., M. Gaborak and B. Cover, *Ann. Trop. Med. Parasitol.* 72, 339 (1978)
- 43 Haberkorn, A., *Zbl. Bakteriell. Parasitenkunde. Infektionskrankh. Hyg. Abt. I. Orig.*, R. A 244, 331 (1979)
- 44 Haraoka, V., A. Sugihara and M. Ito, *Jap. P.* ('66), 1788 (1966)
- 45 Harris, G. H., and B. C. Fischback, *USA P.* 2 951 868 (to Dow Chemical Co), Sept. 6, 1960
- 46 Haugwitz, R. D., B. V. Maurer and V. L. Narayanan, *J. med. Chem.* 15, 856 (1972)
- 47 Hellerbach, J., and A. Szenté, *Swiss P.* 597 226 (to Hoffmann-La Roche AG), 1978
- 48 Henry, D. W., V. H. Brown, M. Cory, J. G. Johansson and E. Bueding, *J. med. Chem.* 16, 1287 (1973)
- 49 Hoyle, W., and M. P. Savage, *Swiss P.* 547 820 (to Ciba-Geigy AG), 1974
- 50 Hulbert, P. B., E. Bueding and C. H. Robinson, *J. med. Chem.* 16, 72 (1973)
- 51 Ingalls, J. W., *Kirk-Othmer Encycl. Chem. Technol.* 3rd ed. vol. 5, p. 451, John Wiley & Sons, New York 1979
- 52 Islip, P. J., M. D. Cosier and J. E. Wcale, *J. med. Chem.* 16, 1027 (1973)
- 53 Islip, P. J., M. D. Cosier and M. C. Neville, *ibid.* 16, 1030 (1973)
- 54 Jennings, F. W., *UK P.* (application) 2 024 008 (to Merck), Sept. 9, 1980
- 55 Jennings, F. W., G. M. Urquhart, P. K. Murray and B. M. Miller, *Inst. J. Parasitol.* 16, 27 (1980)
- 56 Kinnamon, K. E., E. A. Steck, W. L. Hanson and W. L. Chapman, Jr., *J. med. Chem.* 20, 741 (1977)
- 57 Kuppuswamy, N., A. P. Vishwa and T. George, *Stouth African P.* 7 406 157 (to Ciba-Geigy AG), 1978
- 58 Koji, M., I. Masao and Y. Sachiho, *J. pharmac. Soc. Japan [Yakugaku Zasshi]* 75, 57 (1955)
- 59 Lee, R. M., M. W. Mills and G. S. Sach, *Experientia [Basel]* 33, 198 (1977)
- 60 Lin, Y., P. B. Hulbert, E. Bueding and C. H. Robinson, *J. med. Chem.* 17, 835 (1974)
- 61 Mason, R. P., and J. L. Holtzman, *Biochem. biophysic. Res. Commun.* 67, 1267 (1975)
- 62 Mason, R. P., and J. L. Holtzman, *Biochemistry* 14, 1626 (1975)
- 63 Mason, R. P., and J. L. Holtzman, *Federat. Proc.* 34, 665 (1975)
- 64 McGuire, W. C., R. C. O'Neill and G. Brody, *J. Parasitol.* 52, 528 (1966)
- 65 Meiser, W., and M. Federmann, *USA P.* 3 082 151 (to Farbenfabriken Bayer AG), March 19, 1963
- 66 Michels, J. G., *USA P.* 2 920 074 (to Norwich Pharmacol. Co), January 5, 1960
- 67 Michels, J. G., *USA P.* 3 141 880 (to Norwich Pharmacol. Co), July 21, 1964
- 68 Middleton, K. R., F. W. Schaefer III and H. J. Szaz, *Experientia [Basel]* 35, 243 (1979)
- 69 Miller, B. M., C. M. Malangu, J. Taylor and J. Conroy, *Abstr. Int. Conf. Chagas' Disease* 140, Rio de Janeiro 1979
- 70 Mueller, M., D. G. Lindmark and J. McLaughlin, in: H. van den Bossche (Ed.), *Biochem. Parasites Host-Parasites Relat.*, Proc. 2nd Int. Sympos. p. 537, North Holland, Amsterdam 1976
- 71 Nabin, I., M. M. Kamel and M. Saddik, *Pharmazie* 35, 234 (1980)
- 72 Nafissi-Varchei, M. M., *USA P.* 4 021 570 (to Schering Corp), May 3, 1977
- 73 Nagarajan, K., V. P. Arya and T. George, *Austral. P.* 492 051 (to Ciba-Geigy AG), 1978
- 74 Nagarajan, K., V. P. Arya and T. George, *Canad. P.* 1 039 732 (to Ciba-Geigy AG), 1979
- 75 Nakao, H., and K. Kitano, *Jap. Kokai* 77 139 032, November 19, 1977
- 76 Neville, M. C., and J. P. Verge, *J. med. Chem.* 20, 946 (1977)
- 77 Ono, T., and S. Inoki, *Kiseichugaku Zasshi* 22, 320 (1976)
- 78 Pelosi, S. S., Jr., *USA P.* 4 066 660 (1978)
- 79 Peterson, F. J., G. F. Comiss, Jr. and J. L. Holtzman, *Federat. Amer. Soc. exp. Biol.* 6rd Ann. Meeting, Abstr. 1045 (1979)
- 80 Polak, A., and R. Riche, *Ann. Trop. Med. Parasitol.* 72, 45 (1978)
- 81 Raether, W., and N. Deutschländer, *Abstr. Int. Conf. Chagas' Disease*, 142, Rio de Janeiro 1979
- 82 René, L., S. Risse, P. Demerseman, R. Royer and R. Cavier, *Europ. J. med. Chem. chim. Therap.* 14, 281 (1979)
- 83 Report on the Third Meeting of the Scientific Working Group on Filariasis: Filariicide Screening, WHO, February 1979
- 84 Robinson, C. H., E. Bueding and J. Fisher, *Molecular Pharmacol.* 6, 604 (1970)
- 85 Robinson, C. H., S. Spencer and E. Bueding, *J. med. Chem.* 16, 79 (1973)
- 86 Sallazar-Mallen, M., *Ann. Trop. Med. Parasitol.* 65, 393 (1971)
- 87 Salomon, N., *Israel. P.* 11 059, July 23, 1958
- 88 Sanderson, B. E., in: A. E. R. Taylor and R. Muller (Eds.), *Symposia of the British Society for Parasitology*, vol. 11, p. 53, Blackwell Scientific Publications, Oxford 1973
- 89 Sas, H. J., G. A. Dunbar and E. Bueding, *Amer. J. trop. Med. Hyg.* 26, 475 (1977)
- 90 Schier, O., and A. Marxer, in: E. Jucker (Ed.), *Progress in Drug Research*, vol. 22, p. 28, Birkhäuser Verlag, Basel 1978
- 91 Schlemper, B. R., Jr., E. Chiari and Z. Brenner, *J. Protozool.* 24, 544 (1977)
- 92 Sen, H. G., *Acta tropica* 33, 101 (1976)
- 93 Shumard, R. S., *Soap Sanit. Chemicals* 29, 34 (1953)
- 94 Sims, P., and W. E. Gutteridge, in: H. van den Bossche (Ed.), *Biochem. Parasites Host-Parasite Relat.*, Proc. 2nd Int. Sympos., p. 485, North Holland, Amsterdam 1977
- 95 Sims, P., and W. E. Gutteridge, *IRCS Med. Sci.: Libr. Compend.* 6, 299 (1978)
- 96 Singh, H., S. Sharma, A. K. Singh, R. N. Lyer, P. Govil, J. C. Katiyar and A. B. Sen, *Indian J. exp. Biol.* 14, 332 (1976)
- 97 Singh, H., A. K. Singh, S. Sharma, R. N. Lyer and O. P. Srivastava, *J. med. Chem.* 20, 826 (1977)
- 98 Stohler, R., *Tropenmed. Parasitol.* 28, 276 (1977)
- 99 Striebel, H. P., p. 17 in [1]
- 100 Striebel, H. P., *Experientia [Basel]* 32, 457 (1976)
- 101 Struff, R., and R. Goennert, *Z. Tropenmed. Parasitol.* 18, 193 (1967)
- 102 Tchelitcheff, S., *Ger. Offen* 2 214 258 (to Rhone-Poulenc SA), Oct. 5, 1972
- 103 Thomas, H., p. 49 in [1]
- 104 Thorson, R. E., L. Goldman, W. S. Wallace and A. E. Lansilotti, *USA P.* 3 081 224 (to American Cyanamid Co), March 12, 1963
- 105 Ueda