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# Management of gastrointestinal stromal tumours: from diagnosis to treatment

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## Summary

Gastrointestinal stromal tumours (GIST) are the most common mesenchymal tumours of the digestive tract. Most gastrointestinal soft tissue neoplasms, previously classified as leiomyomas, schwannomas, leiomyoblastomas or leiomyosarcomas, are today classified as GIST on the basis of molecular and immunohistological features. They originate from gastrointestinal pacemaker cells and are characterised by over-expression of the tyrosine kinase receptor KIT. Overall 5-year survival after surgical resection of GIST is approximately 60%. However, these tumours span a wide clinical spectrum from benign to highly malignant. Prognostic factors have recently been identified for GIST and include tumour size, mitotic rate and other minor factors. At present, surgery is the standard treatment for primary resectable GIST. Benign GIST have an excellent prognosis after

primary surgical treatment, with over 90% 5-year survival. While recurrent or malignant GIST, which are resistant to radiotherapy and chemotherapy, had until recently an extremely poor prognosis even after surgical resection, with median survival of 12 months. The development of a tyrosine kinase inhibitor has changed the management of unresectable malignant cases. This new tyrosine kinase inhibitor, imatinib mesylate, which inhibits the c-kit receptor, has proved highly effective against GIST and has improved survival in metastatic GIST. This paper reviews the literature and our experience of GIST, including: diagnosis, pathology, treatment and prognosis.

*Key words:* gastrointestinal stromal tumour (GIST); mesenchymal tumour; c-kit (CD117); tyrosine kinase inhibitor; imatinib mesylate (STI-571)

## Introduction

Up to 20 years ago most mesenchymal tumours of the digestive tract were considered to be of smooth muscle or perineural origin. In 1983, Mazur and Clark [1] coined the term *gastrointestinal stromal tumours (GIST)* to indicate a distinctive subgroup of gastrointestinal mesenchymal tumours which could be classified neither as neurogenic- nor smooth muscle-derived. Kindblom et al. [2] proposed that these tumours may originate from the interstitial cell of Cajal, an intestinal pacemaker cell, and suggested the name gastrointestinal pacemaker cell tumour. This hypothesis is supported by the observation that GIST cells show ultrastructural features and express cell markers typical of the normal interstitial cell of Cajal [2]. Histological features of GIST may explain the past diagnostic confusion with other mesenchymal tumours. Today, on the basis of pathological features, most gastrointestinal mesenchymal tumours previously designated as smooth muscle tumours,

such as leiomyomas, leiomyoblastomas and leiomyosarcomas, are GIST [3–5]. However, true gastrointestinal leiomyomas and schwannomas remain to be identified.

Although rare, GIST are the most common mesenchymal tumours of the gastrointestinal tract. GIST are of clinical relevance because in at least 10–30% of cases they are malignant [4, 6]. In the past, GIST chemo- and radio-resistance made treatment a challenging issue for physicians, while criteria for evaluation of the malignant potential of GIST were lacking. Currently, identification of reliable prognostic factors and the development of molecular-targeted anticancer strategies for GIST has given new hope to these patients [7–9].

In this review we report on the clinical aspects of GIST, from diagnosis to prognosis, and review the published literature as well as our experience. We also review the therapeutic options for GIST.

## Clinical presentation

GIST, the most common mesenchymal tumours of the digestive tract, account for 0.1–3% of all gastrointestinal cancers. However, they represent up to 20% of small bowel malignancies [10]. The incidence of primary GIST is around 20/10<sup>6</sup> people/year [11]. GIST predominantly occurs in middle-aged and older persons (5<sup>th</sup> to 7<sup>th</sup> decade), with no significant difference in distribution between males and females [4, 12].

Most GIST arise in the stomach (approximately 60%) and small intestine (approx 30%), and infrequently from the duodenum, the colon and rectum or the mesentery. Oesophageal localisation is extremely rare [4, 13–15].

The symptoms, which depend on tumour size and location, are usually nonspecific [16]. Small GIST are usually asymptomatic and are detected either during investigations or surgical procedures

for unrelated disease. Incidental discovery accounts for approximately one third of the cases. The commonest presentation of GIST is bleeding related to mucosal erosion (approximately 50%) [16], which may be either chronic, with anaemia, or acute, necessitating emergency treatment (approximately 40% of the cases presenting with haemorrhage) [17]. Some 20% of patients present abdominal discomfort, or even pain, which is generally associated with larger size GIST. In the small intestine bowel obstruction is also frequent. Bowel perforation is infrequent.

At first diagnosis some 10% of patients present with metastatic disease [13]. GIST usually, and primarily, metastasise to the liver. Peritoneal surface diffusion is less frequent, while lymph nodes and extra-abdominal sites are rarely involved.

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## Diagnosis

There are no recognised specific radiological examinations for GIST diagnosis. Barium contrast studies or endoscopy may provide useful data on localisation of GIST. As for the other intra-abdominal malignancies, computed tomography scan (CT scan) is the standard preoperative imaging technique [18]. CT scan may reliably localise GIST, as well as determining size and possibly revealing the presence of secondary localisations (i.e. hepatic metastases) [18]. GIST imaging by CT scan usually discloses an extraluminal mass originating from the digestive tract wall, with frequent central necrosis. Percutaneous needle biopsy carries the theoretical risk of peritoneal seeding and is indicated only for clearly unresectable disease or

where treatment would be altered by pathological diagnosis. Demonstration of KIT positive cells may assist in diagnosing GIST on needle biopsy, although the positive predictive value of this approach needs to be defined [19]. Positron emission tomography with 18FDG is particularly useful for detection of secondary localisation of GIST, but hitherto has been chiefly used for patient follow-up [20, 21].

The diagnosis of GIST may be suggested during surgery by the presence of a well defined extraluminal mass, frequently polylobulate with a pseudocapsule. However, it always requires histological and immunohistochemical confirmation.

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## Pathology

On the basis of molecular and immunohistologic features, most gastrointestinal soft tissue sarcomas previously designated as smooth muscle tumours, such as leiomyomas, schwannomas, leiomyoblastomas and leiomyosarcomas, are classified as GIST [5]. The histological and cytological features of GIST may explain the past diagnostic confusion with other mesenchymal tumours. However, true leiomyoma (especially in the oesophagus), leiomyosarcoma or neural cell tumours should still be classified as such and not as GIST [4]. In particular, the former is the most common mesenchymal tumour of the oesophagus and is characterised by a benign course [4].

It has become clear that the tumour cells comprising GIST are closely related to the interstitial cells of Cajal [22, 23]. These cells constitute a complex cellular network, the likely functions of which

are gastrointestinal tract pacemaking and the regulation of intestinal motility [2, 5, 24]. The immunohistochemistry of the interstitial cells of Cajal is similar to that of GIST cells, being positive for KIT [2, 5, 24]. However, some GIST arise from the mesentery or omentum, which lacks interstitial cells of Cajal, suggesting an origin in multipotential mesenchymal stem cells of Cajal cell lineage [11, 25, 26].

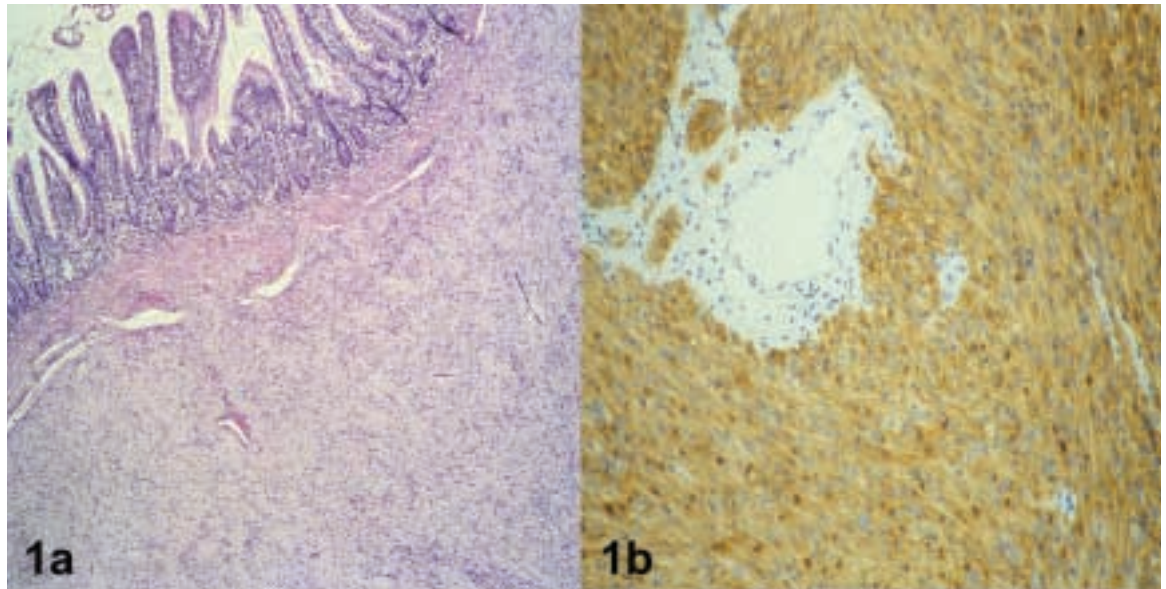
Grossly, GIST vary greatly in size, ranging from 1 cm to more than 20 cm. They not infrequently exhibit areas of necrosis, cystic degeneration or focal haemorrhage. The tumours are well circumscribed and pseudo-encapsulated. They arise in the digestive tract wall, generally with extraluminal development, and not infrequently cause ulceration of the overlying mucosa. They are composed of either spindle-shaped cells (70%) or

**Figure 1**

Histopathology of GIST.

a: Small bowel GIST. Note the dense cellular tumour located in the small bowel wall with regular nucleus (haematoxylin-eosin, 50× magnification).

b: Small bowel GIST. Immunostaining for KIT (CD117). Note the highly and diffusely positive tumoral cells for KIT (brown coloration) (CD117 staining, DAKO, 200× magnification).



epithelioid cells, while mixed cell populations have rarely been encountered (Figure 1) [4].

In contrast to leiomyomas and leiomyosarcomas, GIST are typically immunoreactive for KIT (CD117). They are immunohistochemically positive for KIT, with a reported frequency of 90 to 100% depending on the method used [4], while positivity for CD34, the haematopoietic progenitor cell antigen, is reported in 70–80% of GIST [4]. GIST are frequently vimentin positive. They

may be focally positive for smooth muscle actin (30%) and keratin (<10%) but are negative usually for desmin and protein S100. It should be noted that some other tumour types (e.g. melanoma, mastocytoma) may also be CD117 positive, but the differential diagnosis may depend on histological and immunohistochemical differences. In summary, histological (i.e. cellular type and organisation) and immunohistochemical (i.e. KIT, CD34) findings are the defining features of GIST.

## Molecular biology of GIST

KIT, which is a growth factor transmembrane receptor, is the product of the proto-oncogene *c-kit* (chromosome 4). As a member of the tyrosine kinase receptor, KIT is closely related to the receptors for platelet-derived growth factor (PDGF) and other receptors of this family [27, 28]. KIT is expressed by haematopoietic progenitor cells, mast cells, germ cells and interstitial cells of Cajal. Activation of the KIT receptor by its ligand, known as stem-cell factor, leads to cascades involved in oncogenesis, including proliferation, adhesion, apoptosis and differentiation [27, 28]. While activation of wild-type *c-kit* is required for interstitial Cajal cell development [29], gain of function mutations of *c-kit* occurs in up to 90% of GIST [3, 5, 11, 15], the most frequent being mutations of exon 11 (70% of GIST) and 9 [3, 27, 28, 30, 31]. These mutations lead to constitutional activation of the receptor, which in normal circumstances takes place upon binding of the ligand. The activating mutation perpetuates the KIT signal and the downstream transduction pathway, ultimately leading to activation of cellular proliferation [30, 32, 33]. Inherited germline mutations of exon 11 or 13 produce an autosomal dominant susceptibility to the formation of GIST [34, 35].

Affected individuals develop diffuse hyperplasia of the interstitial cells of Cajal and are subject to the occurrence of multiple GIST during adulthood [34]. Thus mutations in KIT seem to play a gate-keeper role in transformation of the interstitial cells of Cajal into a GIST [27, 29, 35]. However, apparently KIT mutation alone does not cause malignant transformation of GIST. This is consistent with data on the prognosis of patients with sporadic or familial GIST. The proportion of GIST with KIT mutation attains 90%, and comparison of the prognosis of GIST with or without KIT mutation has thus far shown no strong correlation [35–37]. Factors causing the transformation of benign to malignant GIST have still to be identified, but degeneration from benign to malignant GIST is apparently associated with chromosomal depletion and/or acquisition of mutations in various oncogenes [27].

However, some GIST, which are not KIT positive (approximately 5% of GIST), do not present mutation of the *c-kit* proto-oncogene [28], although some of them harbour mutation of the PDGFRA receptor, another tyrosine kinase receptor, and this could be related to oncogenesis in these cases [28, 35, 38].



Prognostic factors

GIST display different degrees of aggressiveness. Indeed, study of site-specific and combined series of GIST have shown that these tumours have a spectrum spanning from small, benign nodules to overtly malignant tumours at all sites of occurrence [13]. Predicting the potential biological behaviour of these tumours remains difficult, and the literature contains many conflicting reports on this issue [9, 39–41]. Since A. P. Stout et al. [42] pointed out that the number of mitoses is a prognostic factor for stromal tumours, many criteria have been correlated with prognosis or survival, including: localisation [43], tumour size (<5 cm in diameter) [14, 43–47], mitotic activity (>5 mitosis/high-power fields) [3, 43, 44, 46, 48], tumour necrosis [44, 48–50], staining for proliferating cells (MiB1 >10%) [3, 51–53], invasive character [41, 48, 53, 54], presence of symptoms [48], histological type [55], immunohistochemical profile [53], presence of metastases or lymph node invasion [14, 40], and others. The role of GIST localisation is debated, some authors having found that progn-

sis is significantly influenced by localisation and others not. However, except the presence of metastases, none of these factors taken independently is predictive for the malignant potential of GIST [6, 36, 39, 55, 56]. It is of importance that the presence of multiple synchronous GIST does not appear to affect outcome [57, 58].

The most easily applicable and recognised pathological criteria for prediction of recurrence risk are tumour size and mitotic rate, and over the last decade prognostic scales combining these two criteria have been evaluated [39, 45, 55, 56]. Recently, via a consensus approach supported by the National Institute of Health, these criteria have been combined in a new scale to define the risk of aggressive-behaviour GIST (Table 1) [6]. This scale is easy to apply for pathological classification. According to this scale the the prognosis of very low risk GIST is excellent, while high risk tumours are associated with a high rate of recurrence and decreased survival. However, the prognosis of low and intermediate risk GIST is in-between and not well defined. Thus the reliability and clinical usefulness of this prognostic scale is questionable, since it is not suited to specifically identifying high-risk patients who may benefit from more aggressive treatments. We have recently proposed a new scaling system for GIST based on their malignant potential [58]. In accordance with the recent studies of Yan et al. [54], it is not only based on tumour size and mitotic rate but refers to other criteria with the aim of improving the classification of intermediate risk cases. The scale is composed of five minor criteria (i.e. tumour size >5 cm, mitotic rate >5/50 HPF, Mib1 >10%, presence of necrosis and invasive components to mucosa or serosa), and two major criteria (presence of either lymph node invasion or metastases) (Table 2). GIST harbouring either four of the five minor criteria, or one major criterion, are considered to have high malignant potential, while others are considered of low malignant potential (benign). This scaling system significantly correlates with overall and event-free survival and makes it possible to detect all GIST at risk of recurrence, which would benefit from more aggressive treatment. With this scale, the 5-year survival of low malignant potential GIST is 95% and of high malignant potential GIST less than 20%. However, the scales of both Fletcher et al. and ourselves need to be validated in large prospective GIST cohorts in terms of positive and negative predictive value for risk of recurrence and reliability, on the basis of patient survival.

Table 1

Proposed approach for assessing risk of aggressive behavior of GIST (adapted from Fletcher et al. [6]).

	Tumour size*	Mitotic count**
Very low risk	<2 cm	<5/50 HPF
Low risk	2–5 cm	<5/50 HPF
Intermediate risk	<5 cm	6–10/50 HPF
	5–10 cm	<5/50 HPF
High risk	>5 cm	>5/50 HPF
	>10 cm	any mitotic rate
	any size	>10/50 HPF

\* Size represents the single largest dimension.  
\*\* Ideally, mitotic count should be standardised according to surface area examined (based on size of high-power fields, HPF). Despite inevitable subjectivity in recognition of mitoses and variability in the area of HPF, mitotic counts still prove useful.

Table 2

Proposed classification of GIST according to malignant potential (adapted from Bucher et al. [58]).

Minor criteria
Tumour size ≥5 cm
Mitotic index ≥5 mitoses/50 hpf
Presence of necrosis
Infiltration of adjacent structures (i.e. mucosa or serosa)
MiB1 index ≥10%
Major criteria
Presence of lymph node invasion
Presence of GIST metastases
Low malignant potential GIST:
GIST with fewer than 4 minor criteria
High malignant potential GIST:
GIST with 4 or 5 minor criteria or 1 major criterion

## Surgical treatment

The treatment of choice for GIST is surgical resection. All GIST should be approached with the intention of performing complete en bloc removal (R0 resection) of the tumour and surrounding tissue [14, 47, 59–62]. It has been shown that completeness of resection correlates with survival after primary resection of GIST [14, 59, 63–65]. This is true of localised disease, but for metastatic disease the role of synchronous hepatic metastasis resection has not been defined in the imatinib mesylate era (Figure 2). However, synchronous liver metastasis resection is advocated when applicable, since a complete and long term response to imatinib mesylate has not thus far been demonstrated. In some cases with primarily non-resectable metastases complementary resection should be done, if possible after response to imatinib mesylate treatment (Figure 2). The rate of resectable GIST, reported in the literature ranges from 50 to 90% [12, 14, 44, 60, 61]. These discrepancies are partly attributable to the type of patient cohort studied. Series concerning GIST with primary presentation in non-specialised centres report a resectability rate higher than those evaluating GIST referred to specialised centres, where the cases collected are chiefly advanced tumours. In our experience GIST resectability is some 90%. However, resection of the primary lesion, even incomplete, is indicated

for the palliation of symptoms associated with mass effect and especially the risk of bleeding [14, 66].

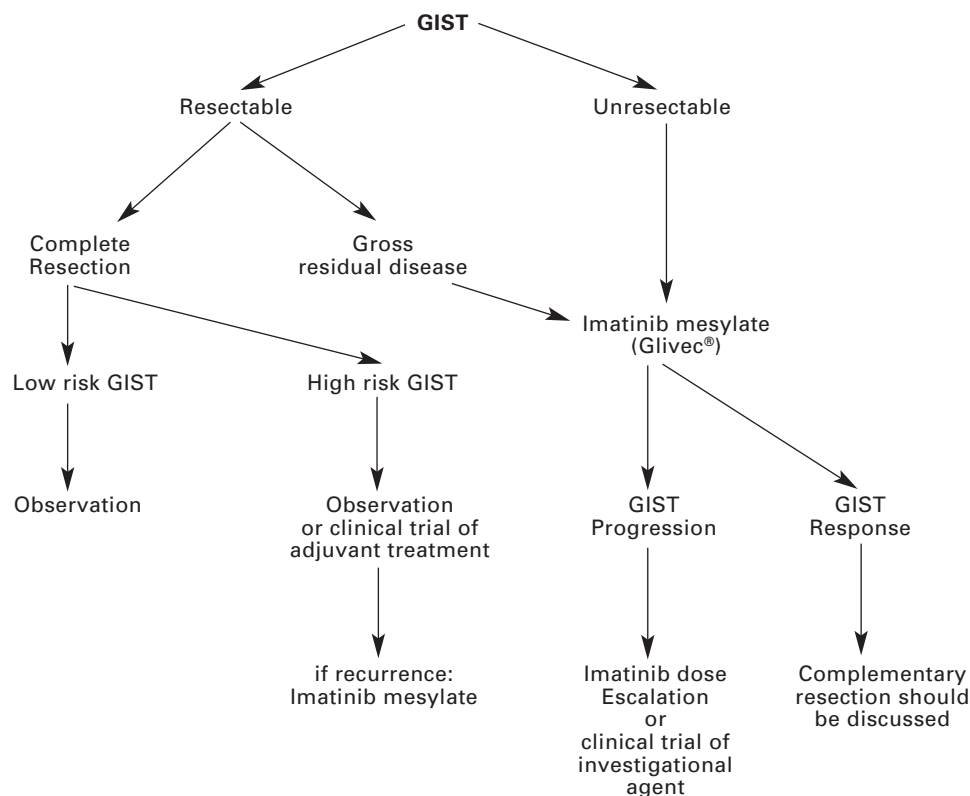
Several reviews have reported that small GIST (2 cm or less) can be treated adequately by wedge (gastric) or segmental (bowel) resection [61]. Larger GIST may require more extensive en bloc resection including adjacent structures or organs if involved [61].

GIST, even with high malignant potential, metastasise to lymph nodes with insufficient frequency to warrant lymph node dissection [3, 14, 58, 59, 61, 67]. Similarly to other sarcomas, lymph node invasion is generally encountered after development of blood-borne metastases and appears to be a late event in the natural history of malignant GIST [68, 69].

Surgical resection in patients with a primary GIST is associated with a 5-year survival rate of 48–70% [12, 14, 43, 44, 48, 59, 61, 69]. However, prognosis of low risk (low malignant potential) GIST after complete resection is excellent [40, 58], while high risk (high malignant potential) GIST have a high rate of recurrence (Figure 2) [6, 14, 40, 58]. There is no indication for chemotherapy and radiation therapy after surgical resection of GIST, as these tumours are notoriously unresponsive to such treatment [12, 14, 37, 67].

**Figure 2**

Treatment algorithm for patient with primary presentation of GIST.



## Molecularly targeted treatment

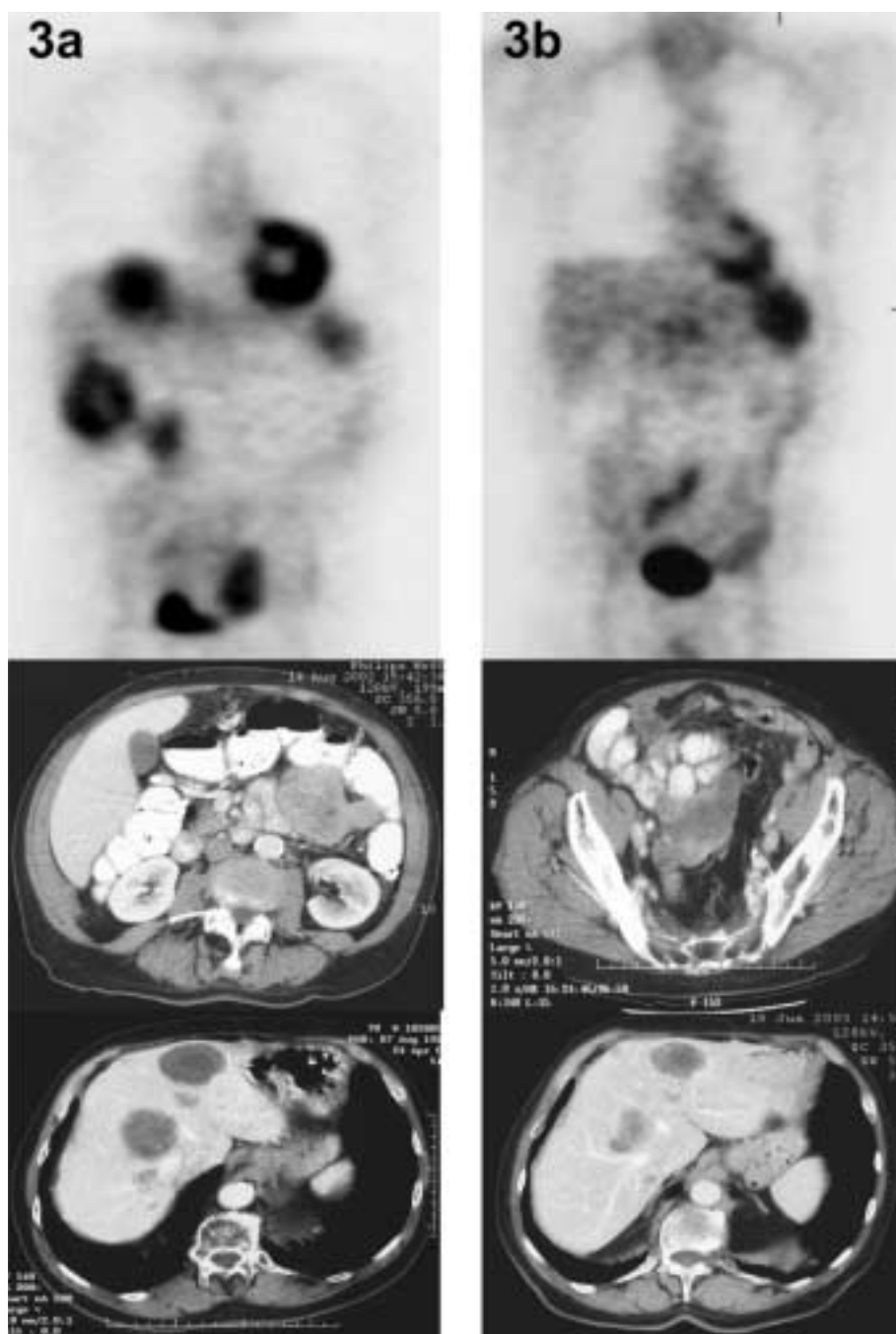
Several protein kinases are deregulated or overexpressed in human cancers due to gene mutations, and are therefore targets for selective pharmacological inhibitors. Imatinib mesylate (formerly referred to as STI571, and now manufactured as Glivec in Europe) is a powerful and relatively selective inhibitor of all ABL tyrosine kinases, including: c-kit, c-ABL, bcr-ABL and platelet-derived growth factor receptor (PDGFRA) [70, 71]. The efficacy of imatinib mesylate as a tyrosine kinase inhibitor and its pharmacokinetics in humans have been assessed in studies of patients with chronic myeloid leukaemia (CML) [72]. In CML, a chromosomal rearrangement in haematopoietic stem cells causes the formation of a fusion protein, BCR-ABL, in which the tyrosine kinase component is constitutively ac-

tive, a ligand-independent activation similar to the role of KIT in GIST cells [73]. In 1996 Druker et al. [74] reported that imatinib specifically inhibited or killed proliferating myeloid cells containing BCR-ABL. Imatinib mesylate at therapeutic drug levels inhibits the receptors KIT and PDGFRA by reversible binding [71, 75]. It has been shown that the vast majority of KIT mutants as well as wild-type KIT are sensitive to imatinib mesylate. Moreover, activation of ligand-stimulated native PDGFRA and PDGFRA mutant are potently inhibited by imatinib mesylate [71, 75, 76].

The response rate of recurrent or metastatic GIST to imatinib mesylate treatment is some 60–70% [7, 9, 15, 37, 77]. Long term results of patient survival on imatinib mesylate treatment are not available, but while patient median survival was

**Figure 3**

Imaging of metastatic GIST before and after treatment with imatinib mesylate.  
a: Before surgical and imatinib mesylate treatment. Top: PET scan with tumoral activity related to hepatic metastases and small bowel GIST and peritoneal second localisation; centre: small bowel GIST; bottom: hepatic metastases of GIST.  
b: After resection of small bowel GIST and 3 months' treatment with imatinib mesylate. Top: PET scan without residual tumoral activity; centre: residual "necrotic" peritoneal GIST mass; bottom: residual hepatic GIST mass.



less than 12 months before the imatinib mesylate era, large series have shown that after 2 years the median of survival was not attained in patients under treatment [7, 15]. Some cases experience stable disease (15%) under treatment, while some patients do not respond at all to treatment and show disease progression after introduction of imatinib mesylate [7, 15]. For these cases an increase in drug dosage may be indicated and, if this is not sufficient, an investigational protocol of complementary treatment could be tried (Figure 2).

It has been shown that the type of KIT mutation influences response to treatment with imatinib mesylate. Patients whose tumours express exon 11 mutant KIT protein have a significantly better response rate to imatinib therapy (83.5%) than patients whose tumours express either exon 9 mutant KIT (47.8%) or contain no detectable mutation of KIT or PDGFRA (0%) [28]. This was, however, associated with longer overall survival. Not all GIST without mutation of the KIT receptor are resistant to imatinib mesylate. Sensitivity and response to therapy in these cases may be explained by the presence of PDGFRA mutations [28, 78].

The duration of treatment with imatinib mesylate is a matter of debate, no clear data having emerged regarding the need to continue therapy indefinitely [8]. However, KIT-positive Ki67-negative cells (non-proliferating cells), probably representing non-cycling or dormant GIST cells,

may be encountered several months after starting imatinib mesylate even in a responding patient, and development of secondary resistance to treatment may supervene [79]. The current recommendation is to continue the treatment indefinitely while trials are under way to assess the evolution of patients after imatinib mesylate withdrawal [15].

For patient follow-up during imatinib mesylate treatment, positron emission tomography (PET scan) with 18FDG is the method of choice [20]. PET scan is a highly sensitive tool for monitoring of response to treatment, while CT scan is unable to detect response for 3–6 months after treatment initiation (Figure 3). Moreover, it has been shown that GIST response assessed by PET scan eight days after the start of imatinib mesylate treatment correlates with prognosis and survival at one year in patients with non-resectable GIST [20, 21].

With regard to the pharmacokinetics of imatinib mesylate, it is well absorbed after oral administration with a half-life in the circulation of approx. 16 hours, a period compatible with administration once daily. While it has been associated with only a few minor side effects [15, 80, 81], gastrointestinal or intra-abdominal haemorrhage have been reported in patients with large GIST after initiation of imatinib mesylate treatment; this may be related to the rapid anti-cancer effect on the tumours, with disruption of tumour-related vasculature [15].

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## Prognosis

In the literature patient survival after primary surgical resection of GIST ranges from 48% to 80% at 5 years [12, 14, 15, 59]. However, these data concern the era before imatinib mesylate and GIST prognosis has drastically changed since its introduction. Moreover, overall GIST prognosis is of little value, as low risk and high risk GIST do not have the same natural history after surgical resection. For low malignant potential (low risk) GIST, the 5-year survival rate (approximately 95%) is similar to the normal population [58], while for high malignant potential (high risk) GIST the 5-year survival rate ranged from 0% to 30% before the introduction of imatinib mesylate [58]. No long-term survival data are available for malignant GIST in the imatinib mesylate era, but current experience show major improvement at

1 year, to 90% versus <50% before the imatinib mesylate era [15, 20, 21]. Recurrences are extremely rare for low malignant potential GIST, while in our experience, which is in agreement with the literature, more than 80% of high malignant potential GIST will recur [40]. Patient follow-up after curative surgical resection of GIST should be tailored to the malignant potential of their tumour [40, 58, 63, 65]. While follow-up on a yearly basis appears to suffice for low malignant potential GIST, closed control is required for high malignant potential GIST as up to 50% of recurrences occur during the first year post resection [65]. For patient follow-up PET scan is currently the most reliable investigation; abdominal CT scan is valuable for detection of recurrence [14, 20, 21, 58].

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## Discussion

In this paper we have reviewed current knowledge of gastrointestinal stromal tumour (GIST) management.

Surgical resection is the treatment of choice for these tumours. GIST studies have shown that after surgical resection these tumours span a wide



clinical spectrum from benign to malignant tumours. Newly developed prognostic scales make it possible to distinguish low malignant (benign or low risk) from high malignant (malignant or high risk) potential GIST. While low malignant potential GIST have an excellent prognosis after resection, high malignant potential GIST involve a high rate of recurrence with poor survival after surgical treatment alone. Imatinib mesylate is a powerful agent against metastatic or recurrent GIST. Imatinib mesylate is one of the first examples of a drug that targets an intracellular signaling molecule in clinical cancer therapy for stromal tumours [82]. However, experience of follow-up with imatinib mesylate therapy is short, and some questions remain [83]. In the light of the results in CML there

is reason to believe that imatinib may be even more effective when given earlier in the management of GIST, for example as adjuvant therapy for high malignant potential GIST. This approach is under investigation [77, 84].

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## References

- Mazur MT, Clark HB. Gastric stromal tumours. Reappraisal of histogenesis. *Am J Surg Pathol* 1983;7:507–19.
- Kindblom LG, Remotti HE, Aldenborg F, Meis-Kindblom JM. Gastrointestinal pacemaker cell tumour (GIPACT): gastrointestinal stromal tumours show phenotypic characteristics of the interstitial cells of Cajal. *Am J Pathol* 1998;152:1259–69.
- Miettinen M, Sarlomo-Rikala M, Lasota J. Gastrointestinal stromal tumours: recent advances in understanding of their biology. *Hum Pathol* 1999;30:1213–20.
- Miettinen M, Majidi M, Lasota J. Pathology and diagnostic criteria of gastrointestinal stromal tumours (GISTs): a review. *Eur J Cancer* 2002;38:S39–S51.
- Graadt van Roggen JF, van Velthuysen ML, Hogendoorn PC. The histopathological differential diagnosis of gastrointestinal stromal tumours. *J Clin Pathol* 2001;54:96–102.
- Fletcher CD, Berman JJ, Corless C, Gorstein F, Lasota J, Longley BJ, et al. Diagnosis of gastrointestinal stromal tumours: A consensus approach. *Hum Pathol* 2002;33:459–65.
- Demetri GD, von Mehren M, Blanke CD, Van den Abbeele AD, Eisenberg B, Roberts PJ, et al. Efficacy and safety of imatinib mesylate in advanced gastrointestinal stromal tumours. *N Engl J Med* 2002;347:472–80.
- Demetri G. Targeting the molecular pathophysiology of gastrointestinal stromal tumours with imatinib. Mechanisms, successes and challenges to rational drug development. *Hematol Oncol Clin North Am* 2002;16:1115–24.
- Joensuu H, Fletcher C, Dimitrijevic S, Silberman S, Roberts P, Demetri G. Management of malignant gastrointestinal stromal tumours. *Lancet Oncol* 2002;3:655–64.
- Blanchard DK, Budde JM, Hatch GF, 3rd, Wertheimer-Hatch L, Hatch KF, Davis GB, et al. Tumours of the small intestine. *World J Surg* 2000;24:421–9.
- Miettinen M, Lasota J. Gastrointestinal stromal tumours – definition, clinical, histological, immunohistochemical, and molecular genetic features and differential diagnosis. *Virchows Arch* 2001;438:1–12.
- Rossi CR, Mocellin S, Mencarelli R, Foletto M, Pilati P, Nitti D, et al. Gastrointestinal stromal tumours: From a surgical to a molecular approach. *Int J Cancer* 2003;107:171–6.
- Miettinen M, Sarlomo-Rikala M, Lasota J. Gastrointestinal stromal tumours. *Ann Chir Gynaecol* 1998;87:278–81.
- Dematteo RP, Lewis JJ, Leung D, Mudan SS, Woodruff JM, Brennan MF. Two hundred gastrointestinal stromal tumours: recurrence patterns and prognostic factors for survival. *Ann Surg* 2000;231:51–8.
- Demetri G. Identification and treatment of chemoresistant inoperable or metastatic GIST: experience with the selective tyrosine kinase inhibitor imatinib mesylate (STI571). *Eur J Cancer* 2002;38:S52–S59.
- Ludwig DJ, Traverso LW. Gut stromal tumours and their clinical behavior. *Am J Surg* 1997;173:390–4.
- Catena F, Pasqualini E, Campione O. Gastrointestinal Stromal Tumours: Experience of an Emergency Surgery Department. *Dig Surg* 2000;17:503–7.
- Buckley JA, Fishman EK. CT evaluation of small bowel neoplasms: spectrum of disease. *Radiographics* 1998;18:379–92.
- Li SQ, O'Leary TJ, Buchner SB, Przygodzki RM, Sobin LH, Erozan YS, et al. Fine needle aspiration of gastrointestinal stromal tumours. *Acta Cytol* 2001;45:9–17.
- Van den Abbeele A, Badawi R. Use of positron emission tomography in oncology and its potential role to assess response to imatinib mesylate therapy in gastrointestinal stromal tumours (GISTs). *Eur J Cancer* 2002;38:S60–S65.
- Stroobants S, Goeminne J, Seegers M, Dimitrijevic S, Dupont P, Nuyts J, et al. 18FDG-Positron emission tomography for the early prediction of response in advanced soft tissue sarcoma treated with imatinib mesylate (Glivec). *Eur J Cancer* 2003;39:2012–20.
- Sakurai S, Fukasawa T, Chong JM, Tanaka A, Fukayama M. Embryonic form of smooth muscle myosin heavy chain (SMemb/MHC-B) in gastrointestinal stromal tumour and interstitial cells of Cajal. *Am J Pathol* 1999;154:23–8.
- Nishida T, Hirota S. Biological and clinical review of stromal tumours in the gastrointestinal tract. *Histol Histopathol* 2000;15:1293–301.
- Chan J. Mesenchymal tumours of the gastrointestinal tract: a paradise for acronyms (STUMP, GIST, GANT, and now GIPACT). Implication of c-kit in genesis, and yet another of the emerging roles of the interstitial cell of Cajal in the pathogenesis of gastrointestinal diseases? *Adv Anat Pathol* 1999;6:19–40.
- Miettinen M, Monihan JM, Sarlomo-Rikala M, Kovatich AJ, Carr NJ, Emory TS, et al. Gastrointestinal stromal tumours/smooth muscle tumours (GISTs) primary in the omentum and mesentery: clinicopathologic and immunohistochemical study of 26 cases. *Am J Surg Pathol* 1999;23:1109–18.
- Wang L, Vargas H, French SW. Cellular origin of gastrointestinal stromal tumours. *Arch Pathol Lab Med* 2000;124:1471–5.
- Heinrich M, Rubin B, Longley B, Fletcher J. Biology and genetic aspects of gastrointestinal stromal tumours: KIT activation and cytogenetic alterations. *Hum Pathol* 2002;33:484–95.
- Heinrich M, Corless C, Demetri G, Blanke C, Von Mehren M, Joensuu H, et al. Kinase mutations and imatinib response in patients with metastatic gastrointestinal stromal tumour. *J Clin Oncol* 2003;21:4342–9.
- Maeda H, Yamagata A, Nishikawa S, Yoshinaga K, Kobayashi S, Nishi K, et al. Requirement of c-kit for development of intestinal pacemaker system. *Development* 1992;116:369–75.
- Hirota S, Nishida T, Isozaki K, Taniguchi M, Nakamura J, Okazaki T, et al. Gain-of-function mutation at the extracellular domain of KIT in gastrointestinal stromal tumours. *J Pathol* 2001;193:505–10.
- Silva M, Reid R. Gastrointestinal stromal tumours (GIST): C-kit mutations, CD117 expression, differential diagnosis and targeted cancer therapy with imatinib. *Pathol Oncol Res* 2003;9:13–9.

- 32 Hirota S, Isozaki K, Moriyama Y, Hashimoto K, Nishida T, Ishiguro S, et al. Gain-of-function mutations of c-kit in human gastrointestinal stromal tumours. *Science* 1998;279:577–80.
- 33 Kitamura Y, Hirota S, Nishida T. Molecular pathology of c-kit proto-oncogene and development of gastrointestinal stromal tumours. *Ann Chir Gynaecol* 1998;86:282–6.
- 34 Isozaki K, Terris B, Belghiti J, Schiffmann S, Hirota S, Vanderwinden J. Germline-activating mutation in the kinase domain of KIT gene in familial gastrointestinal stromal tumours. *Am J Pathol* 2000;157:1581–5.
- 35 Kitamura Y, Hirota S, Nishida T. Gastrointestinal stromal tumours (GIST): A model for molecule-based and treatment of solid tumours. *Cancer Sci* 2003;94:315–20.
- 36 Ernst SI, Hubbs AE, Przygodzki RM, Emory TS, Sobin LH, O'Leary TJ. KIT mutation portends poor prognosis in gastrointestinal stromal/smooth muscle tumours. *Lab Invest* 1998;78:1633–6.
- 37 Dematteo RP, Heinrich MC, El-Rifai WM, Demetri G. Clinical management of gastrointestinal stromal tumours: before and after STI-571. *Hum Pathol* 2002;33:466–77.
- 38 Heinrich M, Corless C, Duensing A, McGreevey L, Chen C, Joseph N, et al. PDGFRA activating mutations in gastrointestinal stromal tumours. *Science* 2003;299:708–10.
- 39 Franquemont DW. Differentiation and risk assessment of gastrointestinal stromal tumours. *Am J Clin Pathol* 1995;103:41–7.
- 40 Emory TS, O'Leary TJ. Prognosis and surveillance of gastrointestinal stromal/smooth muscle tumours. *Ann Chir Gynaecol* 1998;87:306–10.
- 41 Appelman HD. Smooth muscle tumours of the gastrointestinal tract. What we know now that Stout didn't know. *Am J Surg Pathol* 1986;10(Suppl 1):83–99.
- 42 Golden T, Stout AP. Smooth muscle tumours of the gastrointestinal tract and retroperitoneal tissues. *Surg Gynecol Obstet* 1941;73:784–810.
- 43 Emory TS, Sobin LH, Lukes L, Lee DH, O'Leary TJ. Prognosis of gastrointestinal smooth-muscle (stromal) tumours: dependence on anatomic site. *Am J Surg Pathol* 1999;23:82–7.
- 44 Wu P, Langerman B, Ryan C, Hart J, Swiger S, Posner M. Surgical treatment of gastrointestinal stromal tumours in the imatinib (STI-571) era. *Surgery* 2003;134:656–65.
- 45 Ma CK, Amin MB, Kintanar E, Linden MD, Zarbo RJ. Immunohistologic characterization of gastrointestinal stromal tumours: a study of 82 cases compared with 11 cases of leiomyomas. *Mod Pathol* 1993;6:139–44.
- 46 Tworek JA, Appelman HD, Singleton TP, Greenson JK. Stromal tumours of the jejunum and ileum. *Mod Pathol* 1997;10:200–9.
- 47 Pierie JP, Choudry U, Muzikansky A, Yeap BY, Souba WW, Ott MJ. The effect of surgery and grade on outcome of gastrointestinal stromal tumours. *Arch Surg* 2001;136:383–9.
- 48 Brainard JA, Goldblum JR. Stromal tumours of the jejunum and ileum: a clinicopathologic study of 39 cases. *Am J Surg Pathol* 1997;21:407–16.
- 4 Muro-Cacho CA, Cantor AB, Morgan M. Prognostic factors in malignant gastrointestinal stromal tumours. *Ann Clin Lab Sci* 2000;30:239–47.
- 50 Tazawa K, Tsukada K, Makuuchi H, Tsutsumi Y. An immunohistochemical and clinicopathological study of gastrointestinal stromal tumours. *Pathol Int* 1999;49:786–98.
- 51 Rudolf P, Gloeckner K, Parwaresch R, Harms D, Schmidt D. Immunophenotype, proliferation, DNA-ploidy, and biological behavior of gastrointestinal stromal tumours: a multivariate clinicopathologic study. *Hum Pathol* 1998;29:791–800.
- 52 Emory T, Derringer G, Sobin L, O'Leary T. Ki-67 (Mib1) immunohistochemistry as a prognostic factor in gastrointestinal smooth-muscle tumours. *J Surg Pathol* 1997;2:239–42.
- 53 Ma CK, De Peralta MN, Amin MB, Linden MD, Dekovich AA, Kubus JJ, et al. Small intestinal stromal tumours: a clinicopathologic study of 20 cases with immunohistochemical assessment of cell differentiation and the prognostic role of proliferation antigens. *Am J Clin Pathol* 1997;108:641–51.
- 54 Yan H, Marchettini P, Acherman YI, Gething SA, Brun E, Sugarbaker PH. Prognostic assessment of gastrointestinal stromal tumour. *Am J Clin Oncol* 2003;26:221–8.
- 55 Franquemont DW, Frierson HF Jr. Proliferating cell nuclear antigen immunoreactivity and prognosis of gastrointestinal stromal tumours. *Mod Pathol* 1995;8:473–7.
- 56 Miettinen M, El-Rifai W, L HLS, Lasota J. Evaluation of malignancy and prognosis of gastrointestinal stromal tumours: a review. *Hum Pathol* 2002;33:478–83.
- 57 Gall JA, Chetty R, Kemp AJ, Penfold JC. Multiple benign stromal cell tumours of the small bowel. *J Clin Pathol* 1993;46:869–71.
- 58 Bucher P, Taylor S, Villiger P, Morel P, Brundler M. For small intestinal stromal tumours are there any prognostic factors. *Am J Surg* 2004, in press.
- 59 Langer C, Gunawan B, Schuler P, Huber W, Fuzesi L, Becker H. Prognostic factors influencing surgical management and outcome of gastrointestinal stromal tumours. *Br J Surg* 2003;90:332–9.
- 60 Roberts P. GIST. Gastrointestinal stromal tumours. *Ann Chir Gynaecol* 1998;87:277.
- 61 Roberts P, Eisenberg B. Clinical presentation of gastrointestinal stromal tumours and treatment of operable disease. *Eur J Cancer* 2002;38:S37–8.
- 62 Verreer PR, Clausen TA, Schoepp C. Principles of surgical management of stromal tumour. *Chirurg* 2000;71:1335–44.
- 63 Pithorecky I, Cheney RT, Kraybill WG, Gibbs JF. Gastrointestinal stromal tumours: current diagnosis, biologic behavior, and management. *Ann Surg Oncol* 2000;7:705–12.
- 64 Lehnert T. Gastrointestinal sarcoma (GIST) – a review of surgical management. *Ann Chir Gynaecol* 1998;87:297–305.
- 65 Crosby JA, Catton CN, Davis A, Couture J, O'Sullivan B, Kandel R, et al. Malignant gastrointestinal stromal tumours of the small intestine: a review of 50 cases from a prospective database. *Ann Surg Oncol* 2001;8:50–9.
- 66 He LJ, Wang BS, Chen CC. Smooth muscle tumours of the digestive tract: report of 160 cases. *Br J Surg* 1988;75:184–6.
- 67 Pross M, Manger T, Schulz HU, Lippert H, Roessner A, Gunther T. Gastrointestinal stromal tumours—problems in diagnosis and therapy. *Chirurg* 1999;70:807–12.
- 68 Clary B, Dematteo M, Lewis J, Leung D, Brennan M. Gastrointestinal stromal tumours and leiomyosarcoma of the abdomen and retroperitoneum: A clinical comparison. *Ann Surg Oncol* 2001;8:290–9.
- 69 Lehnert T, Schwarzbach M, Willeke F, Herfarth C. Gastrointestinal stroma tumours – a special entity with unique radicality principles. *Langenbecks Arch Chir Suppl Kongressbd* 1998;115:356–8.
- 70 Levitzki A. Tyrosine kinases as targets for cancer therapy. *Eur J Cancer* 2002;38:S11–S18.
- 71 Manley P, Cowan-Jacob S, Buchdunger E, Fabbro D, Fendrich G, Furet P, et al. Imatinib: a selective tyrosine kinase inhibitor. *Eur J Cancer* 2002;38:S19–S27.
- 72 Capdeville R, Silbermann S, Dimitrijevic S. Imatinib: the first 3 years. *Eur J Cancer* 2002;38:S77–S82.
- 73 Pawson T. Regulation and targets of receptor tyrosine kinase. *Eur J Cancer* 2002;38:S3–S10.
- 74 Druker BJ, Tamura S, Buchdunger E, Ohno S, Segal GM, Fanning S, et al. Effects of a selective inhibitor of the Abl tyrosine kinase on the growth of Bcr-Abl positive cells. *Nat Med* 1996;2:561–6.
- 75 Buchdunger E, O'Reilly T, Wood J. Pharmacology of imatinib (STI571). *Eur J Cancer* 2002;38:S28–S36.
- 76 Buchdunger E, Cioffi CL, Law N, Stover D, Ohno-Jones S, Druker BJ, et al. Abl protein-tyrosine kinase inhibitor STI571 inhibits in vitro signal transduction mediated by c-kit and platelet-derived growth factor receptors. *J Pharmacol Exp Ther* 2000;295:139–45.
- 77 Bummig P, Andersson J, Meis-Kindblom J, Klingenstein H, Engstrom K, Stierner U, et al. Neoadjuvant, adjuvant and palliative treatment of gastrointestinal stromal tumours (GIST) with imatinib: a centre-based study of 17 patients. *Br J Cancer* 2003;89:460–4.
- 78 Bauer S, Corless C, Heinrich M, Dirsch O, Antoch G, Kanja J, et al. Response to imatinib mesylate of gastrointestinal stromal tumour with very low expression of KIT. *Cancer Chemother Pharmacol* 2003;51:261–5.
- 79 Joensuu H, Roberts PJ, Sarlomo-Rikala M, Andersson LC, Terahartala P, Tuveson D, et al. Effect of the tyrosine kinase inhibitor STI571 in a patient with a metastatic gastrointestinal stromal tumour. *N Engl J Med* 2001;344:1052–6.
- 80 Eisenberg B, Von Mehren M. Pharmacotherapy of gastrointestinal stromal tumours. *Expert Opin Pharmacother* 2003;4:869–74.
- 81 Connolly E, Gaffney E, Reynolds J. Gastrointestinal stromal tumours. *Br J Surg* 2003;90:1178–86.
- 82 Schwartz R. A molecular star in the war against cancer. *N Engl J Med* 2002;347:462–3.
- 83 Silbermann S, Joensuu H. Overview of issues related to imatinib therapy of advanced gastrointestinal stromal tumours: a discussion among the experts. *Eur J Cancer* 2002;38:S66–S69.
- 84 DeMatteo R. The GIST targeted cancer therapy: a tumour (gastrointestinal stromal tumour), a mutated gene (c-kit), and a molecular inhibitor (STI571). *Am Surg Oncol* 2002;9:831–9.

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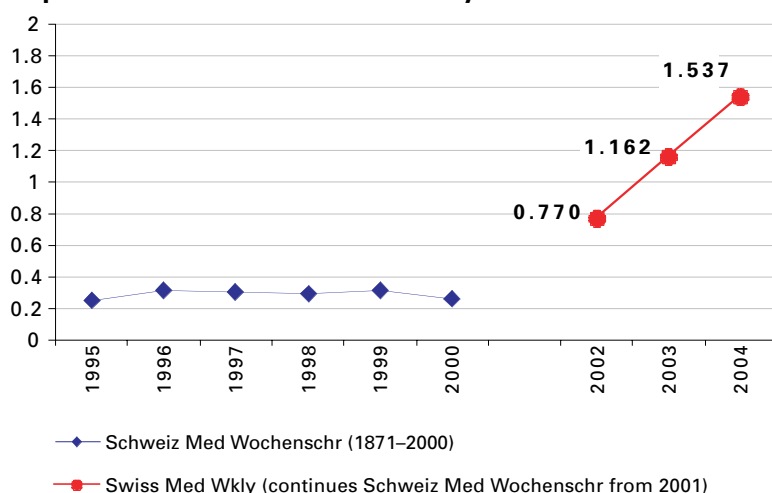
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