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The H90-10 single-chain antibody recognizes Hsp90 β by immunoprecipitation and Western blotting

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Abstract

The recombinant antibody H90-10 detects the endogenous human heat-shock protein 90 beta (Hsp90 β) by immunoprecipitation (IP) and Western blotting.

Introduction

The mouse monoclonal antibody H90-10 specifically recognizes the Hsp90 β (UniProt #P08238) isoform of the Hsp90 family (Holt *et al.*, 1999; Barent *et al.*, 1998). Here, we describe the ability of the single-chain variable antibody (scFv) H90-10 to immunoprecipitate the endogenous human Hsp90 β and to recognize it by immunoblotting.

Materials & Methods

Antibodies: The ABCD_AO870 antibody (ABCD nomenclature, <https://web.expasy.org/abcd>) was produced by the Geneva Antibody Facility (<https://www.unige.ch/medecine/antibodies>) as a mini-antibody with the antigen-binding scFv fused to the Fc region of mouse IgG2a. The synthesized scFv sequence (GeneArt, Invitrogen) corresponds to the sequence of the variable regions of the anti-Hsp90 β monoclonal H90-10 (Holt *et al.*, 1999; Barent *et al.*, 1998) joined by a peptide linker (GGGGS)₃. The H90-10 variable sequences were determined with permission by Brian C. Freeman (University of Illinois, Urbana) from the H90-10 hybridoma originally from David O. Toft (Mayo Clinic, Rochester). HEK293 suspension cells (growing in FreeStyle™ 293 Expression Medium, Gibco #12338) were transiently transfected with the vector coding for the scFv-Fc. Supernatant (~90 mg/L) was collected after 5 days.

Antigen: Both wild-type HEK293T cells, which endogenously express Hsp90 β , and their Hsp90 β knock-out counterpart (Bhattacharya *et al.*, 2020) were grown in Dulbecco's Modified Eagle's Medium supplemented with GlutaMAX, 10% fetal bovine serum, and penicillin/streptomycin (100 U/ml).

Protocol: Cells were pelleted and lysed in lysis buffer (10 mM Tris-HCl pH 7.5, 1 mM EDTA, 10 mM NaCl, 10 mM Na-molybdate, 10% glycerol, 1 mM DTT, 0.1% Triton X-100) with 1x protease inhibitor complex for 50 min at 4 °C using a Bioruptur™ Twin sonicator. Extracts were centrifuged at 16'100 g for 10 min at 4 °C and the pellets were discarded. 500 μ g of proteins were used for the IP. Samples were incubated overnight with murine IgG (Sigma-Aldrich #I5381) as negative control, H90-10, or scFv H90-10, diluted at 1:250, 1:50, and 1:50, respectively. The next day, 50 μ l of Dynabeads™ Protein

G (Invitrogen #10009D) were added for three hours and then washed 6 times for 10 min with lysis buffer. 50 μ g extract (Fig. 1), and 20 μ g input extract and IP samples (Fig. 2) were loaded on separate 10% SDS-polyacrylamide gel and then transferred to a nitrocellulose membrane (100 V, 105 min). The membranes were blocked for 60 min with 5% w/v non-fat dry milk in Tris-buffered saline containing 0.2% Tween 20 (TBST), then incubated overnight at 4 °C with the different antibody dilutions in TBST. As a loading control, corresponding sections of the same membranes were probed with an anti-GAPDH antibody (Hytest, #5G4, dilution 1:5'000). After washing the membranes three times for 15 min with TBST, they were incubated for 90 min at room temperature with horseradish peroxidase-coupled goat anti-mouse antibody (Invitrogen #31430, dilution 1:10'000 in TBST) and washed again three times for 15 min. The immunoblot of the IP experiment was probed similarly as indicated. Chemiluminescent signals were recorded with a LI-COR Odyssey Fc Imaging System.

Results

The scFv version of H90-10 specifically recognizes Hsp90 β (Fig. 1), and it immunoprecipitates Hsp90 β as well as or better than the original monoclonal H90-10 (Fig. 2).

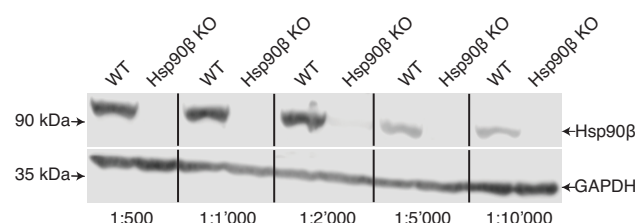


Fig. 1. Immunoblot showing specific recognition of Hsp90 β by the scFv H90-10 antibody at different dilutions. Extracts of Hsp90 β KO cells were used as the negative control and GAPDH as the loading control.

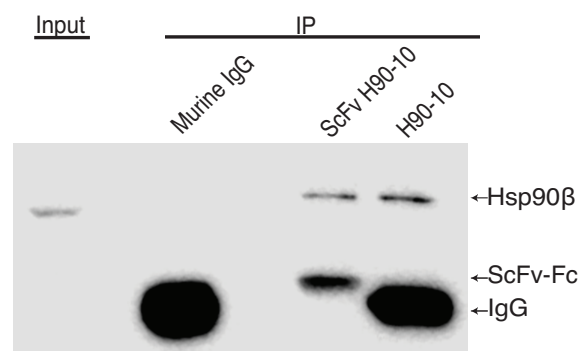


Fig. 2. Immunoblot of an IP experiment comparing the original monoclonal H90-10 and the scFv version, with normal murine IgG as negative control, probed with scFv H90-10 (dilution 1:1'000).

References

Barent RL, Nair SC, Carr DC, *et al.* Analysis of FKBP51/FKBP52 chimeras and mutants for Hsp90 binding and association with progesterone receptor complexes. *Mol Endocrinol.* 1998; 12(3):342–54. PMID:9514152

Bhattacharya K, Weidenauer L, Luengo TM, *et al.* The Hsp70-Hsp90 co-chaperone Hop/Stip1 shifts the proteostatic balance from folding towards degradation. *Nat Commun.* 2020; 11(1):5975. PMID:33239621

Holt SE, Aisner DL, Baur J, *et al.* Functional requirement of p23 and Hsp90 in telomerase complexes. *Genes Dev.* 1999; 13(7):817–26. PMID:10197982

Acknowledgments

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Conflict of interest

The authors declare no conflict of interest.