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Deoxynojirimycin analogs : 1,5-dideoxy-1,5-(*N*-hydroxyimino)-D-lyxitol derivatives

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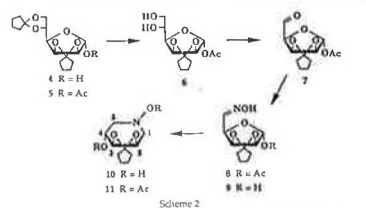
References

1. H. Schöf, *Liebigs Ann. Chem.* 1869, 151, 166.
2. A. Euenne and B. Bonic, *Bull. Soc. Chim. Fr.*, 1975, 1419.
3. A. Orszegovich and G. Orszegovich, *Gazz. Chem. Ital.*, 1936, 66, 48; 1938, 68, 688.
4. C. Ziegner, E. A. Cardella and G. Back, *Monatsh. Chem.* 1961, 92, 31.
5. Y. Ogata, A. Kawasaki and N. Okumura, *J. Org. Chem.*, 1965, 30, 1636; *Tetrahedron*, 1966, 22, 1731.
6. A. R. Butler and E. Leitch, *J. Chem. Soc., Perkin Trans. 2*, 1976, 832.
7. A. R. Butler and E. Leitch, *J. Chem. Soc., Perkin Trans. 2*, 1980, 103.
8. A. R. Butler, I. Hussain and E. Leitch, *J. Chem. Soc., Perkin Trans. 2*, 1980, 106.
9. A. R. Butler and I. Hussain, *J. Chem. Soc., Perkin Trans. 2*, 1981, 310.
10. A. R. Butler, I. Hussain and K. M. Preti, *J. Chem. Soc., Perkin Trans. 2*, 1981, 320.
11. A. R. Butler and I. Hussain, *J. Chem. Soc., Perkin Trans. 2*, 1980, 232.
12. A. R. Butler and I. Hussain, *J. Chem. Soc., Perkin Trans. 2*, 1980, 232.
13. F. Kurtz, *Chem. Reviews*, 1956, 95.
14. A. E. A. Wenzel, *J. Chem. Soc.*, 1913, 103, 1010.
15. C. J. Brian and A. R. Butler, *J. Chem. Soc., Perkin Trans. 2*, 1991, 1501.
16. C. J. Brian and A. R. Butler, *J. Chem. Soc., Perkin Trans. 2*, 1992, 23.
17. O. Hoch, *Chem. Ber.*, 1992, 125, 749.
18. E. Bamberger, *Chem. Ber.*, 1880, 16, 1459.

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Compound 4 was quantitatively acetylated to 5 which was selectively mono-de-O-cyclopentylidenated in high yield to 6. Periodic oxidation of 6 led to 7 which was not isolated but directly converted to the oxime 8 which, in chloroform solution, existed as a 1:1 mixture of *E* and *Z* isomers. De-O-acetylation of 8 yielded 9 (*Z*:*E* mixture in chloroform). The assignment of the *E* or *Z* configuration to the diastereoisomers of 8 and 9 was, as usual, based on $^1\text{H-NMR}$ data,^{5,9} mainly the $\delta_{\text{H-5}}$, $\delta_{\text{H-4}}$ and $I_{\text{H-5}}$ values. Reduction of 9 using sodium cyanoborohydride in acidic conditions led to moderate yields of 10 which was di-O-acetylated to 11 (Scheme 2).



The *N*-(*E*)-phenylprop-2-enyl derivative of glucosylamine having shown interesting anti-HIV activity,¹⁰ we planned to attach a (*E*)-phenylprop-2-enyl group to the nitrogen atom of trihydroxypiperidines, analogs of deoxyglucosylamine. The phthalimide derivative 12 and the *O*-substituted hydroxylamine 13 were prepared using classical methods and the oxime 14, was obtained in good yield by reacting 7 with 13. Deacetylation of 14 gave 15. In chloroform, both 14 and 15 existed in ca 3:2 *E*/*Z* mixtures. Acidic cyanoborohydride reduction of 15 led to a mixture of 16 (98%) and 17 (5%). Compound 17 was quantitatively acetylated to 18 (Scheme 3). At room temperature, the $^1\text{H-NMR}$ spectra of the piperidine derivatives 10, 11, 16-18 were poorly resolved whereas good spectra were obtained either at 50-60 °C (time-averaged) or at -40 to -50 °C (two frozen species) (Tables 3 and 4). The ratio of the concentrations of the more (A) to the less abundant (B) forms and the values of the energy barriers obtained using the Gutowski's approximation¹¹ are collected in Table 5.

References: see frame 0281

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sion. The ΔG^\ddagger values measured are close (somewhat smaller) to those found for nitrogen inversion without chair interconversion in *N*-acetylornithine derivatives.¹²

The conformation of compounds 10-11 is more difficult to ascertain owing to the fact that the presence of a 2,3-O-cyclopentylidene group prohibits their existence as genuine chair forms. An X-ray diffraction study¹⁴ of the L-gulo analog of 10 (Scheme 5) showed this compound to exist in a $^4\text{C}_1$ flattened chair ($F_{1,2}$)¹⁵ form with the *N*-phenylpropenyl group in equatorial position. Interproton coupling constants of 11B (Table 4) indicate an equatorial position of the acetoxy group ($J_{\text{H-5,H-6}} = 10$ Hz) and a close to axial disposition of 2-OR ($J_{\text{H-1,H-2}} = 12$ Hz). All the couplings are in accordance with a flattened $^4\text{C}_1$ form ($F_{1,3}$). The major inversetomer 11A should exist in a flattened $^4\text{C}_1$ ($F_{1,3}$) form with an equatorial *N*-acetoxy group. The reason for which the conformer 11A bearing an axial 4-O-acetyl group is strongly favored over its equatorial counterpart 11B, is not well understood. Conversely, the major conformer of 10 corresponds to a $^4\text{C}_1$ form with both hydroxy groups in equatorial position as appears from its incomplete set of NMR data (Tables 3 and 4). A Monte Carlo conformational search was performed on 10 using the MacroModel 3.5 software¹⁶ in which were introduced the MM2 parameters developed¹⁷ from ab initio studies for the $\text{N}(\text{sp}^3)\text{-O}(\text{sp}^2)$ bond. The search was conducted using the chloroform solvation option and the solvent accessible surface area was analytically recomputed at each optimization step. Two hundred conformers were generated and minimized for each inversetomer. Starting from the (*N*) inversetomer, the most stable form found was a $^4\text{C}_1$ flattened chair ($F_{1,3}$) corresponding to 10A (same conformation as 11B) whereas from the (*N*) inversetomer a $^4\text{C}_1$ flattened chair ($F_{1,3}$) was obtained corresponding to 10B (same conformation as 11A). The two forms were almost isoenergetic (10A more stable than 10B by 0.6 kJ/mol). The coupling constants calculated for 10A using the Allou's equation¹⁸ included in the MacroModel software (Table 4) are in good accordance with the experimental values measured for 10B. The agreement is somewhat less satisfactory between computed 10B and 11A.

In any case, these observations indicate that in this series, the nitrogen inversion provokes a chair inversion whereas in a previously studied¹⁹ morpholine series,

References: see frames 0287 and 0288

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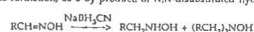
Deoxyglucosylamine Analogs: 1,5-Dideoxy-1,5-N-Hydroxyimino-*D*-Lysitol Derivatives

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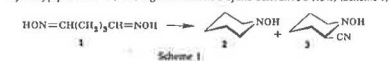
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Deoxy *N*-hydroxylamine sugars¹ ("sugar hydroxylamines") spontaneously oxidize in the air to give a very small stationary concentration of the corresponding aminoxy. This concentration is sufficient to allow the observation of good ESR spectra but too small to broaden significantly NMR signals. This corresponds to an almost unique situation where both good NMR and ESR spectra are obtainable from the same sample. We have described² a number of such sugar analogs in which a hydroxy group has been replaced with a *N*-hydroxylamine group. We report hereunder the extension of these syntheses to anhydrosorbitol analogs in which the ring oxygen has been replaced with a *N*-hydroxylamine bridge. Six-membered ring compounds of this type are analogs of deoxyglucosylamine. Part of these results have been the object of a preliminary communication.³

The sodium cyanoborohydride reduction of oximes into hydroxylamines is accompanied by the formation, as a by-product, of *N,N*-disubstituted hydroxylamines,⁴



the formation of which was decreased by using a large excess of reducing agent and by lowering the pH.⁵ Conversely, at higher pH and with a smaller excess of sodium borohydride, the formation of the *N,N*-disubstituted hydroxylamine should be favored and, starting with an α,α -dioxime, a ring closure could be expected affording a way to *N*-hydroxypiperidines and *N*-hydroxypiperidines. In fact, starting from 1,5-bis(*N*-hydroxylamino)pentane 1, we obtained the expected 1-hydroxypiperidine⁶ 2 (40%) together with its 2-cyano derivative 3 (15%) (Scheme 1).



This reaction, extended from *N,N*-bis(*N*-hydroxylamino) to α,α -*N*-hydroxylamine and α,α -*N*-acetoxyimino compounds, was applied to carbohydrate chemistry.

References: see frame 0287

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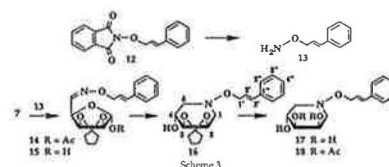


TABLE 3. $^1\text{H-NMR}$ of Piperidine Derivatives. Chemical Shifts of the Ring Protons

Compd	Solvent	$\delta_{\text{H-1}}$	$\delta_{\text{H-2}}$	$\delta_{\text{H-3}}$	$\delta_{\text{H-4}}$	$\delta_{\text{H-5}}$	$\delta_{\text{H-6}}$
10	CDCl_3 +60°	3.10	3.10	4.33	3.91	4.03	3.12
10A	CDCl_3 +40°	2.99	3.33	4.26	3.89	3.89	3.27
10B	CDCl_3 +40°	3.18	2.89	4.26	3.95	4.37	2.58
11	CDCl_3 +22°	3.31	3.20	4.40	4.00	5.30	3.25
11A	CDCl_3 -50°	3.45	3.02	4.46	3.98	5.34	3.12
11B	CDCl_3 -50°	2.97	3.83	4.31	4.07	5.12	3.53
16	CDCl_3 +60°	3.30	2.91	4.38	3.99	4.07	3.10
17	$\text{C}_6\text{D}_6\text{N} +110^\circ$	3.27	3.59	4.53	4.02	4.38	3.68
18	CDCl_3 +55°	3.1	3.30	5.37	5.04	5.21	3.9
18A	CDCl_3 +40°	2.71	4.7	5.39	4.85	5.26	3.72
18B	CDCl_3 -40°	3.41	2.77	5.19	5.12	4.96	2.79

TABLE 4. $^1\text{H-NMR}$ of Piperidine Derivatives. Interproton Couplings in the Ring

Compd	Solvent	$J_{\text{H-1,H-2}}$	$J_{\text{H-2,H-3}}$	$J_{\text{H-3,H-4}}$	$J_{\text{H-4,H-5}}$	$J_{\text{H-5,H-6}}$	$J_{\text{H-1,H-6}}$
10	CDCl_3 +60°	7	4.5	4.5	5.0	5.0	6.8
10A	CDCl_3 -40°	13.0	-7	-7	7	7	9.0
10B	CDCl_3 +40°	11.0	7	7	7	7	7
11	CDCl_3 +22°	12.0	5.5	6.0	5.0	5.0	6.0
11A	CDCl_3 -50°	11.5	5.5	7.0	4.8	3.8	3.09
11B	CDCl_3 -50°	13.0	2.07	3.07	5.5	7.0	4.0
16	CDCl_3 +60°	11.7	6.0	7.5	5.0	4.0	4.0
17	$\text{C}_6\text{D}_6\text{N} +110^\circ$	11.0	3.5	7.0	3.5	7.0	3.5
18	CDCl_3 +55°	11.5	3.0	6.5	3.5	7.5	3.5
18A	CDCl_3 +40°	11.5	1.0	2.0	3.5	10.5	4.5
18B	CDCl_3 -40°	11.0	4.0	11.5	2.5	2.0	2.0

10A calcd 2.6 2.9 5.1 7.1 5.3 10.7

10B calcd 7.0 9.5 4.7 2.9 2.3 3.0

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the energy difference between the two chairs was much larger so we could observe both inversetomers in the same chair conformation.

Upon spontaneous oxidation in the air, 10 gave the corresponding aminoxy radical 10', EPR spectrum of which was measured in diglyme at 50 °C (Fig. 1). Besides the hyperfine coupling with nitrogen, four unequal couplings with the $\text{H}_2\text{C-1}$ and $\text{H}_2\text{C-5}$ vicinal protons were observed, this excluding a conformation having a plane of symmetry perpendicular to the mean plane of the ring and passing through the nitrogen atom and C-3. A recent ab initio study of model aminoxy¹⁹ showed that the angular dependence of vicinal a_{H} couplings upon the 0 rotational angle between the C-H bond and an axis perpendicular to the C-N-C plane and passing through the nitrogen atom could be expressed by the classical equation:²⁰

$$a_{\text{H}}^2 = B_0 + B_2 \cos^2 \theta$$

only if using specific values of B_0 and B_2 depending on the out-of-plane deformation of the aminoxy group (a) and on whether the C-H bond is on the same side of the molecule as aminoxy oxygen or on the opposite side.¹⁹ On the other hand, the a_{H} value depends on the angle. In the case of 10', the experimental value of a_{H} (14.4 G) corresponds to an angle of about 15° which gives B_0 and B_2 values of ca 1.4 and 19.3 respectively.

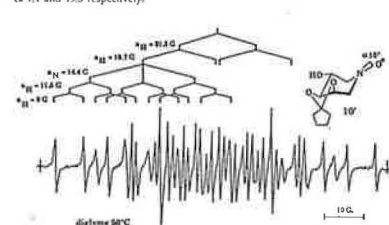


Figure 1. EPR spectrum of 10'.

References: see frame 0288

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Di-O-cyclopentylidenation of α -mannose led in high yield to the crystalline compound 4 obtained as the α anomer as shown by its $^1\text{H-NMR}$ values (Tables 1 and 2), particularly its small $J_{\text{H-5}}$ coupling constant.

TABLE 1. $^1\text{H-NMR}$ of the Furanose Derivatives. Chemical shifts values (6 in p.p.m.).

Compd	H-1	H-2	H-3	H-4	H-5	H-6
4	5.41	4.57	4.75	4.26	4.39	3.99
5	6.11	4.65	4.78	4.03	4.36	3.99
6	6.19	4.67	4.87	4.12	4.02	3.73
(E)-8	6.22	4.70	4.80	4.65	7.49	3.88
(Z)-8	6.20	4.68	5.07	5.13	6.90	
(E)-9	5.48	4.61	4.79	4.73	7.51	
(Z)-9	5.48	4.59	5.04	5.27	6.92	
(E)-14	6.22	4.77	4.77	4.66	7.50	
(Z)-14	6.22	4.77	5.05	5.12	6.90	
(E)-15	5.48	4.58	4.75	4.66	7.53	
(Z)-15	5.48	4.58	5.00	5.21	6.90	

TABLE 2. $^1\text{H-NMR}$ of the Furanose derivatives. Interproton couplings (*J* in Hz).

Compd	$J_{\text{H-1,H-2}}$	$J_{\text{H-2,H-3}}$	$J_{\text{H-3,H-4}}$	$J_{\text{H-4,H-5}}$	$J_{\text{H-5,H-6}}$
4	<0.5	6.0	4.0	7.0	7.0
5	<0.5	6.0	4.0	7.8	6.0
6	<0.5	6.2	4.0	8.0	5.0
(E)-8	<0.5	6.0	4.0	7.5	3.2
(Z)-8	<0.5	6.0	4.0	3.5	
(E)-9	<0.5	6.0	4.0	7.0	
(Z)-9	<0.5	6.0	4.0	4.0	
(E)-14	<0.5	6.0	4.0	7.0	
(Z)-14	<0.5	6.0	4.0	4.0	
(E)-15	<0.5	6.0	4.0	7.0	

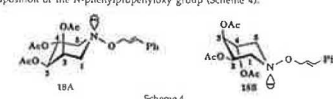
References: see frame 0287

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TABLE 5. Inversion Isomerism of Piperidine Derivatives.

Compd	ΔH (K)	ΔG^\ddagger (KJ/mol) (K)
10	1.8 (223)	52.9 (263)
11	4.5 (223)	52.3 (253)
16	1.3 (223)	54.4 (293)
17	1.0 (233)	57.7 (283)
18	2.7 (233)	55.1 (293)

As variable temperature $^1\text{H-NMR}$ (400 MHz) of 18 could be fully interpreted, it was chosen as a model compound for structure assignments. The interproton coupling values of 18A and 18B (Table 4) indicate a $^4\text{C}_1$ chair for A and a $^4\text{C}_1$ chair for B. A reliable way to establish the orientation (axial or equatorial) of the phenylpropenyl group borne by the nitrogen atom consists in the examination of the effect of vicinal groups²¹ on the absolute value of geminal coupling constants (i.e. $J_{\text{H-1,H-2}}$ and $J_{\text{H-5,H-6}}$). An axial lone pair decreases this value by 3-4 Hz whereas an axial electron-withdrawing (i.e. acetoxy) group increases it by 1-2 Hz. In the absence of either an axial lone pair or an electron-withdrawing group, the expected absolute value of these geminal coupling constants should amount to 14.0-14.5 Hz.¹⁹ For 18A, the very low value (9.5 Hz) of $J_{\text{H-1,H-2}}$ is explained by the presence of a vicinal axial lone pair and no axial acetoxy group whereas the intermediate value (11.5 Hz) of $J_{\text{H-5,H-6}}$ corresponds to the partial cancellation of the effect of the lone pair by that of an axial acetoxy group, thus indicating an equatorial disposition of the *N*-phenylpropenyl group (Scheme 4).

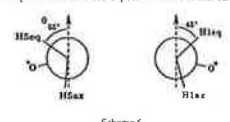


For 18B, $J_{\text{H-1,H-2}}$ and $J_{\text{H-5,H-6}}$ are both smaller than it would be expected for an axial *N*-phenylpropenyl group. $J_{\text{H-1,H-2}}$ being slightly increased by the vicinal axial acetoxy group. The most probable structure of 18B thus corresponds also to an equatorial *N*-phenylpropenyl group. In these conditions, 18A and 18B are inversetomers and the transformation observed by variable temperature NMR is a slow (at the NMR time scale) nitrogen inversion followed by a rapid chair inversion.

References: see frame 0287

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From this, the following assignments were made (Scheme 6) $\delta_{\text{H-1}}$ 8 G, calcd $\delta_{\text{H-1}}$ 7.8 G, $\delta_{\text{H-5}}$ 17.5° (exp. $\delta_{\text{H-5}}$ 21.3 G, calcd $\delta_{\text{H-5}}$ 20.6 G), $\delta_{\text{H-1,H-2}}$ 4.5° (exp. $\delta_{\text{H-1,H-2}}$ 11.1 G, calcd $\delta_{\text{H-1,H-2}}$ 11.4 G), $\delta_{\text{H-5,H-6}}$ 16.5° (exp. $\delta_{\text{H-5,H-6}}$ 19.7 G, calcd $\delta_{\text{H-5,H-6}}$ 19.4 G). These angular values correspond well to those expected from a flattened $^4\text{C}_1$ chair ($F_{1,3}$).



Compounds 10, 16, and 17 have been submitted to antibacterial and antiviral (including HIV-1 and HIV-2) testing following previously described procedures.²¹ Nonetheless, no notable activity, except a partial inhibition of the onco-virus SV₄₀ by 16 at a concentration of 105 μM , half the minimum cytotoxic concentration. No inhibition of either α -mannosidase or α -D-glucosidase was exhibited by compounds 16 and 17. Compound 17 constitutes an analog of *manno*-deoxyglucosylamine missing its terminal hydroxymethyl group. It is probable that the presence of this group is a prerequisite to anti-HIV activity.

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EXPERIMENTAL

General methods. See ref. 22. 400 MHz $^1\text{H-NMR}$ spectra were measured using a Bruker AMX400 spectrometer.

Sodium cyanoborohydride reduction of 1,5-bis(*N*-hydroxylamino)pentane (1). To a solution of 1 (6.5 g, 0.05 mol) in water (300 mL, 80 °C) NaOH/CN (6 g, 0.1 mol) was added and the pH kept at 4.5 by dropwise addition of aqueous 1 M HCl. After completion of the reaction (TLC), the pH was brought to 7 (saturated aqueous NaHCO₃) and the organic products were extracted (AcOEt, 3x100 mL). The collected organic fractions were dried (Na₂SO₄), concentrated and submitted to a column chromatography (4:1 AcOEt/hexane) yielding 2 (2 g, 40%) and 3 (0.9 g, 15%).

References: see

2-Cyano-1-hydroxyethylpyridine (1). Obtained as described hereupon: mp 96.7-100.6 °C; $\nu_{\text{max}}^{\text{IR}}$ 3335 (OH), 2946 and 2849 (CH), and 2518 (CN) cm^{-1} ; MS: m/z (%) 99 (100), 59 (70), 54 (64), 69 (54), 67 (47), 109 (47, M^{+} - OH), 53 (32), 126 (32, M^{+}), and 100 (25, M^{+} - CN); Anal. Calcd for $\text{C}_7\text{H}_7\text{N}_2\text{O}$ (126.16): C, 57.12; H, 7.99; N, 22.20. Found: C, 57.14; H, 7.94; N, 22.19.

2,3,5,6-Di-O-cyclopentylidene- α -D-mannofuranose (4). To a solution of D-mannose (15 g, 80.35 mmol) in freshly distilled cyclopentanone (400 mL), dried CuSO_4 (20 g, 94 mmol) and concentrated H_2SO_4 (1 mL) were added. After 36 h, the solution was filtered, neutralized (NaHCO_3 , 15 g, 0.18 mmol) for 1 h, then concentrated and extracted with ether (250 mL). The organic phase was washed with water (3x200 mL), dried (Na_2SO_4), then concentrated. Crystallization (heptane) gave 4 (21.48 g, 82.4%); mp 118.4 °C; R_f : 0.2 (1:4 AcOEt/hexane); $[\alpha]_D^{25}$ +16.3° (c 1.2); $\nu_{\text{max}}^{\text{IR}}$ 3420 (OH) cm^{-1} ; MS: m/z (%) 55 (100), 56 (22), 57 (24), 67 (13), 69 (19), 81 (12), 83 (11), 87 (27), 127 (16), and 212 (0.4, M^{+}).

Anal. Calcd for $\text{C}_{18}\text{H}_{28}\text{O}_{10}$ (312.37): C, 61.52; H, 7.74. Found: C, 61.49; H, 7.56.

1-O-Acetyl-2,3,5,6-di-O-cyclopentylidene- α -mannofuranose (5). A solution of 4 (21 g, 67.2 mmol) in a mixture of pyridine (30 mL) and Ac_2O (30 mL) was kept 12 h at room temperature, then extracted as usual. Column chromatography (1:2 AcOEt/hexane) gave 5 (23.5 g, 98.6%); syrup; R_f : 0.4 (1:4 AcOEt/hexane); $[\alpha]_D^{25}$ +27° (c 0.5); $\nu_{\text{max}}^{\text{IR}}$ 1750 (CO) cm^{-1} ; MS: m/z (%) 55 (100), 56 (25), 57 (16), 67 (11), 69 (23), 81 (12), 84 (19), 85 (24), 127 (52), and 354 (0.5, M^{+}).

Anal. Calcd for $\text{C}_{18}\text{H}_{26}\text{O}_{10}$ (334.40): C, 61.00; H, 7.39. Found: C, 60.88; H, 7.31.

1-O-Acetyl-2,3-O-cyclopentylidene- α -mannofuranose (6). A solution of 5 (12.62 g, 0.43 mmol) in MeOH (250 mL) and 1*N* HCl (5 mL) was kept 3 h (TLC) at room temperature then neutralized (saturated aqueous NaHCO_3), diluted with water (100 mL), and extracted with CHCl_3 (3x200 mL). The organic phase was dried (Na_2SO_4), then concentrated. Crystallization (toluene) gave 6 (13 g, 71%); mp 151.1-151.3 °C; R_f : 0.5 (6:1 AcOEt/hexane); $[\alpha]_D^{25}$ +52° (c 0.9); $\nu_{\text{max}}^{\text{IR}}$ 3400 and 3180 (OH), and 1746 (CO) cm^{-1} ; MS: m/z (%) 55 (94), 56 (28), 57 (24), 61 (17), 69 (20), 71 (18), 84 (25), 85 (100), 127 (26), and 288 (1, M^{+}).

Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_{10}$ (338.30): C, 54.16; H, 6.99. Found: C, 54.42; H, 7.06.

1-O-Acetyl-2,3-O-cyclopentylidene-5-deoxy-5-hydroxymethyl- α -lyxofuranose (8). To a solution of 6 (11.4 g, 39.5 mmol) in water (200 mL), NaIO_4 (8.45 g, 39.5 mmol) and NaHCO_3 (3.32 g, 39.5 mmol) were added. After 45 min at room temperature,

the mixture was extracted with CHCl_3 (5x100 mL), the organic phase dried (Na_2SO_4) and concentrated to give 7 (10 g) which was not further purified but dissolved in pyridine (100 mL), then hydroxylamine hydrochloride (8.14 g, 117.3 mmol) was added. After 12 h at room temperature, the solvent was evaporated with toluene (3x100 mL) and the residue extracted with AcOEt (200 mL). The organic phase was washed with water (5x100 mL), dried (Na_2SO_4) and concentrated. Column chromatography (1:1 AcOEt/hexane) gave 8 (5.54 g, 32% from 6); mp 130.7-130.8 °C; R_f : 0.4 and 0.5 (1:1 AcOEt/hexane); $\lambda_{\text{max}}^{\text{UV}}$ 204 nm (c 2400); $\nu_{\text{max}}^{\text{IR}}$ 3233 (OH) and 1754 (CO) cm^{-1} ; MS: m/z (%) 55 (100), 56 (24), 57 (13), 67 (13), 84 (11), 85 (27), 86 (16), 128 (22), 242 (13), and 271 (2, M^{+}).

Anal. Calcd for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}_6$ (271.27): C, 53.13; H, 6.32; N, 5.16. Found: C, 52.99; H, 6.18; N, 5.41.

2,3-O-Cyclopentylidene-5-deoxy-5-hydroxymethyl- α -lyxofuranose (9). A solution of 8 (4.78 g, 17.63 mmol) and NaOMe (925 mg, 17.6 mmol) in MeOH (75 mL) was kept at room temperature for 30 min, then concentrated and the residue extracted with EtOAc (100 mL). The organic phase was washed with water (3x50 mL), dried (Na_2SO_4), and concentrated. Column chromatography (1:1 AcOEt/hexane) gave 9 (2.5 g, 62%); mp 109.7-109.8 °C; R_f : 0.2 and 0.3 (1:1 AcOEt/hexane); $\lambda_{\text{max}}^{\text{UV}}$ 204 nm (c 3200); $\nu_{\text{max}}^{\text{IR}}$ 3370 (OH) cm^{-1} ; MS: m/z (%) 52 (16), 55 (100), 56 (29), 57 (19), 67 (19), 69 (11), 75 (11), 85 (20), 200 (23), and 229 (1, M^{+}).

Anal. Calcd for $\text{C}_{15}\text{H}_{18}\text{O}_6$ (229.23): C, 52.40; H, 6.60; N, 6.11. Found: C, 52.53; H, 6.66; N, 5.99.

2,3-O-Cyclopentylidene-1,5-dideoxy-1,5-*N*-(3'-phenylprop-2'-enyloxy)imino- α -lyxitol (10). To a solution of 9 (2.27 g, 9.9 mmol) in MeOH (250 mL), NaBH_4CN (0.13 g, 0.98 mmol) was added and the pH was kept at 2 by dropwise addition of 1 M HCl. After 30 min, the mixture was brought to pH 8 (saturated aqueous NaHCO_3), diluted with water (200 mL) then extracted with CHCl_3 (3x200 mL). The organic phases were dried (Na_2SO_4), then concentrated, and after column chromatography (2:1 AcOEt/hexane) gave 10 (300 mg, 38%); syrup; R_f : 0.15 (1:1 AcOEt/hexane); $[\alpha]_D^{25}$ -13.5° (c 0.8); $\lambda_{\text{max}}^{\text{UV}}$ 204 nm (c 570), 240 (200), and 290 (90); $\nu_{\text{max}}^{\text{IR}}$ 3400 (OH) cm^{-1} ; MS: m/z (%) 45 (70), 48 (26), 55 (57), 64 (28), 67 (25), 77 (36), 79 (28), 91 (100), 105 (23), and 215 (1, M^{+}).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{N}_2\text{O}_6$ (312.25): C, 55.80; H, 7.56; N, 6.51. Found: C, 55.66; H, 8.06; N, 6.51.

4-O-Acetyl-2,3-O-cyclopentylidene-1,5-dideoxy-1,5-*N*-(acetoxyimino)- α -lyxitol (11). To a solution of 10 (45 mg, 0.209 mmol) in Py (3 mL) Ac_2O (0.5 mL, 5 mmol) was added. After 5 h stirring at room temp, the solvent was evaporated with toluene (5x10 mL), and the residue, extracted with column chromatography (1:5 AcOEt/hexane), gave 11 (50 mg, 96%); syrup; R_f : 0.6 (1:5 AcOEt/hexane); $[\alpha]_D^{25}$ -37° (c 0.9); $\lambda_{\text{max}}^{\text{UV}}$ 204 nm (c 1023); $\nu_{\text{max}}^{\text{IR}}$ 2950 (CH sat.), 1750 and 1730 (C=O) cm^{-1} ; MS: m/z (%) 55 (86), 68 (53), 84 (58), 85 (46), 113 (61), 173 (100), 240 (27), and 299 (1.3, M^{+}).

Anal. Calcd for $\text{C}_{21}\text{H}_{28}\text{N}_2\text{O}_8$ (299.33): C, 56.18; H, 7.07; N, 4.68. Found: C, 55.93; H, 7.11; N, 4.60.

(*E*)-3-Phenylprop-2-enyloxyphthalimide (12). To a solution of (*E*)-3-phenylprop-2-enol (10 g, 74.5 mmol) in THF (500 mL), *N*-hydroxyphthalimide (13.24 g, 74.5 mmol), triphenylphosphine (21.5 g, 74.5 mmol) and then dropwise, diethyl azodicarboxylate (12.74 mL, 74.5 mmol) were added at 0 °C. After 4 days at room temperature, the reaction mixture was concentrated, and the residue purified by crystallization (EtOAc) gave 12 (16 g, 81%); mp 128.6-130.1 °C; $\lambda_{\text{max}}^{\text{UV}}$ 210 nm (c 45200), 216 (45500), and 241 (26300); $\nu_{\text{max}}^{\text{IR}}$ 1787, 1727 (CO), 1580, 1440, and 1453 (Ar) cm^{-1} ; ^1H NMR: δ 4.49 (dd, 2 H, $J_{1,2}$ 7 Hz, $J_{1,3}$ 1 Hz, $\text{H}_2\text{C}-1$), 6.48 (dt, 1 H, $J_{1,2}$ 16 Hz, $\text{H}_2\text{C}-2$), 6.70 (dt, 1 H, $\text{H}-3$), 7.39 (m, 5 H, Ar), and 7.80 (m, 4 H, Ar). MS: m/z (%) 117 (100), 91 (19), 147 (5), and 223 (0.5).

Anal. Calcd for $\text{C}_{19}\text{H}_{17}\text{NO}_2$ (279.30): C, 73.11; H, 4.69; N, 5.01. Found: C, 72.95; H, 4.65; N, 5.10.

(*E*)-3-Phenylprop-2-enylhydroxylamine (13). To a solution of 12 (16 g, 60.7 mmol) in EtOH (300 mL), hydrazine hydrate (8.8 mL, 182.1 mmol) was added. After 5 min stirring at room temperature, the reaction mixture filtered and concentrated was submitted to column chromatography (1:1 AcOEt/hexane) to give 13 (6.4 g, 79%); syrup; R_f : 0.5 (1:1 AcOEt/hexane); $\lambda_{\text{max}}^{\text{UV}}$ 205 nm (c 20000), and 250 (16400); $\nu_{\text{max}}^{\text{IR}}$ 3314, 3240 (NH), 1598, 1580, 1495, and 1450 (Ar) cm^{-1} ; ^1H NMR: δ 4.35 (dd, 2 H, $J_{1,2}$ 6 Hz, $J_{1,3}$ 1 Hz, $\text{H}_2\text{C}-1$), 4.45 (br, 2 H, OH), 6.35 (dt, 1 H, $J_{1,2}$ 16 Hz, $\text{H}_2\text{C}-2$), 6.68 (dt, 1 H, $\text{H}-3$), and 7.35 (m, 5 H, Ar). MS: m/z (%) 117 (100, M^{+} - NH_2), 91 (19), 77 (10), 51 (8), 103 (3), and 149 (2, M^{+}).

Anal. Calcd for $\text{C}_{17}\text{H}_{17}\text{NO}$ (149.19): C, 72.46; H, 7.43; N, 9.39. Found: C, 72.42; H, 7.58; N, 9.16.

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(*E* and *Z*)-1-O-Acetyl-2,3-O-cyclopentylidene-5-deoxy-5-(2',3'-phenylprop-2'-enyloxyimino)- α -lyxofuranose (14). To a solution of 7 (2.47 g, 9.61 mmol) in EtOH (100 mL), 13 (1.5 g, 10.05 mmol) was added and the mixture stirred for 12 h at room temperature. The reaction mixture, concentrated, was submitted to column chromatography (1:1 AcOEt/hexane) to give 14 (3 g, 80.6%); syrup; R_f : 0.71 (1:1 AcOEt/hexane); $[\alpha]_D^{25}$ -59° (c 0.6); $\lambda_{\text{max}}^{\text{UV}}$ 206 nm (c 17600) and 250 (12400); $\nu_{\text{max}}^{\text{IR}}$ 2950 (CH), and 1751 (CO) cm^{-1} ; ^1H NMR (2:3 EtOAc): δ 1.70 (m, 6 H, (E + Z) cyclopentylidene), 1.92 (m, 2 H, (E + Z) cyclopentylidene), 2.09 (s, 3 H, (E + Z) OAc), 4.66 (dd, 1 H, $J_{1,2}$ 4 Hz, $J_{1,3}$ 7.5 Hz, (Z) HC-4), 4.77 (m, (E + Z) $\text{H}_2\text{C}-1$), (E + Z) HC-2, 5.05 (dd, 1 H, $J_{1,2}$ 6 Hz, $J_{1,3}$ 4 Hz, (E) HC-3), 5.12 (t, 1 H, $J_{1,2}$ 4 Hz, (E) HC-4), 6.22 (s, 1 H, (E + Z) HC-1), 6.36 and 6.38 (2 dt, 1 H, $J_{1,2}$ 6.5 Hz, $J_{1,3}$ 6 Hz, (E + Z) HC-2), 6.64 (dd, 1 H, $J_{1,2}$ 1 Hz, (E + Z) HC-3), 6.90 (d, 1 H, (E) HC-5), 7.35 (m, 5 H, (E + Z) Ar), and 7.50 (d, 1 H, (Z) HC-5). MS: m/z (%) 117 (100), 55 (15), 91 (10), 74 (1), 325 (0.6), and 387 (0.2, M^{+}).

Anal. Calcd for $\text{C}_{27}\text{H}_{34}\text{N}_2\text{O}_{10}$ (587.44): C, 65.10; H, 6.50; N, 3.62. Found: C, 65.01; H, 6.52; N, 3.74.

(*E* and *Z*)-2,3-O-Cyclopentylidene-5-deoxy-5-(2',3'-phenylprop-2'-enyloxyimino)- α -lyxofuranose (15). To a solution of 14 (2.37 g, 6.11 mmol) in EtOH (70 mL), NaOMe (0.33 g, 6.11 mmol) was added. After 1 h at room temperature, the mixture (150 mL) was added and the reaction mixture extracted with CHCl_3 (3x100 mL). The collected organic phases, dried (Na_2SO_4) and concentrated, were submitted to column chromatography (1:1 AcOEt/hexane) to give 15 (2 g, 94.7%); mp 68-73.4 °C; R_f : 0.6 (1:1 AcOEt/hexane); $[\alpha]_D^{25}$ -61° (c 1); $\lambda_{\text{max}}^{\text{UV}}$ 205 nm (c 26300), 210 (25000), 216 (16000), and 250 (19000); $\nu_{\text{max}}^{\text{IR}}$ 3360 (OH), 1635 (C=N), 1690, 1670, 1490, and 1445 (C=C and Ar) cm^{-1} ; ^1H NMR (2:1 EtOH): δ 1.19 (m, 6 H, (E + Z) cyclopentylidene), 1.90 (m, 2 H, (E + Z) cyclopentylidene), 2.53 (d, 1 H, $J_{1,2}$ 2.5 Hz, (E) HC-1), 2.62 (d, 1 H, $J_{1,2}$ 2.5 Hz, (Z) HC-1), 2.58 (d, 1 H, $J_{1,2}$ 6 Hz, (E) HC-2), 2.56 (d, 1 H, $J_{1,2}$ 6 Hz, (Z) HC-2), 4.75 (m, 4 H, (E + Z) $\text{H}_2\text{C}-1$), (Z) HC-3, 5.00 (dd, 1 H, $J_{1,2}$ 4 Hz, (E) HC-3), 5.48 (d, 1 H, (E + Z) HC-1), 6.35 (dt, 1 H, $J_{1,2}$ 6 Hz, $J_{1,3}$ 16 Hz, (E + Z) HC-2), 6.66 (d, 1 H, (E + Z) HC-3), 6.90 (d, 1 H, (E) HC-5), 7.25 (m, 5 H, (E + Z) Ph), and 7.53 (d, 1 H, (Z) HC-5). MS: m/z (%) 117 (100), 55 (36), 91 (30), 77 (20), 199 (1), and 345 (0.2, M^{+}).

Anal. Calcd for $\text{C}_{27}\text{H}_{32}\text{N}_2\text{O}_8$ (545.40): C, 66.07; H, 6.71; N, 4.06. Found: C, 65.94; H, 6.72; N, 4.10.

(*E*)-2,3-O-Tri-O-acetyl-1,5-dideoxy-1,5-*N*-(3'-phenylprop-2'-enyloxyimino)- α -lyxitol (16). To a solution of 15 (2.31 g, 6.4 mmol) in EtOH (150 mL), NaBH_4CN (0.12 g, 19.1 mmol) was added at room temperature and the pH kept at 1 by dropwise addition of 1 M HCl. After 16 h, water (200 mL) was added and the reaction mixture extracted with CHCl_3 (4x100 mL). The organic phases, dried (Na_2SO_4) were concentrated and submitted to column chromatography to afford 16 (810 mg, 38.2%) and 17 (600 mg, 35.4%). Properties of 16: syrup; R_f : 0.52 (1:1 AcOEt/hexane); $[\alpha]_D^{25}$ +14.3° (c 0.9); $\lambda_{\text{max}}^{\text{UV}}$ 205 nm (c 35000), 210 (32000), 216 (20000), and 250 (28500); $\nu_{\text{max}}^{\text{IR}}$ 3450 (OH) cm^{-1} ; MS: m/z (%) 117 (100, phenylpropenyl), 55 (16), 91 (8), 130 (7), 77 (4), 68 (3), 214 (0.4), and 332 (0.1, M^{+} + H).

Anal. Calcd for $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_6$ (331.42): C, 68.86; H, 7.60; N, 4.23. Found: C, 68.73; H, 7.71; N, 4.19.

(*E*)-1,5-Dideoxy-1,5-*N*-(3'-phenylprop-2'-enyloxyimino)- α -lyxitol (17). Obtained as described for 16: mp 138.0-140.6 °C; R_f : 0.1 (9:1 AcOEt/hexane); $[\alpha]_D^{25}$ -5.4° (c 0.7); $\lambda_{\text{max}}^{\text{UV}}$ 205 nm (c 19000), 210 (17000), 216 (10000), and 250 (15000); $\nu_{\text{max}}^{\text{IR}}$ 3350 (OH) cm^{-1} ; MS: m/z (%) 117 (100, phenylpropenyl), 91 (10), 103 (5), 77 (8), 235 (1), 194 (0.9), 180 (0.5), 259 (0.5), 222 (0.4), and 249 (0.3).

Anal. Calcd for $\text{C}_{19}\text{H}_{24}\text{N}_2\text{O}_6$ (265.31): C, 63.38; H, 7.22; N, 5.28. Found: C, 63.24; H, 7.15; N, 5.38.

(*E*)-2,3,4-Tri-O-acetyl-1,5-dideoxy-1,5-*N*-(3'-phenylprop-2'-enyloxyimino)- α -lyxitol (18). To a solution of 17 (95 mg, 0.358 mmol) in pyridine (10 mL), acetic anhydride (1 mL, 10.6 mmol) was added and the mixture stirred for 12 h at room temperature. After codistillation of the pyridine with toluene (4x20 mL), the residue was submitted to a column chromatography (1:1 AcOEt/hexane) to give 18 (136 mg, 97%); syrup; R_f : 0.8 (1:1 AcOEt/hexane); $[\alpha]_D^{25}$ -19.9° (c 0.8); $\lambda_{\text{max}}^{\text{UV}}$ 206 nm (c 21900), and 252 (17900); $\nu_{\text{max}}^{\text{IR}}$ 1745 (CO) cm^{-1} ; ^{13}C NMR (100.6 MHz, CDCl_3) δ 55 (c): 169.61 (CO), 136.92 (C1'), 133.70 (C2'), 128.56 (C3'), 127.78 (C4'), 126.56 (C2'), 125.27 (C2'), 73.08 (C1'), 69.82 (C3'), 67.91 and 67.21 (C2 and C4), 56.01 and 55.68 (C1 and C5), 20.69, 20.66, and 20.53 (CH_3); -40 °C a 3:1 mixture of 18A and 18B: 18A, 170.80, 170.80, and 170.49 (1, 6, 3aCO), 135.60 (5, C1'), 134.66 (6, C3'), 128.41 (d, C3'), 127.89 (d, C4'), 126.31 (d, C2'), 122.99 (d, C2'), 73.12 (t, C1'), 70.77 (d, C2'), 66.35 (t, C2 and C4) 57.00 and 56.91 (2 d, C5 and C1), 21.20, 21.15 and 21.06 (3 q, 3aMe); 18B, 170.40, 170.17, and 169.84 (3 s, 3aCO), 135.90 (5, C1'), 134.40

(d, C3'), 128.41 (d, C3'), 127.89 (d, C4'), 126.31 (d, C2'), 122.28 (d, C2'), 72.98 (t, C1'), 67.88 (d, C4), 66.07 (d, C2), 65.25 (d, C3), 53.86 and 53.02 (2 d, C1 and C5), 21.20, 21.15, and 21.06 (3 q, 3aMe). MS: m/z (%) 117 (100, phenylpropenyl), 91 (10), 57 (9), 342 (9, M^{+} - AcO), and 220 (5).

Anal. Calcd for $\text{C}_{29}\text{H}_{34}\text{N}_2\text{O}_{10}$ (591.42): C, 61.37; H, 6.44; N, 3.58. Found: C, 61.33; H, 6.57; N, 3.45.

REFERENCES

- J. M. J. Tronchet, E. Winter-Mihaly, F. Habashi, D. Schwarzenbach, U. Liskic, and M. Geoffroy, *Helv. Chim. Acta*, 1982, 64, 610.
- J. M. J. Tronchet in *Bioactive Spin Labels*, R. L. Zhdanov Ed., Springer-Verlag Berlin, 1992, p. 355.
- J. M. J. Tronchet, G. Zosimo-Landolfo, M. Balkadjan, A. Ricca, M. Zedly, F. Barbalat-Rey, D. Cabrin, P. Lichle, and M. Geoffroy, *Tetrahedron Lett.*, 1991, 33, 4129.
- R.F. Borch, M. D. Bernstein, and H. D. Durst, *J. Am. Chem. Soc.*, 1971, 93, 2897.
- J. M. J. Tronchet, G. Zosimo-Landolfo, N. Bizzozero, D. Cabrin, F. Habashi, E. Jean, and M. Geoffroy, *J. Carbohydr. Chem.*, 1988, 7, 169.
- R. Wolfenstein, *Ber.*, 1892, 25, 1777; *ibid.*, 1893, 26, 2991.
- J. M. J. Tronchet, G. Zosimo-Landolfo, F. Villedon Denalde, M. Balkadjan, D. Cabrin, and F. Barbalat-Rey, *J. Carbohydr. Chem.*, 1990, 9, 823.
- J. M. J. Tronchet, F. Barbalat-Rey, and N. Le Hong, *Helv. Chim. Acta*, 1971, 54, 2615.
- G. J. Karabatsos and D. J. Fenoglio, *Topics Stereochemistry*, 1970, 5, 167.
- H. Shimizu, H. Tsuchie, H. Honma, K. Yoshida, T. Tsunokawa, H. Ushijima, and T. Kitamura, *Jpn. J. Med. Sci. Biol.*, 1990, 43, 75; H. Shimizu, H. Tsuchie, K. Yoshida, S. Morikawa, T. Tsunokawa, H. Yamamoto, H. Ushijima, and T. Kitamura, *AIDS*, 1990, 4, 925.
- A. Allerhand, H. S. Gutowski, J. Jonas, and R. A. Maitzner, *J. Am. Chem. Soc.*, 1966, 88, 3185.
- J. A. Pople and A. A. Boheer-Bij, *J. Chem. Phys.*, 1965, 42, 1339.
- J. M. J. Tronchet, M. Zedly, D. Cabrin, C. Jorand, F. Barbalat-Rey, I. Komaromi, A. Ricca, and M. Geoffroy, *Nucleosides & Nucleotides*, 1993, 12, 615.

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- J. M. J. Tronchet, M. Balkadjan, and G. Bernardinelli, unpublished results.
- J. M. J. Tronchet, F. Barbalat-Rey, and J.-M. Chale, *Carbohydr. Res.*, 1973, 30, 220.
- F. Mohamadi, N. G. Richards, W. C. Guida, R. Liskamp, M. Lipton, C. Caufield, G. Chang, T. Henrickson and W. C. Still, *J. Comp. Chem.*, 1990, 11, 440.
- J. M. J. Tronchet and I. Komaromi, *Int. J. Biol. Macromol.*, 1993, 15, 69.
- C. A. Haasnot, F. A. M. De Leduc, and C. Altona, *Tetrahedron*, 1980, 36, 2783.
- A. Ricca, J. M. J. Tronchet, J. Weber, and Y. Ellinger, *J. Phys. Chem.*, 1992, 96, 10779.
- J. Doudy, Y. Ellinger, A. Rastal, R. Subra, and G. Berthier, *Mol. Phys.*, 1967, 3, 217.
- J. M. J. Tronchet, M. Iznaola, F. Barbalat-Rey, H. Dhimane, A. Ricca, J. Balzarini, and E. De Clercq, *Eur. J. Med. Chem.*, 1992, 27, 55.
- J. M. J. Tronchet, M. Koufaki, G. Zosimo-Landolfo, and G. Bernardinelli, *J. Chem. Res.*, 1992, (M), 2901.

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